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<p>(21) International Application Number: PCT/US98/11692</p> <p>(22) International Filing Date: 5 June 1998 (05.06.98)</p> <p>(30) Priority Data: 60/048,771 6 June 1997 (06.06.97) US 60/049,443 12 June 1997 (12.06.97) US</p> <p>(71) Applicant (for all designated States except US): E.I. DU PONT DE NEMOURS AND COMPANY [US/US]; 1007 Market Street, Wilmington, DE 19898 (US).</p> <p>(72) Inventors; and</p> <p>(75) Inventors/Applicants (for US only): FALCO, Saverio, Carl [US/US]; 1902 Millers Road, Arden, DE 19810 (US). ALLEN, Stephen, M. [US/US]; 12 Stanton Avenue, West Chester, PA 19382 (US). RAFALSKI, J., Antoni [US/US]; 2028 Longcome Drive, Wilmington, DE 19810 (US). HITZ, William, D. [US/US]; 404 Hillside Road, Wilmington, DE 19807 (US). KINNEY, Anthony, John [US/US]; 609 Lore Avenue, Wilmington, DE 19809 (US). ABELL, Lynn, Marie [US/US]; 5 Laurel Court, Wilmington, DE 19808 (US). THORPE, Catherine, Jane [GB/GB]; 120 Ross Street, Cambridge CB1 3BU (GB).</p>		<p>(74) Agent: MAJARIAN, William, R.; E.I. du Pont de Nemours and Company, Legal Patent Records Center, 1007 Market Street, Wilmington, DE 19898 (US).</p> <p>(81) Designated States: AL, AM, AU, AZ, BA, BB, BG, BR, BY, CA, CN, CU, CZ, EE, GE, GW, HU, ID, IL, IS, JP, KG, KP, KR, KZ, LC, LK, LR, LT, LV, MD, MG, MK, MN, MX, NO, NZ, PL, RO, RU, SG, SI, SK, SL, TJ, TM, TR, TT, UA, US, UZ, VN, YU, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).</p> <p>Published Without international search report and to be republished upon receipt of that report.</p>	
<p>(54) Title: PLANT AMINO ACID BIOSYNTHETIC ENZYMES</p> <p></p> <p>(57) Abstract</p> <p>This invention relates to an isolated nucleic acid fragment encoding a plant enzyme that catalyzes steps in the biosynthesis of lysine, threonine, methionine, cysteine and isoleucine from aspartate, the enzyme a member selected from the group consisting of: dihydrodipicolinate reductase, diaminopimelate epimerase, threonine synthase, threonine deaminase and S-adenosylmethionine synthetase. The invention also relates to the construction of a chimeric gene encoding all or a portion of the enzyme, in sense or antisense orientation, wherein expression of the chimeric gene results in production of altered levels of the enzyme in a transformed host cell.</p>			

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TITLE

PLANT AMINO ACID BIOSYNTHETIC ENZYMES

This application claims the benefit of U.S. Provisional Application No. 60/048,771, filed June 6, 1997, and U.S. Provisional Application No. 60/049,443, filed June 12, 1997.

5 FIELD OF THE INVENTION

This invention is in the field of plant molecular biology. More specifically, this invention pertains to nucleic acid fragments encoding enzymes involved in amino acid biosynthesis in plants and seeds.

BACKGROUND OF THE INVENTION

10 Many vertebrates, including man, lack the ability to manufacture a number of amino acids and therefore require these amino acids preformed in the diet. These are called essential amino acids. Human food and animal feed, derived from many grains, are deficient in essential amino acids, such as lysine, the sulfur amino acids methionine and cysteine, threonine and tryptophan. For example, in corn (*Zea mays L.*) lysine is the most limiting 15 amino acid for the dietary requirements of many animals. Soybean (*Glycine max L.*) meal is used as an additive to corn-based animal feeds primarily as a lysine supplement. Thus, an increase in the lysine content of either corn or soybean would reduce or eliminate the need to supplement mixed grain feeds with lysine produced via fermentation of microbes. Furthermore, in corn the sulfur amino acids are the third most limiting amino acids, after 20 lysine and tryptophan, for the dietary requirements of many animals. The use of soybean meal, which is rich in lysine and tryptophan, to supplement corn in animal feed is limited by the low sulfur amino acid content of the legume. Thus, an increase in the sulfur amino acid content of either corn or soybean would improve the nutritional quality of the mixtures and reduce the need for further supplementation through addition of more expensive methionine.

25 Lysine, threonine, methionine, cysteine and isoleucine are amino acids derived from aspartate. Regulation of the biosynthesis of each member of this family is interconnected (see Figure 1). One approach to increasing the nutritional quality of human foods and animal feed is to increase the production and accumulation of specific free amino acids via genetic engineering of this biosynthetic pathway. Alteration of the activity of enzymes in this 30 pathway could lead to altered levels of lysine, threonine, methionine, cysteine and isoleucine. However, few of the genes encoding enzymes that regulate this pathway in plants, especially corn, soybeans and wheat, are available.

The organization of the pathway leading to biosynthesis of lysine, threonine, methionine, cysteine and isoleucine indicates that over-expression or reduction of expression 35 of genes encoding, *inter alia*, threonine synthase, dihydrodipicolinate reductase, diaminopimelate epimerase, threonine deaminase and S-adenosylmethionine synthetase in corn, soybean, wheat and other crop plants could be used to alter levels of these amino acids in human food and animal feed. Accordingly, availability of nucleic acid sequences

encoding all or a portion of these enzymes would facilitate development of nutritionally improved crop plants.

SUMMARY OF THE INVENTION

The instant invention relates to isolated nucleic acid fragments encoding plant enzymes involved in amino acid biosynthesis. Specifically, this invention concerns isolated nucleic acid fragments encoding the following plant enzymes that catalyze steps in the biosynthesis of lysine, threonine, methionine, cysteine and isoleucine from aspartate: dihydrodipicolinate reductase, diaminopimelate epimerase, threonine synthase, threonine deaminase and S-adenosylmethionine synthetase. In addition, this invention relates to nucleic acid fragments that are complementary to nucleic acid fragments encoding the listed plant biosynthetic enzymes.

In another embodiment, the instant invention relates to chimeric genes encoding the amino acid biosynthetic acid enzymes listed above or to chimeric genes that comprise nucleic acid fragments that are complementary to the nucleic acid fragments encoding the enzymes, operably linked to suitable regulatory sequences, wherein expression of the chimeric genes results in production of levels of the encoded enzymes in transformed host cells that are altered (i.e., increased or decreased) from the levels produced in untransformed host cells.

In a further embodiment, the instant invention concerns a transformed host cell comprising in its genome a chimeric gene encoding a plant amino acid biosynthetic enzyme operably linked to suitable regulatory sequences, the enzyme selected from the group consisting of: dihydrodipicolinate reductase, diaminopimelate epimerase, threonine synthase, threonine deaminase and S-adenosylmethionine synthetase. Expression of the chimeric gene results in production of altered levels of the biosynthetic enzyme in the transformed host cell. The transformed host cells can be of eukaryotic or prokaryotic origin, and include cells derived from higher plants and microorganisms. The invention also includes transformed plants that arise from transformed host cells of higher plants, and seeds derived from such transformed plants.

An additional embodiment of the instant invention concerns a method of altering the level of expression of a plant biosynthetic enzyme in a transformed host cell comprising: a) transforming a host cell with a chimeric gene comprising a nucleic acid fragment encoding a plant biosynthetic enzyme selected from the group consisting of dihydrodipicolinate reductase, diaminopimelate epimerase, threonine synthase, threonine deaminase and S-adenosylmethionine synthetase, operably linked to suitable regulatory sequences; and b) growing the transformed host cell under conditions that are suitable for expression of the chimeric gene wherein expression of the chimeric gene results in production of altered levels of the biosynthetic enzyme in the transformed host cell.

An additional embodiment of the instant invention concerns a method for obtaining a nucleic acid fragment encoding all or substantially all of an amino acid sequence encoding a

plant dihydrodipicolinate reductase, diaminopimelate epimerase, threonine synthase, threonine deaminase and S-adenosylmethionine synthetase.

A further embodiment of the instant invention is a method for evaluating at least one compound for its ability to inhibit the activity of a plant biosynthetic enzyme selected from the group consisting of dihydrodipicolinate reductase, diaminopimelate epimerase, threonine synthase, threonine deaminase and S-adenosylmethionine synthetase, the method comprising the steps of: (a) transforming a host cell with a chimeric gene comprising a nucleic acid fragment encoding a plant biosynthetic enzyme selected from the group consisting of dihydrodipicolinate reductase, diaminopimelate epimerase, threonine synthase, threonine deaminase and S-adenosylmethionine synthetase, operably linked to suitable regulatory sequences; (b) growing the transformed host cell under conditions that are suitable for expression of the chimeric gene wherein expression of the chimeric gene results in production of the biosynthetic enzyme in the transformed host cell; (c) optionally purifying the biosynthetic enzyme expressed by the transformed host cell; (d) treating the biosynthetic enzyme with a compound to be tested; and (e) comparing the activity of the biosynthetic enzyme that has been treated with a test compound to the activity of an untreated biosynthetic enzyme, thereby selecting compounds with potential for inhibitory activity.

BRIEF DESCRIPTION OF THE
DRAWINGS AND SEQUENCE DESCRIPTIONS

The invention can be more fully understood from the following detailed description and the accompanying drawings and sequence descriptions which form a part of this application.

Figure 1 depicts the biosynthetic pathway for the aspartate family of amino acids. The following abbreviations are used: AK = aspartokinase; ASADH = aspartic semialdehyde dehydrogenase; DHDPS = dihydrodipicolinate synthase; DHDPR = dihydrodipicolinate reductase; DAPEP = diaminopimelate epimerase; DAPDC = diaminopimelate decarboxylase; HDH = homoserine dehydrogenase; HK = homoserine kinase; TS = threonine synthase; TD = threonine deaminase; C γ S = cystathionine γ -synthase; C β L = cystathionine β -lyase; MS = methionine synthase; CS = cysteine synthase; and SAMS = S-adenosylmethionine synthase.

Figure 2 shows a multiple alignment of the amino acid sequence fragments reported herein encoding dihydrodipicolinate reductase (SEQ ID NOs:2 and 4) and the *Synechocystis* sp. dihydrodipicolinate reductase sequence set forth in DDBJ Accession No. D90899 (SEQ ID NO:5).

Figure 3 shows a multiple alignment of the amino acid sequence fragments reported herein encoding diaminopimelate epimerase (SEQ ID NOs:7, 9, 11, and 13) and the *Synechocystis* sp. diaminopimelate epimerase sequence set forth in DDBJ Accession No. D90917 (SEQ ID NO:14).

Figure 4 shows a multiple alignment of the amino acid sequence fragments reported herein encoding threonine synthase (SEQ ID NOs:16, 18, 20, 22, 24, and 26) and the *Arabidopsis thaliana* threonine synthase sequence set forth in GenBank Accession No. L41666 (SEQ ID NO:27).

5 Figure 5 shows a multiple alignment of the amino acid sequence fragments reported herein encoding threonine deaminase (SEQ ID NOs:29, 31, and 33) to the *Brukholderia capacia* threonine synthase set forth in GenBank Accession No. U40630 (SEQ ID NO:34).

Figure 6 shows the nucleotide sequence alignment of the S-adenosylmethionine synthetase reported herein for corn (SEQ ID NO:35) with the *Oryza sativa*

10 S-adenosylmethionine synthetase nucleotide sequence set forth in EMBL Accession No. Z26867 (SEQ ID NO:37).

Figure 7 shows the nucleotide sequence alignment of the S-adenosylmethionine synthetase reported here for soybean (SEQ ID NO:38) with the *Lycopersicon esculentum* S-adenosyl-methionine synthetase nucleotide sequence set forth in EMBL Accession

15 No. Z24741 (SEQ ID NO:40).

Figure 8 shows the nucleotide sequence alignment of the S-adenosylmethionine synthetase reported here for wheat (SEQ ID NO:41) with the *Hordeum vulgare* S-adenosylmethionine synthetase nucleotide sequence set forth in DDBJ Accession No. D63835 (SEQ ID NO:43).

20 Amino acid sequence alignments were performed using the Clustal method of alignment (Higgins, D.G. and Sharp, P.M. (1989) *CABIOS* 5:151-153), from the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI). Nucleotide sequence alignments were a result of the BLASTN search performed with each individual S-adenosylmethionine sequence.

25 The following sequence descriptions and sequence listings attached hereto comply with the rules governing nucleotide and/or amino acid sequence disclosures in patent applications as set forth in 37 C.F.R. §1.821-1.825.

SEQ ID NO:1 is the nucleotide sequence comprising the entire cDNA insert in clone csi1n.pk0042.a3 encoding a corn dihydrodipicolinate reductase.

30 SEQ ID NO:2 is the deduced amino acid sequence of a portion of a corn dihydrodipicolinate reductase derived from the nucleotide sequence of SEQ ID NO:1.

SEQ ID NO:3 is the nucleotide sequence comprising a portion of the cDNA insert in clone rls2.pk0017.d3 encoding a rice dihydrodipicolinate reductase.

35 SEQ ID NO:4 is the deduced amino acid sequence of a portion of a rice dihydrodipicolinate reductase derived from the nucleotide sequence of SEQ ID NO:3.

SEQ ID NO:5 is the amino acid sequence of the entire *Synechocystis sp.* dihydrodipicolinate reductase DDBJ Accession No. D90899.

SEQ ID NO:6 is the nucleotide sequence comprising the entire cDNA insert in clone chp2.pk0008.h4 encoding a corn diaminopimelate epimerase.

SEQ ID NO:7 is the deduced amino acid sequence of a portion of a corn diaminopimelate epimerase derived from the nucleotide sequence of SEQ ID NO:6.

SEQ ID NO:8 is the nucleotide sequence comprising a portion of the cDNA insert in clone rls48.pk0036.h10 encoding a rice diaminopimelate epimerase.

5 SEQ ID NO:9 is the deduced amino acid sequence of a portion of a rice diaminopimelate epimerase derived from the nucleotide sequence of SEQ ID NO:8.

SEQ ID NO:10 is the nucleotide sequence comprising a contig formed of portions of sfl1.pk0031.h3, and sgs1c.pk002.k12, and the entire cDNA insert from clones se2.pk0005.f1, and ses8w.pk0010.h11 encoding a soybean diaminopimelate epimerase.

10 SEQ ID NO:11 is the deduced amino acid sequence of a soybean diaminopimelate epimerase derived from the nucleotide sequence of SEQ ID NO:10.

SEQ ID NO:12 is the nucleotide sequence comprising a portion of the cDNA insert in clone wlm24.pk0030.g4 encoding a wheat diaminopimelate epimerase.

15 SEQ ID NO:13 is the deduced amino acid sequence of a portion of a wheat diaminopimelate epimerase derived from the nucleotide sequence of SEQ ID NO:12.

SEQ ID NO:14 is the nucleotide sequence comprising the entire *Synechocystis sp.* diaminopimelate epimerase DDBJ Accession No. D90917.

SEQ ID NO:15 is the nucleotide sequence comprising the entire cDNA insert in clone cc2.pk0031.c9 encoding a corn threonine synthase.

20 SEQ ID NO:16 is the deduced amino acid sequence of a portion of a corn threonine synthase derived from the nucleotide sequence set forth in SEQ ID NO:15.

SEQ ID NO:17 is the nucleotide sequence comprising part of the cDNA insert in clone cs1.pk0058.g5 encoding a corn threonine synthase.

25 SEQ ID NO:18 is the deduced amino acid sequence of a portion of a corn threonine synthase derived from the nucleotide sequence of SEQ ID NO:17.

SEQ ID NO:19 is the nucleotide sequence comprising part of the cDNA insert in clone rls72.pk0018.e7 encoding a rice threonine synthase.

SEQ ID NO:20 is deduced amino acid sequence of a portion of a rice threonine synthase derived from the nucleotide sequence set forth in SEQ ID NO:19.

30 SEQ ID NO:21 is the nucleotide sequence comprising part of the cDNA insert in clone se1.06a03 encoding a soybean threonine synthase.

SEQ ID NO:22 is the deduced amino acid sequence of a portion of a soybean threonine synthase derived from the nucleotide sequence set forth in SEQ ID NO:21.

35 SEQ ID NO:23 is the nucleotide sequence comprising the entire cDNA insert in clone sr1.pk0003.f6 encoding a soybean threonine synthase.

SEQ ID NO:24 is the deduced amino acid sequence of a portion of a soybean threonine synthase derived from the nucleotide sequence set forth in SEQ ID NO:23.

SEQ ID NO:25 is the nucleotide sequence comprising part of the cDNA insert in clone wr1.pk0085.h2 encoding a wheat threonine synthase.

SEQ ID NO:26 is the deduced amino acid sequence of a portion of a wheat threonine synthase derived from the nucleotide sequence set forth in SEQ ID NO:25.

SEQ ID NO:27 is the entire amino acid sequence of an *Arabidopsis thaliana* threonine synthase found in GenBank Accession No. L41666.

5 SEQ ID NO:28 is the nucleotide sequence comprising the entire cDNA insert in clone cen1.pk0064.f4 encoding a corn threonine deaminase.

SEQ ID NO:29 is the deduced amino acid sequence of a portion of a corn threonine deaminase derived from the nucleotide sequence set forth in SEQ ID NO:28.

10 SEQ ID NO:30 is the nucleotide sequence comprising a portion of the cDNA insert in clone sfl1.pk0055.h7 encoding a soybean threonine deaminase.

SEQ ID NO:31 is the deduced amino acid sequence of a portion of a soybean threonine deaminase derived from the nucleotide sequence set forth in SEQ ID NO:30.

SEQ ID NO:32 is the nucleotide sequence comprising the entire cDNA insert in clone stre.pk0044.f3 encoding a soybean threonine deaminase.

15 SEQ ID NO:33 is the deduced amino acid sequence of a portion of a soybean threonine deaminase derived from the nucleotide sequence set forth in SEQ ID NO:32.

SEQ ID NO:34 is the entire amino acid sequence of a *Burkholderia capacia* threonine deaminase found in GenBank Accession No. U49630.

20 SEQ ID NO:35 is the nucleotide sequence comprising the entire cDNA insert in clone cc3.mn0002.d2 encoding the entire corn S-adenosylmethionine synthetase.

SEQ ID NO:36 is the deduced amino acid sequence of a corn S-adenosylmethionine synthetase derived from the nucleotide sequence set forth in SEQ ID NO:35.

SEQ ID NO:37 is the entire nucleotide sequence of a *Oryza sativa* S-adenosyl-methionine synthetase found in EMBL Accession No. Z26867.

25 SEQ ID NO:38 is the nucleotide sequence of the entire cDNA insert in clone s2.12b06 encoding the entire soybean S-adenosyl-methionine synthetase.

SEQ ID NO:39 is the deduced amino acid sequence of the entire soybean S-adenosyl-methionine synthetase derived from the nucleotide sequence set forth in SEQ ID NO:38.

30 SEQ ID NO:40 is the entire nucleotide sequence of a *Lycopersicon esculentum* S-adenosyl-methionine synthetase found in EMBL Accession No. Z24741.

SEQ ID NO:41 is the nucleotide sequence comprising a contig formed of portions of the cDNA inserts in clones wre1.pk0002.c12, wre1n.pk0070.b8, wkm1c.pk0003.g4, wlk1.pk0028.d3, wre1n.pk170.d8, wre1.pk0086.d5, wre1.pk0103.h8, and wre1n.pk0082.b2 encoding a portion of a wheat S-adenosyl-methionine synthetase.

35 SEQ ID NO:42 is the deduced amino acid sequence of a wheat S-adenosyl-methionine synthetase derived from the nucleotide sequence set forth in SEQ ID NO:41.

SEQ ID NO:43 is the entire nucleotide sequence of a *Hordeum vulgare* S-adenosyl-methionine synthetase found in DDBJ Accession No. D63835.

The Sequence Descriptions contain the one letter code for nucleotide sequence characters and the three letter codes for amino acids as defined in conformity with the IUPAC-IYUB standards described in *Nucleic Acids Research* 13:3021-3030 (1985) and in the *Biochemical Journal* 219 (No. 2):345-373 (1984) which are herein incorporated by reference. The symbols and format used for nucleotide and amino acid sequence data comply with the rules set forth in 37 C.F.R. §1.822.

DETAILED DESCRIPTION OF THE INVENTION

In the context of this disclosure, a number of terms shall be utilized. As used herein, an "isolated nucleic acid fragment" is a polymer of RNA or DNA that is single- or double-stranded, optionally containing synthetic, non-natural or altered nucleotide bases. An isolated nucleic acid fragment in the form of a polymer of DNA may be comprised of one or more segments of cDNA, genomic DNA or synthetic DNA. As used herein, "contig" refers to an assemblage of overlapping nucleic acid sequences to form one contiguous nucleotide sequence. For example, several DNA sequences can be compared and aligned to identify common or overlapping regions. The individual sequences can then be assembled into a single contiguous nucleotide sequence.

As used herein, "substantially similar" refers to nucleic acid fragments wherein changes in one or more nucleotide bases results in substitution of one or more amino acids, but do not affect the functional properties of the protein encoded by the DNA sequence. "Substantially similar" also refers to nucleic acid fragments wherein changes in one or more nucleotide bases does not affect the ability of the nucleic acid fragment to mediate alteration of gene expression by antisense or co-suppression technology. "Substantially similar" also refers to modifications of the nucleic acid fragments of the instant invention such as deletion or insertion of one or more nucleotides that do not substantially affect the functional properties of the resulting transcript vis-à-vis the ability to mediate alteration of gene expression by antisense or co-suppression technology or alteration of the functional properties of the resulting protein molecule. It is therefore understood that the invention encompasses more than the specific exemplary sequences.

For example, it is well known in the art that antisense suppression and co-suppression of gene expression may be accomplished using nucleic acid fragments representing less than the entire coding region of a gene, and by nucleic acid fragments that do not share 100% identity with the gene to be suppressed. Moreover, alterations in a gene which result in the production of a chemically equivalent amino acid at a given site, but do not effect the functional properties of the encoded protein, are well known in the art. Thus, a codon for the amino acid alanine, a hydrophobic amino acid, may be substituted by a codon encoding another less hydrophobic residue, such as glycine, or a more hydrophobic residue, such as valine, leucine, or isoleucine. Similarly, changes which result in substitution of one negatively charged residue for another, such as aspartic acid for glutamic acid, or one positively charged residue for another, such as lysine for arginine, can also be expected to

produce a functionally equivalent product. Nucleotide changes which result in alteration of the N-terminal and C-terminal portions of the protein molecule would also not be expected to alter the activity of the protein. Each of the proposed modifications is well within the routine skill in the art, as is determination of retention of biological activity of the encoded products. Moreover, the skilled artisan recognizes that substantially similar sequences encompassed by this invention are also defined by their ability to hybridize, under stringent conditions (0.1X SSC, 0.1% SDS, 65°C), with the sequences exemplified herein. Preferred substantially similar nucleic acid fragments of the instant invention are those nucleic acid fragments whose DNA sequences are 80% identical to the DNA sequence of the nucleic acid fragments reported herein. More preferred nucleic acid fragments are 90% identical to the identical to the DNA sequence of the nucleic acid fragments reported herein. Most preferred are nucleic acid fragments that are 95% identical to the DNA sequence of the nucleic acid fragments reported herein. The Clustal multiple alignment algorithm (Higgins, D.G. and Sharp, P.M. (1989) CABIOS 5:151-153) was used here with a GAP PENALTY of 10 and a GAP LENGTH PENALTY of 10.

A "substantial portion" of an amino acid or nucleotide sequence comprises enough of the amino acid sequence of a polypeptide or the nucleotide sequence of a gene to afford putative identification of that polypeptide or gene, either by manual evaluation of the sequence by one skilled in the art, or by computer-automated sequence comparison and identification using algorithms such as BLAST (Basic Local Alignment Search Tool; Altschul, S. F., et al., (1993) J. Mol. Biol. 215:403-410; see also www.ncbi.nlm.nih.gov/BLAST/). In general, a sequence of ten or more contiguous amino acids or thirty or more nucleotides is necessary in order to putatively identify a polypeptide or nucleic acid sequence as homologous to a known protein or gene. Moreover, with respect to nucleotide sequences, gene specific oligonucleotide probes comprising 20-30 contiguous nucleotides may be used in sequence-dependent methods of gene identification (e.g., Southern hybridization) and isolation (e.g., *in situ* hybridization of bacterial colonies or bacteriophage plaques). In addition, short oligonucleotides of 12-15 bases may be used as amplification primers in PCR in order to obtain a particular nucleic acid fragment comprising the primers. Accordingly, a "substantial portion" of a nucleotide sequence comprises enough of the sequence to afford specific identification and/or isolation of a nucleic acid fragment comprising the sequence. The instant specification teaches partial or complete amino acid and nucleotide sequences encoding one or more particular plant proteins. The skilled artisan, having the benefit of the sequences as reported herein, may now use all or a substantial portion of the disclosed sequences for purposes known to those skilled in this art. Accordingly, the instant invention comprises the complete sequences as reported in the accompanying Sequence Listing, as well as substantial portions of those sequences as defined above.

“Codon degeneracy” refers to divergence in the genetic code permitting variation of the nucleotide sequence without effecting the amino acid sequence of an encoded polypeptide. Accordingly, the instant invention relates to any nucleic acid fragment that encodes all or a substantial portion of the amino acid sequence encoding the amino acid biosynthetic enzymes as set forth in SEQ ID NOs:2, 4, 7, 9, 11, 13, 16, 18, 20, 22, 24, 26, 29, 31, and 33. The skilled artisan is well aware of the “codon-bias” exhibited by a specific host cell in usage of nucleotide codons to specify a given amino acid. Therefore, when synthesizing a gene for improved expression in a host cell, it is desirable to design the gene such that its frequency of codon usage approaches the frequency of preferred codon usage of the host cell.

“Synthetic genes” can be assembled from oligonucleotide building blocks that are chemically synthesized using procedures known to those skilled in the art. These building blocks are ligated and annealed to form gene segments which are then enzymatically assembled to construct the entire gene. “Chemically synthesized”, as related to a sequence of DNA, means that the component nucleotides were assembled *in vitro*. Manual chemical synthesis of DNA may be accomplished using well established procedures, or automated chemical synthesis can be performed using one of a number of commercially available machines. Accordingly, the genes can be tailored for optimal gene expression based on optimization of nucleotide sequence to reflect the codon bias of the host cell. The skilled artisan appreciates the likelihood of successful gene expression if codon usage is biased towards those codons favored by the host. Determination of preferred codons can be based on a survey of genes derived from the host cell where sequence information is available.

“Gene” refers to a nucleic acid fragment that expresses a specific protein, including regulatory sequences preceding (5' non-coding sequences) and following (3' non-coding sequences) the coding sequence. “Native gene” refers to a gene as found in nature with its own regulatory sequences. “Chimeric gene” refers any gene that is not a native gene, comprising regulatory and coding sequences that are not found together in nature. Accordingly, a chimeric gene may comprise regulatory sequences and coding sequences that are derived from different sources, or regulatory sequences and coding sequences derived from the same source, but arranged in a manner different than that found in nature. “Endogenous gene” refers to a native gene in its natural location in the genome of an organism. A “foreign” gene refers to a gene not normally found in the host organism, but that is introduced into the host organism by gene transfer. Foreign genes can comprise native genes inserted into a non-native organism, or chimeric genes. A “transgene” is a gene that has been introduced into the genome by a transformation procedure.

“Coding sequence” refers to a DNA sequence that codes for a specific amino acid sequence. “Regulatory sequences” refer to nucleotide sequences located upstream (5' non-coding sequences), within, or downstream (3' non-coding sequences) of a coding sequence, and which influence the transcription, RNA processing or stability, or translation of the

associated coding sequence. Regulatory sequences may include promoters, translation leader sequences, introns, and polyadenylation recognition sequences.

“Promoter” refers to a DNA sequence capable of controlling the expression of a coding sequence or functional RNA. In general, a coding sequence is located 3' to a

5 promoter sequence. The promoter sequence consists of proximal and more distal upstream elements, the latter elements often referred to as enhancers. Accordingly, an “enhancer” is a DNA sequence which can stimulate promoter activity and may be an innate element of the promoter or a heterologous element inserted to enhance the level or tissue-specificity of a promoter. Promoters may be derived in their entirety from a native gene, or be composed of
10 different elements derived from different promoters found in nature, or even comprise synthetic DNA segments. It is understood by those skilled in the art that different promoters may direct the expression of a gene in different tissues or cell types, or at different stages of development, or in response to different environmental conditions. Promoters which cause a gene to be expressed in most cell types at most times are commonly referred to as
15 “constitutive promoters”. New promoters of various types useful in plant cells are constantly being discovered; numerous examples may be found in the compilation by Okamuro and Goldberg, (1989) *Biochemistry of Plants* 15:1-82. It is further recognized that since in most cases the exact boundaries of regulatory sequences have not been completely defined, DNA fragments of different lengths may have identical promoter activity.

20 The “translation leader sequence” refers to a DNA sequence located between the promoter sequence of a gene and the coding sequence. The translation leader sequence is present in the fully processed mRNA upstream of the translation start sequence. The translation leader sequence may affect processing of the primary transcript to mRNA, mRNA stability or translation efficiency. Examples of translation leader sequences have
25 been described (Turner, R. and Foster, G.D. (1995) *Molecular Biotechnology* 3:225).

The “3' non-coding sequences” refer to DNA sequences located downstream of a coding sequence and include polyadenylation recognition sequences and other sequences encoding regulatory signals capable of affecting mRNA processing or gene expression. The polyadenylation signal is usually characterized by affecting the addition of polyadenylic acid tracts to the 3' end of the mRNA precursor. The use of different 3' non-coding sequences is exemplified by Ingelbrecht et al., (1989) *Plant Cell* 1:671-680.

30 “RNA transcript” refers to the product resulting from RNA polymerase-catalyzed transcription of a DNA sequence. When the RNA transcript is a perfect complementary copy of the DNA sequence, it is referred to as the primary transcript or it may be a RNA
35 sequence derived from posttranscriptional processing of the primary transcript and is referred to as the mature RNA. “Messenger RNA (mRNA)” refers to the RNA that is without introns and that can be translated into protein by the cell. “cDNA” refers to a double-stranded DNA that is complementary to and derived from mRNA. “Sense” RNA refers to RNA transcript that includes the mRNA and so can be translated into protein by the

cell. "Antisense RNA" refers to a RNA transcript that is complementary to all or part of a target primary transcript or mRNA and that blocks the expression of a target gene (U.S. Pat. No. 5,107,065). The complementarity of an antisense RNA may be with any part of the specific gene transcript, i.e., at the 5' non-coding sequence, 3' non-coding sequence, introns, 5 or the coding sequence. "Functional RNA" refers to antisense RNA, ribozyme RNA, or other RNA that is not translated yet has an effect on cellular processes.

The term "operably linked" refers to the association of nucleic acid sequences on a single nucleic acid fragment so that the function of one is affected by the other. For example, a promoter is operably linked with a coding sequence when it is capable of 10 affecting the expression of that coding sequence (i.e., that the coding sequence is under the transcriptional control of the promoter). Coding sequences can be operably linked to regulatory sequences in sense or antisense orientation.

The term "expression", as used herein, refers to the transcription and stable accumulation of sense (mRNA) or antisense RNA derived from the nucleic acid fragment of 15 the invention. Expression may also refer to translation of mRNA into a polypeptide. "Antisense inhibition" refers to the production of antisense RNA transcripts capable of suppressing the expression of the target protein. "Overexpression" refers to the production of a gene product in transgenic organisms that exceeds levels of production in normal or non-transformed organisms. "Co-suppression" refers to the production of sense RNA 20 transcripts capable of suppressing the expression of identical or substantially similar foreign or endogenous genes (U.S. Patent No. 5,231,020).

"Altered levels" refers to the production of gene product(s) in transgenic organisms in amounts or proportions that differ from that of normal or non-transformed organisms.

"Mature" protein refers to a post-translationally processed polypeptide; i.e., one from 25 which any pre- or propeptides present in the primary translation product have been removed. "Precursor" protein refers to the primary product of translation of mRNA; i.e., with pre- and propeptides still present. Pre- and propeptides may be but are not limited to intracellular localization signals.

A "chloroplast transit peptide" is an amino acid sequence which is translated in 30 conjunction with a protein and directs the protein to the chloroplast or other plastid types present in the cell in which the protein is made. "Chloroplast transit sequence" refers to a nucleotide sequence that encodes a chloroplast transit peptide. A "signal peptide" is an amino acid sequence which is translated in conjunction with a protein and directs the protein to the secretory system (Chrispeels, J.J., (1991) *Ann. Rev. Plant Phys. Plant Mol. Biol.* 42:21-53). If the protein is to be directed to a vacuole, a vacuolar targeting signal (*supra*) 35 can further be added, or if to the endoplasmic reticulum, an endoplasmic reticulum retention signal (*supra*) may be added. If the protein is to be directed to the nucleus, any signal peptide present should be removed and instead a nuclear localization signal included (Raikhel (1992) *Plant Phys.* 100:1627-1632).

“Transformation” refers to the transfer of a nucleic acid fragment into the genome of a host organism, resulting in genetically stable inheritance. Host organisms containing the transformed nucleic acid fragments are referred to as “transgenic” organisms. Examples of methods of plant transformation include Agrobacterium-mediated transformation (De Blaere 5 et al. (1987) *Meth. Enzymol.* 143:277) and particle-accelerated or “gene gun” transformation technology (Klein et al. (1987) *Nature (London)* 327:70-73; U.S. Pat. No. 4,945,050).

Standard recombinant DNA and molecular cloning techniques used herein are well known in the art and are described more fully in Sambrook, J., Fritsch, E.F. and Maniatis, T. 10 *Molecular Cloning: A Laboratory Manual*; Cold Spring Harbor Laboratory Press: Cold Spring Harbor, 1989 (hereinafter “Maniatis”).

Nucleic acid fragments encoding at least a portion of several plant amino acid biosynthetic enzymes have been isolated and identified by comparison of random plant cDNA sequences to public databases containing nucleotide and protein sequences using the 15 BLAST algorithms well known to those skilled in the art. Table 1 lists the amino acid biosynthetic enzymes that are described herein, and the designation of the cDNA clones that comprise the nucleic acid fragments encoding these enzymes.

TABLE 1
Amino Acid Biosynthetic Enzymes

Enzyme	Clone	Plant
dihydrodipicolinate reductase	cs1.pk0083.b10	corn
	rls2.pk0017.d3	rice
diaminopimelate epimerase	chp2.pk0008.h4	corn
	rls48.pk0036.h10	rice
	se2.pk0005.f1	soybean
	ses8w.pk0010.f11	soybean
	sfl1.pk0031.h3	soybean
	sgs1c.pk002.k12	soybean
	wlm24.pk0030.g4	wheat
threonine synthase	cc2.pk0031.c9	corn
	cs1.pk0058.g5	corn
	rls72.pk0018.e7	rice
	se1.06a03	soybean
	srl.pk0003.f6	soybean
	wrl.pk0085.h2	wheat
threonine deaminase	cen1.pk0064.f4	corn
	sfl1.pk0055.h7	soybean
	sre.pk0044.f3	soybean

Enzyme	Clone	Plant
s-adenosylmethionine synthase	cc3.mn0002.d2	corn
	se2.12b06	soybean
	wre1.pk0002.c12	wheat
	wle1n.pk0070.b8	wheat
	wkm1c.pk0003.g4	wheat
	wlk1.pk0028.d3	wheat
	wre1n.pk170.d8	wheat
	wr1.pk0086.d5	wheat
	wr1.pk0103.h8	wheat
	wre1n.pk0082.b2	wheat

The nucleic acid fragments of the instant invention may be used to isolate cDNAs and genes encoding homologous enzymes from the same or other plant species. Isolation of 5 homologous genes using sequence-dependent protocols is well known in the art. Examples of sequence-dependent protocols include, but are not limited to, methods of nucleic acid hybridization, and methods of DNA and RNA amplification as exemplified by various uses of nucleic acid amplification technologies (e.g., polymerase chain reaction, ligase chain reaction).

10 For example, genes encoding other amino acid biosynthetic enzymes, either as cDNAs or genomic DNAs, could be isolated directly by using all or a portion of the instant nucleic acid fragments as DNA hybridization probes to screen libraries from any desired plant employing methodology well known to those skilled in the art. Specific oligonucleotide probes based upon the instant nucleic acid sequences can be designed and synthesized by 15 methods known in the art (Maniatis). Moreover, the entire sequences can be used directly to synthesize DNA probes by methods known to the skilled artisan such as random primers DNA labeling, nick translation, or end-labeling techniques, or RNA probes using available *in vitro* transcription systems. In addition, specific primers can be designed and used to amplify a part of or full-length of the instant sequences. The resulting amplification 20 products can be labeled directly during amplification reactions or labeled after amplification reactions, and used as probes to isolate full length cDNA or genomic fragments under conditions of appropriate stringency.

25 In addition, two short segments of the instant nucleic acid fragments may be used in polymerase chain reaction protocols to amplify longer nucleic acid fragments encoding homologous genes from DNA or RNA. The polymerase chain reaction may also be performed on a library of cloned nucleic acid fragments wherein the sequence of one primer is derived from the instant nucleic acid fragments, and the sequence of the other primer takes advantage of the presence of the polyadenylic acid tracts to the 3' end of the mRNA 30 precursor encoding plant genes. Alternatively, the second primer sequence may be based upon sequences derived from the cloning vector. For example, the skilled artisan can follow

the RACE protocol (Frohman et al., (1988) *PNAS USA* 85:8998) to generate cDNAs by using PCR to amplify copies of the region between a single point in the transcript and the 3' or 5' end. Primers oriented in the 3' and 5' directions can be designed from the instant sequences. Using commercially available 3' RACE or 5' RACE systems (BRL), specific 3' or 5' cDNA fragments can be isolated (Ohara et al., (1989) *PNAS USA* 86:5673; Loh et al., (1989) *Science* 243:217). Products generated by the 3' and 5' RACE procedures can be combined to generate full-length cDNAs (Frohman, M.A. and Martin, G.R., (1989) *Techniques* 1:165).

Availability of the instant nucleotide and deduced amino acid sequences facilitates immunological screening cDNA expression libraries. Synthetic peptides representing portions of the instant amino acid sequences may be synthesized. These peptides can be used to immunize animals to produce polyclonal or monoclonal antibodies with specificity for peptides or proteins comprising the amino acid sequences. These antibodies can be then be used to screen cDNA expression libraries to isolate full-length cDNA clones of interest (Lerner, R.A. (1984) *Adv. Immunol.* 36:1; Maniatis).

The nucleic acid fragments of the instant invention may be used to create transgenic plants in which the disclosed biosynthetic enzymes are present at higher or lower levels than normal or in cell types or developmental stages in which they are not normally found. This would have the effect of altering the level of free amino acids in those cells.

Overexpression of the biosynthetic enzymes of the instant invention may be accomplished by first constructing chimeric genes in which the coding region are operably linked to promoters capable of directing expression of a gene in the desired tissues at the desired stage of development. For reasons of convenience, the chimeric genes may comprise promoter sequences and translation leader sequences derived from the same genes. 3' Non-coding sequences encoding transcription termination signals may also be provided. The instant chimeric genes may also comprise one or more introns in order to facilitate gene expression.

Plasmid vectors comprising the instant chimeric genes can then constructed. The choice of plasmid vector is dependent upon the method that will be used to transform host plants. The skilled artisan is well aware of the genetic elements that must be present on the plasmid vector in order to successfully transform, select and propagate host cells containing the chimeric gene. The skilled artisan will also recognize that different independent transformation events will result in different levels and patterns of expression (Jones et al., (1985) *EMBO J.* 4:2411-2418; De Almeida et al., (1989) *Mol. Gen. Genetics* 218:78-86), and thus that multiple events must be screened in order to obtain lines displaying the desired expression level and pattern. Such screening may be accomplished by Southern analysis of DNA, Northern analysis of mRNA expression, Western analysis of protein expression, or phenotypic analysis.

For some applications it may be useful to direct the instant biosynthetic enzymes to different cellular compartments, or to facilitate their secretion from the cell. It is thus envisioned that the chimeric genes described above may be further supplemented by altering the coding sequences to encode enzymes with appropriate intracellular targeting sequences 5 such as transit sequences (Keegstra, K. (1989) *Cell* 56:247-253), signal sequences or sequences encoding endoplasmic reticulum localization (Chrispeels, J.J., (1991) *Ann. Rev. Plant Phys. Plant Mol. Biol.* 42:21-53), or nuclear localization signals (Raikhel, N. (1992) *Plant Phys.* 100:1627-1632) added and/or with targeting sequences that are already present removed. While the references cited give examples of each of these, the list is not 10 exhaustive and more targeting signals of utility may be discovered in the future.

It may also be desirable to reduce or eliminate expression of the genes encoding the instant biosynthetic enzymes in plants for some applications. In order to accomplish this, chimeric genes designed for co-suppression of the instant biosynthetic enzymes can be constructed by linking the genes or gene fragments encoding the enzymes to plant promoter 15 sequences. Alternatively, chimeric genes designed to express antisense RNA for all or part of the instant nucleic acid fragments can be constructed by linking the genes or gene fragment in reverse orientation to plant promoter sequences. Either the co-suppression or antisense chimeric genes could be introduced into plants via transformation wherein expression of the corresponding endogenous genes are reduced or eliminated.

20 The instant amino acid biosynthetic enzymes (or portions of the enzymes) may be produced in heterologous host cells, particularly in the cells of microbial hosts, and can be used to prepare antibodies to the enzymes by methods well known to those skilled in the art. The antibodies are useful for detecting the enzymes *in situ* in cells or *in vitro* in cell extracts. Preferred heterologous host cells for production of the instant amino acid biosynthetic 25 enzymes are microbial hosts. Microbial expression systems and expression vectors containing regulatory sequences that direct high level expression of foreign proteins are well known to those skilled in the art. Any of these could be used to construct chimeric genes for production of the instant amino acid biosynthetic enzymes. These chimeric genes could then be introduced into appropriate microorganisms via transformation to provide high level 30 expression of the enzymes. An example of a vector for high level expression of the instant amino acid biosynthetic enzymes in a bacterial host is provided (Example 11).

35 Additionally, the instant plant amino acid biosynthetic enzymes can be used as a targets to facilitate design and/or identification of inhibitors of the enzymes that may be useful as herbicides. This is desirable because the enzymes described herein catalyze various steps in a pathway leading to production of several essential amino acids. Accordingly, inhibition of the activity of one or more of the enzymes described herein could lead to inhibition of amino acid biosynthesis sufficient to inhibit plant growth. Thus, the instant plant amino acid biosynthetic enzymes could be appropriate for new herbicide discovery and design.

All or a substantial portion of the nucleic acid fragments of the instant invention may also be used as probes for genetically and physically mapping the genes that they are a part of, and as markers for traits linked to those genes. Such information may be useful in plant breeding in order to develop lines with desired phenotypes. For example, the instant nucleic acid fragments may be used as restriction fragment length polymorphism (RFLP) markers. Southern blots (Maniatis) of restriction-digested plant genomic DNA may be probed with the nucleic acid fragments of the instant invention. The resulting banding patterns may then be subjected to genetic analyses using computer programs such as MapMaker (Lander et al., (1987) *Genomics* 1:174-181) in order to construct a genetic map. In addition, the nucleic acid fragments of the instant invention may be used to probe Southern blots containing restriction endonuclease-treated genomic DNAs of a set of individuals representing parent and progeny of a defined genetic cross. Segregation of the DNA polymorphisms is noted and used to calculate the position of the instant nucleic acid sequence in the genetic map previously obtained using this population (Botstein, D. et al., (1980) *Am. J. Hum. Genet.* 32:314-331).

The production and use of plant gene-derived probes for use in genetic mapping is described in R. Bernatzky, R. and Tanksley, S. D. (1986) *Plant Mol. Biol. Reporter* 4(1):37-41. Numerous publications describe genetic mapping of specific cDNA clones using the methodology outlined above or variations thereof. For example, F2 intercross populations, backcross populations, randomly mated populations, near isogenic lines, and other sets of individuals may be used for mapping. Such methodologies are well known to those skilled in the art.

Nucleic acid probes derived from the instant nucleic acid sequences may also be used for physical mapping (i.e., placement of sequences on physical maps; see Hoheisel, J. D., et al., In: *Nonmammalian Genomic Analysis: A Practical Guide*, Academic press 1996, pp. 319-346, and references cited therein).

In another embodiment, nucleic acid probes derived from the instant nucleic acid sequences may be used in direct fluorescence *in situ* hybridization (FISH) mapping (Trask, B. J. (1991) *Trends Genet.* 7:149-154). Although current methods of FISH mapping favor use of large clones (several to several hundred KB; see Laan, M. et al. (1995) *Genome Research* 5:13-20), improvements in sensitivity may allow performance of FISH mapping using shorter probes.

A variety of nucleic acid amplification-based methods of genetic and physical mapping may be carried out using the instant nucleic acid sequences. Examples include allele-specific amplification (Kazazian, H. H. (1989) *J. Lab. Clin. Med.* 114(2):95-96), polymorphism of PCR-amplified fragments (CAPS; Sheffield, V. C. et al. (1993) *Genomics* 16:325-332), allele-specific ligation (Landegren, U. et al. (1988) *Science* 241:1077-1080), nucleotide extension reactions (Sokolov, B. P. (1990) *Nucleic Acid Res.* 18:3671), Radiation Hybrid Mapping (Walter, M. A. et al. (1997) *Nature Genetics* 7:22-28) and Happy Mapping

(Dear, P. H. and Cook, P. R. (1989) *Nucleic Acid Res.* 17:6795-6807). For these methods, the sequence of a nucleic acid fragment is used to design and produce primer pairs for use in the amplification reaction or in primer extension reactions. The design of such primers is well known to those skilled in the art. In methods employing PCR-based genetic mapping, 5 it may be necessary to identify DNA sequence differences between the parents of the mapping cross in the region corresponding to the instant nucleic acid sequence. This, however, is generally not necessary for mapping methods.

Loss of function mutant phenotypes may be identified for the instant cDNA clones either by targeted gene disruption protocols or by identifying specific mutants for these 10 genes contained in a maize population carrying mutations in all possible genes (Ballinger and Benzer, (1989) *Proc. Natl. Acad. Sci USA* 86:9402; Koes et al., (1995) *Proc. Natl. Acad. Sci USA* 92:8149; Bensen et al., (1995) *Plant Cell* 7:75). The latter approach may be accomplished in two ways. First, short segments of the instant nucleic acid fragments may be used in polymerase chain reaction protocols in conjunction with a mutation tag sequence 15 primer on DNAs prepared from a population of plants in which Mutator transposons or some other mutation-causing DNA element has been introduced (see Bensen, *supra*). The amplification of a specific DNA fragment with these primers indicates the insertion of the mutation tag element in or near the plant gene encoding dihydrodipicolinate reductase, diaminopimelate epimerase, threonine synthase, threonine deaminase or S-adenosyl- 20 methionine synthetase. Alternatively, the instant nucleic acid fragment may be used as a hybridization probe against PCR amplification products generated from the mutation population using the mutation tag sequence primer in conjunction with an arbitrary genomic site primer, such as that for a restriction enzyme site-anchored synthetic adaptor. With either method, a plant containing a mutation in the endogenous gene encoding a 25 dihydrodipicolinate reductase, diaminopimelate epimerase, threonine synthase, threonine deaminase or S-adenosylmethionine synthetase can be identified and obtained. This mutant plant can then be used to determine or confirm the natural function of the dihydro- dipicolinate reductase, diaminopimelate epimerase, threonine synthase, threonine deaminase and S-adenosylmethionine synthetase gene product.

EXAMPLES

The present invention is further defined in the following Examples, in which all parts and percentages are by weight and degrees are Celsius, unless otherwise stated. It should be understood that these Examples, while indicating preferred embodiments of the invention, are given by way of illustration only. From the above discussion and these Examples, one 35 skilled in the art can ascertain the essential characteristics of this invention, and without departing from the spirit and scope thereof, can make various changes and modifications of the invention to adapt it to various usages and conditions.

EXAMPLE 1Composition of cDNA Libraries; Isolation and Sequencing of cDNA Clones

cDNA libraries representing mRNAs from various corn, rice, soybean and wheat tissues were prepared. The characteristics of the libraries are described below.

5

TABLE 2

cDNA Libraries from Corn and Soybean Tissues

Library	Tissue	Clone
cc2	Corn Callus, Partially Differentiated, 2 Weeks After Subculture	cc2.pk0031.c9
cc3	Corn Callus, Mature Somatic Embryo	cc3.mn0002.d2
cen1	Corn Endosperm 12 Days After Pollination	cen1.pk0064.f4
chp2	Corn Leaf, 11 Day Old Plant	chp2.pk0008.h4
cs1	Corn Leaf, Sheath 5 Week Old Plant	cs1.pk0058.g5
csi1n	Corn Silk*	csi1n.pk0042.a3
rls2	Rice Leaf 15 Days After Germination, 2 Hours After Infection of Strain <i>Magaporthe grisea</i> 4360-R-67 (AVR2-YAMO); Susceptible	rls2.pk0017.d3
rls48	Rice Leaf 15 Days After Germination, 48 Hours After Infection of Strain <i>Magaporthe grisea</i> 4360-R-67 (AVR2-YAMO); Susceptible	rls48.pk0036.h10
rls72	Rice Leaf 15 Days After Germination, 72 Hours After Infection of Strain <i>Magaporthe grisea</i> 4360-R-67 (AVR2-YAMO); Susceptible	rls72.pk0018.e7
s2	Soybean Seed, 19 Days After Flowering	s2.12b06
se1	Soybean Embryo 7 Days After Flowering	se1.06a03
se2	Soybean Embryo 10 Days After Flowering	se2.pk0005.f1
ses8w	Mature Soybean Embryo 8 Weeks After Subculture	ses8w.pk0010.h11
sfl1	Soybean Immature Flower	sfl1.pk0055.h7
sgs1c	Soybean Seeds 4 Hours After Germination	sgs1c.pk002.k12
sr1	Soybean Root From 10 Day Old Seedlings	sr1.pk0003.f6
sre	Soybean Root Elongation 4-5 Days After Germination	sre.pk0044.f3
wkm1c	Wheat Kernel Malted 55 Hours at 22 Degrees Celsius	wkm1c.pk0003.g4
wle1n	Wheat Leaf From 7 Day Old Etiolated Seedling*	wle1n.pk0070.b8
wlk1	Wheat Seedlings 1 Hour After Treatment with Fungicide**	wlk1.pk0028.d3
wlm24	Wheat Seedlings 24 Hours After Inoculation With <i>Erysiphe graminis</i> f. sp <i>tritici</i>	wlm24.pk0030.g4
wr1	Wheat Root From 7 Day Old Seedling	wr1.pk0085.h2
wre1	Wheat Root From 7 Day Old Etiolated Seedling	wre1.pk0002.c12
wre1n	Wheat Root From 7 Day Old Etiolated Seedling*	wre1n.pk0082.b2
		wre1n.pk170.d8

*These libraries were normalized essentially as described in U.S. Pat. No. 5,482,845

** Application of 6-iodo-2-propoxy-3-propyl-4(3*H*)-quinazolinone; synthesis and methods of using this compound are described in USSN 08/545,827, incorporated herein by reference.

5 cDNA libraries were prepared in Uni-ZAP™ XR vectors according to the manufacturer's protocol (Stratagene Cloning Systems, La Jolla, CA). Conversion of the Uni-ZAP™ XR libraries into plasmid libraries was accomplished according to the protocol provided by Stratagene. Upon conversion, cDNA inserts were contained in the plasmid vector pBluescript. cDNA inserts from randomly picked bacterial colonies containing
10 recombinant pBluescript plasmids were amplified via polymerase chain reaction using primers specific for vector sequences flanking the inserted cDNA sequences, or plasmid DNA was prepared from cultured bacterial cells. Amplified insert DNAs or plasmid DNAs were sequenced in dye-primer sequencing reactions to generate partial cDNA sequences (expressed sequence tags or "ESTs"; see Adams, M. D. et al., (1991) *Science* 252:1651).
15 The resulting ESTs were analyzed using a Perkin Elmer Model 377 fluorescent sequencer.

EXAMPLE 2

Identification and Characterization of cDNA Clones

ESTs encoding plant amino acid biosynthetic enzymes were identified by conducting BLAST (Basic Local Alignment Search Tool; Altschul, S. F., et al., (1993) *J. Mol. Biol.* 215:403-410; see also www.ncbi.nlm.nih.gov/BLAST/) searches for similarity to sequences contained in the BLAST "nr" database (comprising all non-redundant GenBank CDS translations, sequences derived from the 3-dimensional structure Brookhaven Protein Data Bank, the last major release of the SWISS-PROT protein sequence database, EMBL, and DDBJ databases). The cDNA sequences obtained in Example 1 were analyzed for similarity
25 to all publicly available DNA sequences contained in the "nr" database using the BLASTN algorithm provided by the National Center for Biotechnology Information (NCBI). The DNA sequences were translated in all reading frames and compared for similarity to all publicly available protein sequences contained in the "nr" database using the BLASTX algorithm (Gish, W. and States, D. J. (1993) *Nature Genetics* 3:266-272) provided by the
30 NCBI. For convenience, the P-value (probability) of observing a match of a cDNA sequence to a sequence contained in the searched databases merely by chance as calculated by BLAST are reported herein as "pLog" values, which represent the negative of the logarithm of the reported P-value. Accordingly, the greater the pLog value, the greater the likelihood that the cDNA sequence and the BLAST "hit" represent homologous proteins.
35

EXAMPLE 3

Characterization of cDNA Clones Encoding Polypeptides Homologous to Dihydrodipicolinate Reductase

The BLASTX search using the nucleotide sequences from clones csl1n.pk0042.a3 and rls2.pk0017.d3 revealed similarity of the protein encoded by the cDNA to *Synechocystis sp.*

dihydrodipicolinate reductase enzyme (DDBJ Accession No. D90899). BLAST pLog values were 12.60 and 11.68 for csi1n.pk0042.a3 and rls2.pk0017.d3, respectively.

The sequence of the entire cDNA insert in clone csi1n.pk0042.a3 was determined and is shown in SEQ ID NO:1; the deduced amino acid sequence of this cDNA is shown in SEQ 5 ID NO:2. The amino acid sequence set forth in SEQ ID NO:2 was evaluated by BLASTP, yielding a pLog value of 36.72 versus the *Synechocystis* sp. dihydrodipicolinate reductase sequence. The sequence of a portion of the cDNA insert from clone rls2.pk0017.d3 is shown in SEQ ID NO:3; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:4. Figure 2 presents an alignment of the amino acid sequences set forth in SEQ ID NO:2 and 10 the *Synechocystis* sp. dihydrodipicolinate reductase sequence (SEQ ID NO:5). SEQ ID NO:2 is 40% identical to the *Synechocystis* sp. dihydrodipicolinate reductase sequence (SEQ ID NO:5). Sequence alignments were performed by the Clustal method of alignment (Higgins, D.G. and Sharp, P.M. (1989) CABIOS 5:151-153), using the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI). 15 Sequence percent identity calculations were performed by the Jotun Hein method (Hein, J. J. (1990) *Meth. Enz.* 183:626-645) using the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI).

Sequence alignments and BLAST scores and probabilities indicate that the instant 20 nucleic acid fragments encode a nearly entire corn dihydrodipicolinate reductase, and a portion of a rice dihydrodipicolinate reductase. These sequences represent the first plant sequences encoding dihydrodipicolinate reductase.

EXAMPLE 4

Characterization of cDNA Clones Encoding Diaminopimelate Epimerase

The BLASTX search using the nucleotide sequences from clones chp2.pk0008.h4, 25 rls48.pk0036.h10, wlm24.pk0030.g4, and the contig sequences assembled from clones se2.pk0005.f1, ses8w.pk0010.h11, sfl1.pk0031.h3, and sgs1c.pk002.k12 revealed similarity of the proteins encoded by the cDNAs to diaminopimelate epimerase from *Synechocystis* sp. (DDBJ Accession No. D90917). The BLAST results for each of these ESTs are shown in Table 3:

30

TABLE 3
 BLAST Results for Clones Encoding Polypeptides Homologous
 to Diaminopimelate Epimerase

Clone	BLAST pLog Score DDBJ Accession No. D90917
chp2.pk0008.h4	59.16
rls48.pk0036.h10	40.82
The contig of:	98.30
se2.pk0005.f1	
ses8w.pk0010.h11	
sfl1.pk0031.h3	
sgs1c.pk002.k12	
wlm24.pk0030.g4	23.46

5 The sequence of the entire cDNA insert in clone chp2.pk0008.h4 was determined and
 is shown in SEQ ID NO:6; the deduced amino acid sequence of this cDNA is shown in SEQ
 ID NO:7. The amino acid sequence set forth in SEQ ID NO:7 was evaluated by BLASTP,
 yielding a pLog value of 75.66 versus the *Synechocystis sp.* sequence. The sequence of a
 portion of the cDNA insert from clone rls48.pk0036.h10 is shown in SEQ ID NO:8; the
 10 deduced amino acid sequence of this cDNA is shown in SEQ ID NO:9. The nucleotide
 sequence of the contig assembled from clones se2.pk0005.f1, ses8w.pk0010.h11,
 sfl1.pk0031.h3, and sgs1c.pk002.k12 was determined and is shown in SEQ ID NO:10; the
 deduced amino acid sequence of this cDNA is shown in SEQ ID NO:11. The amino acid
 sequence set forth in SEQ ID NO:11 was evaluated by BLASTP, yielding a pLog value of
 15 98.57 versus the *Synechocystis sp.* sequence. The sequence of a portion of the cDNA insert
 from clone wlm24.pk0030.g4 is shown in SEQ ID NO:12; the deduced amino acid sequence
 of this cDNA is shown in SEQ ID NO:13. Figure 3 presents an alignment of the amino acid
 sequences set forth in SEQ ID NOs: 7, 9, 11, and 13 and the *Synechocystis sp.* sequence
 (SEQ ID NO:14). The data in Table 4 represents a calculation of the percent identity of the
 20 amino acid sequences set forth in SEQ ID NOs: 7, 9, 11, and 13 and the *Synechocystis sp.*
 sequence.

TABLE 4

Percent Identity of Amino Acid Sequences Deduced From the Nucleotide Sequences of cDNA Clones Encoding Polypeptides Homologous to Diaminopimelate Epimerase

Clone	SEQ ID NO.	Percent Identity to DDBJ Accession No. D90917 (SEQ ID NO:16)
chp2.pk0008.h4	7	59
rls48.pk0036.h10	9	74
Contig of:		
se2.pk0005.f1		
ses8w.pk0010.h11		
sfl1.pk0031.h3		
sgs1c.pk002.k12		
wlm24.pk0030.g4	13	65

5

Sequence alignments were performed by the Clustal method of alignment (Higgins, D.G. and Sharp, P.M. (1989) CABIOS 5:151-153), using the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI). Sequence percent identity calculations were performed by the Jotun Hein method (Hein, J. J. (1990) *Meth. Enz.* 183:626-645) using the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI).

Sequence alignments and BLAST scores and probabilities indicate that the instant nucleic acid fragments encode a nearly entire corn diaminopimelate epimerase (chp2.pk0008.h4), a portion of a rice diaminopimelate epimerase (rls48.pk0036.h10), and an entire soybean diaminopimelate epimerase (se2.pk0005.f1, ses8w.pk0010.h11, sfl1.pk0031.h3, and sgs1c.pk002.k12), and a portion of a wheat diaminopimelate epimerase (wlm24.pk0030.g4). These sequences represent the first plant sequences encoding diaminopimelate epimerase enzyme.

EXAMPLE 5**Characterization of cDNA Clones Encoding Threonine Synthase**

The BLASTX search using the EST sequences from clones cc2.pk0031.c9, cs1.pk0058.g5, rls72.pk0018.e7, se1.06a03, sr1.pk0003.f6, and wr1.pk0085.h2 revealed similarity of the proteins encoded by the cDNAs to threonine synthase from *Arabidopsis thaliana* (GenBank Accession No. L41666). The BLAST results for each of these ESTs are shown in Table 5:

TABLE 5
 BLAST Results for Clones Encoding Polypeptides Homologous
 to Threonine Synthase

Clone	BLAST pLog Score L41666
cc2.pk0031.c9	56.19
cs1.pk0058.g5	8.00
rls72.pk0018.e7	29.47
se1.06a03	34.15
srl.pk0003.f6	21.13
wrl.pk0085.h2	29.47

5 The sequence of the entire cDNA insert in clone cc2.pk0031.c9 was determined and is shown in SEQ ID NO:15; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:16. The amino acid sequence set forth in SEQ ID NO:16 was evaluated by BLASTP, yielding a pLog value of 166.11 versus the *Arabidopsis thaliana* sequence. BLASTN against dbest indicated identity of nucleotides 520 through 684 from cc2.pk0031.c9 with
 10 nucleotides 1 through 162 of a corn EST (GenBank Accession No. T18847). The sequence of a portion of the cDNA insert from clone cs1.pk0058.g5 is shown in SEQ ID NO:17; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:18. The sequence of a portion of the cDNA insert from clone rls72.pk0018.e7 is shown in SEQ ID NO:19; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:20. The sequence of a portion of the cDNA insert from clone se1.06a03 is shown in SEQ ID NO:21; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:22. The sequence of the entire
 15 cDNA insert in clone srl.pk0003.f6 was determined and is shown in SEQ ID NO:23; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:24. The amino acid sequence set forth in SEQ ID NO:24 was evaluated by BLASTP, yielding a pLog value of
 20 275.06 versus the *Arabidopsis thaliana* sequence. The sequence of a portion of the cDNA insert from clone wrl.pk0085.h2 is shown in SEQ ID NO:25; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:26. Figure 4 presents an alignment of the amino acid sequences set forth in SEQ ID NOs:16, 18, 20, 22, 24, and 26 and the
 25 *Arabidopsis thaliana* sequence. The data in Table 6 represents a calculation of the percent identity of the amino acid sequences set forth in SEQ ID NOs:16, 18, 20, 22, 24, and 26 and the *Arabidopsis thaliana* sequence (SEQ ID NO:27).

TABLE 6

Percent Identity of Amino Acid Sequences Deduced From the Nucleotide Sequences of cDNA Clones Encoding Polypeptides Homologous to Threonine Synthase

Clone	SEQ ID NO.	Percent Identity to L41666 (SEQ ID NO:29)
cc2.pk0031.c9	16	81.0
cs1.pk0058.g5	18	81.0
rls72.pk0018.e7	20	55.3
se1.06a03	22	80.0
sr1.pk0003.f6	24	84.4
wr1.pk0085.h2	26	50.4

5 Sequence alignments were performed by the Clustal method of alignment (Higgins, D.G. and Sharp, P.M. (1989) *CABIOS* 5:151-153), using the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI). Sequence percent identity calculations were performed by the Jotun Hein method (Hein, J. J. (1990) 10 *Meth. Enz.* 183:626-645) using the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI).

Sequence alignments and BLAST scores and probabilities indicate that the instant nucleic acid fragments encode portions of a corn threonine synthase (cc2.pk0031.c9 and 15 cs1.pk0058.g5), a portion of a rice threonine synthase (rls72.pk0018.e7), portions of a soybean threonine synthase (se1.06a03 and sr1.pk0003.f6), and a portion of a wheat threonine synthase (wr1.pk0085.h2). These sequences represent the first corn, rice, soybean, and wheat sequences encoding threonine synthase.

EXAMPLE 6

Characterization of cDNA Clones Encoding Threonine Deaminase

20 The BLASTX search using the EST sequence from clone cen1.pk0064.f4 revealed similarity of the protein encoded by the cDNA to threonine deaminase from *Brukholderia capacia* (GenBank Accession No. U40630; pLog = 31.38). The BLASTX search using the EST sequences from clones sfl1.pk0055.h7 and sre.pk0044.f3 revealed similarity of the proteins encoded by the cDNAs to threonine deaminase from *Solanum tuberosum* and 25 *Brukholderia capacia* (EMBL Accession No. X67846 and GenBank Accession No. U40630, respectively). BLAST pLog values were 36.55 and 31.79 for sfl1.pk0055.h7, and 19.47 and 14.51 for sre.pk0044.f3.

The sequence of the entire cDNA insert in clone cen1.pk0064.f4 was determined and is shown in SEQ ID NO:28; the deduced amino acid sequence of this cDNA is shown in 30 SEQ ID NO:29. The amino acid sequence set forth in SEQ ID NO:29 was evaluated by BLASTP, yielding a pLog value of 134.85 versus the *Brukholderia capacia* sequence. The sequence of a portion of the cDNA insert from clone sfl1.pk0055.h7 is shown in SEQ ID

NO:30; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:31. The sequence of the entire cDNA insert in clone sre.pk0044.f3 was determined and is shown in SEQ ID NO:32; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:33. The amino acid sequence set forth in SEQ ID NO:33 was evaluated by BLASTP, yielding 5 pLog values of 19.24 versus the *Solanum tuberosum* sequence and 15.19 versus the *Brukholderia capacia* threonine deaminase sequence. Figure 5 presents an alignment of the amino acid sequences set forth in SEQ ID NOs:29, 31, and 33 and the *Brukholderia capacia* (SEQ ID NO:34) sequence. The data in Table 7 represents a calculation of the percent identity of the amino acid sequences set forth in SEQ ID NOs:29, 31, and 33 35 and the 10 *Brukholderia capacia* sequence.

TABLE 7

Percent Identity of Amino Acid Sequences Deduced From the Nucleotide Sequences of cDNA Clones Encoding Polypeptides Homologous to Threonine Deaminase

Clone	SEQ ID NO.	Percent Identity to U40630 (SEQ ID NO:36)
cen1.pk0064.f4	29	61.0
sfl1.pk0055.h7	31	47.9
sre.pk0044.f3	33	46.0

Sequence alignments were performed by the Clustal method of alignment (Higgins, D.G. and Sharp, P.M. (1989) *CABIOS* 5:151-153), using the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI). Sequence 20 percent identity calculations were performed by the Jotun Hein method (Hein, J. J. (1990) *Meth. Enz.* 183:626-645) using the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI)

Sequence alignments and BLAST scores and probabilities indicate that the instant nucleic acid fragments encode a nearly entire corn threonine deaminase (cen1.pk0064.f4) 25 and portions of a soybean threonine deaminase (sfl1.pk0055.h7 and sre.pk0044.f3). These sequences represent the first corn and soybean sequences encoding threonine deaminase.

EXAMPLE 7**Characterization of cDNA Clones Encoding S-adenosylmethionine synthetase**

The BLASTX search using the nucleotide sequence from clone cc3.mn0002.d2 30 revealed similarity of the protein encoded by the cDNA to S-adenosylmethionine synthetase from *Oryza sativa* (EMBL Accession No. Z26867; pLog = 99.03). The sequence of the entire cDNA insert in clone cc3.mn0002.d2 was determined and is shown in SEQ ID NO:35; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:36. The nucleotide sequence set forth in SEQ ID NO:35 was evaluated by BLASTN, yielding a pLog value 35 larger than 200 versus the *Oryza sativa* sequence. Figure 6 presents an alignment of the

nucleotide sequences set forth in SEQ ID NO:35 and the *Oryza sativa* sequence (SEQ ID NO:37). The nucleotide sequence in SEQ ID NO:35 is 88% identical over 1216 nucleotides to the nucleotide sequence of the *Oryza sativa* S-adenosylmethionine synthetase.

The BLASTX search using the nucleotide sequence from clone s2.12b06 revealed 5 similarity of the protein encoded by the cDNA to S-adenosylmethionine synthetase from *Lycopersicon esculentum* (EMBL Accession No. Z24741; pLog = 62.62). The sequence of the entire cDNA insert in clone s2.12b06 was determined and is shown in SEQ ID NO:38; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:39. The nucleotide sequence set forth in SEQ ID NO:38 was evaluated by BLASTN, yielding a pLog value 10 larger than 200 versus the *Lycopersicon esculentum* sequence. Figure 7 presents an alignment of the nucleotide sequences set forth in SEQ ID NO:38 and the *Lycopersicon esculentum* sequence (SEQ ID NO:40). The nucleotide sequence set forth in SEQ ID NO:38 is 82 % identical over 1210 nucleotides to the *Lycopersicon esculentum* sequence.

The BLASTX search using the nucleotide sequence from the contig assembled from 15 clones wre1.pk0002.c12, wle1n.pk0070.b8, wkm1c.pk0003.g4, wlk1.pk0028.d3, wre1n.pk170.d8, wr1.pk0086.d5, wr1.pk0103.h8, and wre1n.pk0082.b2 revealed similarity of the protein encoded by the contig to S-adenosylmethionine synthetase from *Hordeum vulgare* (DDBJ Accession No. D63835) with a pLog value larger than 200. The nucleotide sequence of the contig assembled from clones wre1.pk0002.c12, wle1n.pk0070.b8, 20 wkm1c.pk0003.g4, wlk1.pk0028.d3, wre1n.pk170.d8, wr1.pk0086.d5, wr1.pk0103.h8, and wre1n.pk0082.b2 is shown in SEQ ID NO:41; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:42. Figure 8 presents an alignment of the nucleotide sequence set forth in SEQ ID NO:41 and the *Hordeum vulgare* sequence (SEQ ID NO:43). The SEQ ID NO:41 is 92% identical to the *Hordeum vulgare* sequence.

25 Sequence alignments and BLAST scores and probabilities indicate that the instant nucleic acid fragments encode entire or nearly entire corn, soybean, or wheat S-adenosylmethionine synthetase. These sequences represent the first corn, soybean, or wheat sequences encoding S-adenosylmethionine synthetase.

EXAMPLE 8

Expression of Chimeric Genes in Monocot Cells

30 A chimeric gene comprising a cDNA encoding an amino acid biosynthetic enzyme in sense orientation with respect to the maize 27 kD zein promoter that is located 5' to the cDNA fragment, and the 10 kD zein 3' end that is located 3' to the cDNA fragment, can be constructed. The cDNA fragment of this gene may be generated by polymerase chain 35 reaction (PCR) of the cDNA clone using appropriate oligonucleotide primers and under appropriate experimental conditions. Cloning sites (NcoI or SmaI) can be incorporated into the oligonucleotides to provide proper orientation of the DNA fragment when inserted into the digested vector pML103 as described below. The amplified DNA can then be digested with restriction enzymes NcoI and SmaI and fractionated on a 0.7% low melting point

agarose gel in 40 mM Tris-acetate, pH 8.5, 1 mM EDTA. The appropriate band can be excised from the gel, melted at 68°C and combined with a 4.9 kb NcoI-SmaI fragment of the plasmid pML103. Plasmid pML103 has been deposited under the terms of the Budapest Treaty at ATCC (American Type Culture Collection, 10801 University Boulevard, 5 Manassas, VA 20110-2209), and bears accession number ATCC 97366. The DNA segment from pML103 contains a 1.05 kb SalI-NcoI promoter fragment of the maize 27 kD zein gene and a 0.96 kb SmaI-SalI fragment from the 3' end of the maize 10 kD zein gene in the vector pGem9Zf(+) (Promega). Vector and insert DNA can be ligated at 15°C overnight, essentially as described (Maniatis). The ligated DNA may then be used to transform *E. coli* 10 XL1-Blue (Epicurian Coli XL-1 Blue™; Stratagene). Bacterial transformants can be screened by restriction enzyme digestion of plasmid DNA and limited nucleotide sequence analysis using the dideoxy chain termination method (Sequenase™ DNA Sequencing Kit; U.S. Biochemical). The resulting plasmid construct would comprise a chimeric gene 15 encoding, in the 5' to 3' direction, the maize 27 kD zein promoter, a cDNA fragment encoding a plant amino acid biosynthetic enzyme, and the 10 kD zein 3' region.

The chimeric gene described above can then be introduced into corn cells by the following procedure. Immature corn embryos can be dissected from developing caryopses derived from crosses of the inbred corn lines H99 and LH132. The embryos are isolated 10 to 11 days after pollination when they are 1.0 to 1.5 mm long. The embryos are then placed 20 with the axis-side facing down and in contact with agarose-solidified N6 medium (Chu et al., (1975) *Sci. Sin. Peking* 18:659-668). The embryos are kept in the dark at 27°C. Friable embryogenic callus consisting of undifferentiated masses of cells with somatic proembryoids and embryoids borne on suspensor structures proliferates from the scutellum 25 of these immature embryos. The embryogenic callus isolated from the primary explant can be cultured on N6 medium and sub-cultured on this medium every 2 to 3 weeks.

The plasmid, p35S/Ac (obtained from Dr. Peter Eckes, Hoechst Ag, Frankfurt, Germany) may be used in transformation experiments in order to provide for a selectable marker. This plasmid contains the *Pat* gene (see European Patent Publication 0 242 236) which encodes phosphinothricin acetyl transferase (PAT). The enzyme PAT confers 30 resistance to herbicidal glutamine synthetase inhibitors such as phosphinothricin. The *pat* gene in p35S/Ac is under the control of the 35S promoter from Cauliflower Mosaic Virus (Odell et al. (1985) *Nature* 313:810-812) and the 3' region of the nopaline synthase gene from the T-DNA of the Ti plasmid of *Agrobacterium tumefaciens*.

The particle bombardment method (Klein et al., (1987) *Nature* 327:70-73) may be 35 used to transfer genes to the callus culture cells. According to this method, gold particles (1 µm in diameter) are coated with DNA using the following technique. Ten µg of plasmid DNAs are added to 50 µL of a suspension of gold particles (60 mg per mL). Calcium chloride (50 µL of a 2.5 M solution) and spermidine free base (20 µL of a 1.0 M solution) are added to the particles. The suspension is vortexed during the addition of these solutions.

After 10 minutes, the tubes are briefly centrifuged (5 sec at 15,000 rpm) and the supernatant removed. The particles are resuspended in 200 μ L of absolute ethanol, centrifuged again and the supernatant removed. The ethanol rinse is performed again and the particles resuspended in a final volume of 30 μ L of ethanol. An aliquot (5 μ L) of the DNA-coated 5 gold particles can be placed in the center of a KaptonTM flying disc (Bio-Rad Labs). The particles are then accelerated into the corn tissue with a BiostaticTM PDS-1000/He (Bio-Rad Instruments, Hercules CA), using a helium pressure of 1000 psi, a gap distance of 0.5 cm and a flying distance of 1.0 cm.

For bombardment, the embryogenic tissue is placed on filter paper over agarose-10 solidified N6 medium. The tissue is arranged as a thin lawn and covered a circular area of about 5 cm in diameter. The petri dish containing the tissue can be placed in the chamber of the PDS-1000/He approximately 8 cm from the stopping screen. The air in the chamber is then evacuated to a vacuum of 28 inches of Hg. The macrocarrier is accelerated with a helium shock wave using a rupture membrane that bursts when the He pressure in the shock 15 tube reaches 1000 psi.

Seven days after bombardment the tissue can be transferred to N6 medium that contains glufosinate (2 mg per liter) and lacks casein or proline. The tissue continues to grow slowly on this medium. After an additional 2 weeks the tissue can be transferred to fresh N6 medium containing glufosinate. After 6 weeks, areas of about 1 cm in diameter 20 of actively growing callus can be identified on some of the plates containing the glufosinate-supplemented medium. These calli may continue to grow when sub-cultured on the selective medium.

Plants can be regenerated from the transgenic callus by first transferring clusters of tissue to N6 medium supplemented with 0.2 mg per liter of 2,4-D. After two weeks the 25 tissue can be transferred to regeneration medium (Fromm et al., (1990) *Bio/Technology* 8:833-839).

EXAMPLE 9

Expression of Chimeric Genes in Dicot Cells

A seed-specific expression cassette composed of the promoter and transcription 30 terminator from the gene encoding the β subunit of the seed storage protein phaseolin from the bean *Phaseolus vulgaris* (Doyle et al. (1986) *J. Biol. Chem.* 261:9228-9238) can be used for expression of the instant amino acid biosynthetic enzymes in transformed soybean. The phaseolin cassette includes about 500 nucleotides upstream (5') from the translation initiation codon and about 1650 nucleotides downstream (3') from the translation stop codon of 35 phaseolin. Between the 5' and 3' regions are the unique restriction endonuclease sites Nco I (which includes the ATG translation initiation codon), Sma I, Kpn I and Xba I. The entire cassette is flanked by Hind III sites.

The cDNA fragment of this gene may be generated by polymerase chain reaction (PCR) of the cDNA clone using appropriate oligonucleotide primers. Cloning sites can be

incorporated into the oligonucleotides to provide proper orientation of the DNA fragment when inserted into the expression vector. Amplification is then performed, and the isolated fragment is inserted into a pUC18 vector carrying the seed expression cassette.

Plant amino acid biosynthetic enzymes are known to be localized in the chloroplasts.

5 Accordingly, for those enzymes (or polypeptides representing part of the instant amino acid biosynthetic enzymes) that lack a chloroplast targeting signal, the DNA fragment to be inserted into the expression vector can be synthesized by PCR with primers encoding a chloroplast targeting signal. For example, a chloroplast transit sequence equivalent to the cts of the small subunit of ribulose 1,5-bisphosphate carboxylase from soybean (Berry-Lowe et 10 al. (1982) *J. Mol. Appl. Genet.* 1:483-498) may be used.

Soybean embryos may then be transformed with the expression vector comprising sequences encoding a plant amino acid biosynthetic enzyme. To induce somatic embryos, cotyledons, 3-5 mm in length dissected from surface sterilized, immature seeds of the soybean cultivar A2872, can be cultured in the light or dark at 26°C on an appropriate agar 15 medium for 6-10 weeks. Somatic embryos which produce secondary embryos are then excised and placed into a suitable liquid medium. After repeated selection for clusters of somatic embryos which multiplied as early, globular staged embryos, the suspensions are maintained as described below.

Soybean embryogenic suspension cultures can be maintained in 35 mL liquid media on a 20 rotary shaker, 150 rpm, at 26°C with fluorescent lights on a 16:8 hour day/night schedule. Cultures are subcultured every two weeks by inoculating approximately 35 mg of tissue into 35 mL of liquid medium.

Soybean embryogenic suspension cultures may then be transformed by the method of particle gun bombardment (Kline et al. (1987) *Nature* (London) 327:70, U.S. Patent 25 No. 4,945,050). A Du Pont Biolistic™ PDS1000/HE instrument (helium retrofit) can be used for these transformations.

A selectable marker gene which can be used to facilitate soybean transformation is a chimeric gene composed of the 35S promoter from Cauliflower Mosaic Virus (Odell et al. (1985) *Nature* 313:810-812), the hygromycin phosphotransferase gene from plasmid 30 pJR225 (from *E. coli*; Gritz et al. (1983) *Gene* 25:179-188) and the 3' region of the nopaline synthase gene from the T-DNA of the Ti plasmid of *Agrobacterium tumefaciens*. The seed expression cassette comprising the phaseolin 5' region, the fragment encoding the biosynthetic enzyme and the phaseolin 3' region can be isolated as a restriction fragment. This fragment can then be inserted into a unique restriction site of the vector carrying the 35 marker gene.

To 50 µL of a 60 mg/mL 1 µm gold particle suspension is added (in order): 5 µL DNA (1 µg/µL), 20 µL spermidine (0.1 M), and 50 µL CaCl₂ (2.5 M). The particle preparation is then agitated for three minutes, spun in a microfuge for 10 seconds and the supernatant removed. The DNA-coated particles are then washed once in 400 µL 70% ethanol and

resuspended in 40 μ L of anhydrous ethanol. The DNA/particle suspension can be sonicated three times for one second each. Five μ L of the DNA-coated gold particles are then loaded on each macro carrier disk.

Approximately 300-400 mg of a two-week-old suspension culture is placed in an empty 60x15 mm petri dish and the residual liquid removed from the tissue with a pipette. For each transformation experiment, approximately 5-10 plates of tissue are normally bombarded. Membrane rupture pressure is set at 1100 psi and the chamber is evacuated to a vacuum of 28 inches mercury. The tissue is placed approximately 3.5 inches away from the retaining screen and bombarded three times. Following bombardment, the tissue can be divided in half and placed back into liquid and cultured as described above.

Five to seven days post bombardment, the liquid media may be exchanged with fresh media, and eleven to twelve days post bombardment with fresh media containing 50 mg/mL hygromycin. This selective media can be refreshed weekly. Seven to eight weeks post bombardment, green, transformed tissue may be observed growing from untransformed, necrotic embryogenic clusters. Isolated green tissue is removed and inoculated into individual flasks to generate new, clonally propagated, transformed embryogenic suspension cultures. Each new line may be treated as an independent transformation event. These suspensions can then be subcultured and maintained as clusters of immature embryos or regenerated into whole plants by maturation and germination of individual somatic embryos.

20 EXAMPLE 10

Analysis of Amino Acid Content of the Seeds of Transformed Plants

To analyze for expression of the chimeric genes in seeds and for the consequences of expression on the amino acid content in the seeds, a seed meal can be prepared by any of a number of suitable methods known to those skilled in the art. The seed meal can be partially or completely defatted, via hexane extraction for example, if desired. Protein extracts can be prepared from the meal and analyzed for enzyme activity. Alternatively the presence of any of the expressed enzymes can be tested for immunologically by methods well-known to those skilled in the art. To measure free amino acid composition of the seeds, free amino acids can be extracted from the meal and analyzed by methods known to those skilled in the art (Bielecki et al. (1966) *Anal. Biochem.* 17:278-293). Amino acid composition can then be determined using any commercially available amino acid analyzer. To measure total amino acid composition of the seeds, meal containing both protein-bound and free amino acids can be acid hydrolyzed to release the protein-bound amino acids and the composition can then be determined using any commercially available amino acid analyzer. Seeds expressing the instant amino acid biosynthetic enzymes and with altered lysine, threonine, methionine, cysteine and/or isoleucine content as compared to the wild type seeds can thus be identified and propagated.

To measure free amino acid composition of the seeds, free amino acids can be extracted from 8-10 milligrams of the seed meal in 1.0 mL of methanol/chloroform/water

mixed in ratio of 12v/5v/3v (MCW) at room temperature. The mixture can be vortexed and then centrifuged in an eppendorf microcentrifuge for about 3 min; approximately 0.8 mL of supernatant is then decanted. To this supernatant, 0.2 mL of chloroform is added followed by 0.3 mL of water. The mixture is then vortexed and centrifuged in an 5 eppendorf microcentrifuge for about 3 min. The upper aqueous phase, approximately 1.0 mL, can then be removed and dried down in a Savant Speed Vac Concentrator. The samples are then hydrolyzed in 6N hydrochloric acid, 0.4% β -mercaptoethanol under nitrogen for 24 h at 110-120°C. Ten percent of the sample can then be analyzed using a Beckman Model 6300 amino acid analyzer using post-column ninhydrin detection.

10 Relative free amino acid levels in the seeds are then compared as ratios of lysine, threonine, methionine, cysteine and/or isoleucine to leucine, thus using leucine as an internal standard.

EXAMPLE 11

Expression of Chimeric Genes in Microbial Cells

15 The cDNAs encoding the instant plant amino acid biosynthetic enzymes can be inserted into the T7 *E. coli* expression vector pET24d (Novagen). Plasmid DNA containing a cDNA may be appropriately digested to release a nucleic acid fragment encoding the enzyme. This fragment may then be purified on a 1% NuSieve GTG™ low melting agarose gel (FMC). Buffer and agarose contain 10 μ g/ml ethidium bromide for visualization of the 20 DNA fragment. The fragment can then be purified from the agarose gel by digestion with GELase™ (Epicentre Technologies) according to the manufacturer's instructions, ethanol precipitated, dried and resuspended in 20 μ L of water. Appropriate oligonucleotide adapters may be ligated to the fragment using T4 DNA ligase (New England Biolabs, Beverly, MA). The fragment containing the ligated adapters can be purified from the excess adapters using 25 low melting agarose as described above. The vector pET24d is digested, dephosphorylated with alkaline phosphatase (NEB) and deproteinized with phenol/chloroform as described above. The prepared vector pET24d and fragment can then be ligated at 16°C for 15 hours followed by transformation into DH5 electrocompetent cells (GIBCO BRL). Transformants can be selected on agar plates containing 2x YT media and 50 μ g/mL kanamycin.

30 Transformants containing gene encoding the enzyme are then screened for the correct orientation with respect to pET24d T7 promoter by restriction enzyme analysis.

Clones in the correct orientation with respect to the T7 promoter can be transformed into BL21(DE3) competent cells (Novagen) and selected on 2x YT agar plates containing 50 μ g/ml kanamycin. A colony arising from this transformation construct can be grown 35 overnight at 30°C in 2x YT media with 50 μ g/mL kanamycin. The culture is then diluted two fold with fresh media, allowed to re-grow for 1 h, and induced by adding isopropyl-thiogalactopyranoside to 1 mM final concentration. Cells are then harvested by centrifugation after 3 h and re-suspended in 50 μ L of 50 mM Tris-HCl at pH 8.0 containing 0.1 mM DTT and 0.2 mM phenyl methylsulfonyl fluoride. A small amount of 1 mm glass

beads can be added and the mixture sonicated 3 times for about 5 seconds each time with a microprobe sonicator. The mixture is centrifuged and the protein concentration of the supernatant determined. One μ g of protein from the soluble fraction of the culture can be separated by SDS-polyacrylamide gel electrophoresis. Gels can be observed for protein
5 bands migrating at the expected molecular weight.

EXAMPLE 12

Evaluating Compounds for Their Ability to Inhibit the Activity of a Plant Amino Acid Biosynthetic Enzyme

The plant amino acid biosynthetic enzymes described herein may be produced using
10 any number of methods known to those skilled in the art. Such methods include, but are not limited to, expression in bacteria as described in Example 6, or expression in eukaryotic cell culture, *in planta*, and using viral expression systems in suitably infected organisms or cell lines. The instant enzymes may be expressed separately as mature proteins, or may be co-expressed in *E. coli* or another suitable expression background. In addition, whether
15 expressed separately or in combination, the instant enzymes may be expressed either as mature forms of the proteins as observed *in vivo* or as fusion proteins by covalent attachment to a variety of enzymes, proteins or affinity tags. Common fusion protein partners include glutathione S-transferase ("GST"), thioredoxin ("Trx"), maltose binding protein, and C- and/or N-terminal hexahistidine polypeptide ("(His)₆"). The fusion proteins may be
20 engineered with a protease recognition site at the fusion point so that fusion partners can be separated by protease digestion to yield intact mature enzymes. Examples of such proteases include thrombin, enterokinase and factor Xa. However, any protease can be used which specifically cleaves the peptide connecting the fusion protein and the biosynthetic enzyme.

Purification of the instant enzymes, if desired, may utilize any number of separation
25 technologies familiar to those skilled in the art of protein purification. Examples of such methods include, but are not limited to, homogenization, filtration, centrifugation, heat denaturation, ammonium sulfate precipitation, desalting, pH precipitation, ion exchange chromatography, hydrophobic interaction chromatography and affinity chromatography, wherein the affinity ligand represents a substrate, substrate analog or inhibitor. When the
30 enzymes are expressed as fusion proteins, the purification protocol may include the use of an affinity resin which is specific for the fusion protein tag attached to the expressed enzyme or an affinity resin containing ligands which are specific for the enzyme. For example, an enzyme may be expressed as a fusion protein coupled to the C-terminus of thioredoxin. In addition, a (His)₆ peptide may be engineered into the N-terminus of the fused thioredoxin
35 moiety to afford additional opportunities for affinity purification. Other suitable affinity resins could be synthesized by linking the appropriate ligands to any suitable resin such as Sepharose-4B. In an alternate embodiment, a thioredoxin fusion protein may be eluted using dithiothreitol; however, elution may be accomplished using other reagents which interact to displace the thioredoxin from the resin. These reagents include β -mercaptoethanol or other

reduced thiol. The eluted fusion protein may be subjected to further purification by traditional means as stated above, if desired. Proteolytic cleavage of the thioredoxin fusion protein and the biosynthetic enzyme may be accomplished after the fusion protein is purified or while the protein is still bound to the ThioBond™ affinity resin or other resin.

5 Crude, partially purified or purified enzyme, either alone or as a fusion protein, may be utilized in assays for the evaluation of compounds for their ability to inhibit enzymatic activation of the plant amino acid biosynthetic enzymes disclosed herein. Assays may be conducted under well known experimental conditions which permit optimal enzymatic activity. Examples of assays for many of these enzymes can be found in *Methods in*

10 *Enzymology* Vol. V, (Colowick and Kaplan eds.) Academic Press, New York or *Methods in Enzymology* Vol. XVII, (Tabor and Tabor eds.) Academic Press, New York. Specific examples may be found in the following references, each of which is incorporated herein by reference: dihydrodipicolinate reductase may be assayed as described in Farkas et al. (1965) *J. Biol. Chem.* 240: 4717-4722, or Cremer et al. (1988) *J. Gen. Microbiol.* 134:3221-3229;

15 diaminopimelate epimerase may be assayed as described in Work (1962) in *Methods in Enzymology* Vol. V, (Colowick and Kaplan eds.) 858-864, Academic Press, New York; threonine synthase may be assayed as described in Giovanelli et al. (1984) *Plant Physiol.* 76: 285-292 or Curien et al. (1996) *FEBS Lett.* 390: 85-90; threonine deaminase may be assayed as described in Tomova et al. (1968) *Biochemistry (USSR)* 33: 200-208 or Dougall (1970)

20 *Phytochemistry* 9: 959-964; and S-adenosylmethionine synthetase may be assayed as described in Mudd (1960) *Biochim. Biophys. Acta* 38:354-355 or Boerjan et al. (1994) *Plant Cell* 6:1401-1414.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

- (A) ADDRESSEE: E. I. DU PONT DE NEMOURS AND COMPANY
- (B) STREET: 1007 MARKET STREET
- (C) CITY: WILMINGTON
- (D) STATE: DELAWARE
- (E) COUNTRY: USA
- (F) ZIP: 19898
- (G) TELEPHONE: 302-992-4926
- (H) TELEFAX: 302-773-0164
- (I) TELEX: 6717325

(ii) TITLE OF INVENTION: PLANT AMINO ACID BIOSYNTHETIC ENZYMES

(iii) NUMBER OF SEQUENCES: 43

(iv) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: DISKETTE, 3.50 INCH
- (B) COMPUTER: IBM PC COMPATIBLE
- (C) OPERATING SYSTEM: MICROSOFT WINDOWS 95
- (D) SOFTWARE: MICROSOFT WORD VERSION 7.0A

(v) CURRENT APPLICATION DATA:

- (A) APPLICATION NUMBER:
- (B) FILING DATE:
- (C) CLASSIFICATION:

(vi) PRIOR APPLICATION DATA:

- (A) APPLICATION NUMBER: 60/048,771
- (B) FILING DATE: JUNE 6, 1997

(vii) ATTORNEY/AGENT INFORMATION:

- (A) NAME: MAJARIAN, WILLIAM R.
- (B) REGISTRATION NUMBER: 41,173
- (C) REFERENCE/DOCKET NUMBER: BB-1087

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 908 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: csiln.pk0042.a3
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

ACGCAGGGACA	GATAAGTGGC	ATGGACGAGC	CGCTGGAGAT	CCCTGTGCTG	AACGACCTCA	60
CCATGGTTCT	GGGCTCCATA	GCCGAGTCGA	GAGCAACCGG	CGTGGTGGTC	GACTTCAGCG	120
AGCCTTCAGC	TGTTTACGAC	AATGTCAAGC	AGGCAGCGGC	GTTTGGTCTG	AGCAGCGTCG	180
TCTACGTTCC	GAAAATCGAG	CTAGAGACAG	TGACTGAAC	GTCAGCGTTC	TGCGAGAAAGG	240
CAAGCGGCTG	CTTGGTTGCG	CCAACGCTGT	CGATTGGGTC	CGTGCTCCTT	CAGCAAGCGG	300
CTATACAGGC	CTCGTTCCAC	TACAGCAACG	TTGAGATTGT	GGAATCGAGA	CCAAACCCAT	360
CGGATCTTCC	ATCGCAAGAT	GCAATCCAGA	TTGCAAACAA	CATATCAGAC	CTTGGTCAGA	420
TATACAACAG	GGAAGATATG	GATTCCAGCA	GTCCAGCCAG	AGGCCAGCTG	CTCGGGGAAG	480
ACGGAGTGCG	CGTGCACAGC	ATGGTTCTCC	CTGGTCTCGT	CTCCAGCAGC	TCGATCAACT	540
TCTCTGGCCC	AGGAGAGATG	TACACCTTAC	GGCATGACGT	TGCGAATGTT	CAGTGCCTGA	600
TGCCAGGACT	GATCCTGGCG	ATACGGAAGG	TGGTGCGGTT	CAAGAACTTG	ATTATGGGC	660
TAGAGAAAGTT	CTTGTAGTGA	ACAACAAACA	ACCAATGCAA	AACATCGACA	GGCAACAGGC	720
AAGGCAGATA	TCATCTGACG	TCGCAACAAAC	CAAAACGACA	GAGATTGGA	AAATAAAGGC	780
TGCCACAGAAG	ACGTCTGGGG	TTTTGTGTGC	ACCAGGCTGC	GCAGAGAACG	TCTGTCATT	840
TGTGTGCACC	ACTACGGCAC	TACCTGCTGA	GCGCGATTT	TATAAAAAAG	GCATGGGAGG	900
GAGATCAT						908

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 224 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: csiln.pk0042.a3
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Ala	Gly	Gln	Ile	Ser	Gly	Met	Asp	Glu	Pro	Leu	Glu	Ile	Pro	Val	Leu
1				5				10				15			

Asn	Asp	Leu	Thr	Met	Val	Leu	Gly	Ser	Ile	Ala	Gln	Ser	Arg	Ala	Thr
20															30
Gly	Val	Val	Val	Asp	Phe	Ser	Glu	Pro	Ser	Ala	Val	Tyr	Asp	Asn	Val
35															45
Lys	Gln	Ala	Ala	Ala	Phe	Gly	Leu	Ser	Ser	Val	Val	Tyr	Val	Pro	Lys
50															60
Ile	Glu	Leu	Glu	Thr	Val	Thr	Glu	Leu	Ser	Ala	Phe	Cys	Glu	Lys	Ala
65															80
Ser	Gly	Cys	Leu	Val	Ala	Pro	Thr	Leu	Ser	Ile	Gly	Ser	Val	Leu	Leu
															95
Gln	Gln	Ala	Ala	Ile	Gln	Ala	Ser	Phe	His	Tyr	Ser	Asn	Val	Glu	Ile
100															110
Val	Glu	Ser	Arg	Pro	Asn	Pro	Ser	Asp	Leu	Pro	Ser	Gln	Asp	Ala	Ile
115															125
Gln	Ile	Ala	Asn	Asn	Ile	Ser	Asp	Leu	Gly	Gln	Ile	Tyr	Asn	Arg	Glu
130															140
Asp	Met	Asp	Ser	Ser	Ser	Pro	Ala	Arg	Gly	Gln	Leu	Leu	Gly	Glu	Asp
145															160
Gly	Val	Arg	Val	His	Ser	Met	Val	Leu	Pro	Gly	Leu	Val	Ser	Ser	Thr
165															175
Ser	Ile	Asn	Phe	Ser	Gly	Pro	Gly	Glu	Met	Tyr	Thr	Leu	Arg	His	Asp
180															190
Val	Ala	Asn	Val	Gln	Cys	Leu	Met	Pro	Gly	Leu	Ile	Leu	Ala	Ile	Arg
195															205
Lys	Val	Val	Arg	Phe	Lys	Asn	Leu	Ile	Tyr	Gly	Leu	Glu	Lys	Phe	Leu
210															220

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 339 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vii) IMMEDIATE SOURCE:
 (B) CLONE: rls2.pk0017.d3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

AAGATTGGCA	GGAGAAATGC	AGCAAAGGTC	CTCTGCTCAA	CGCAGATGCC	GCCATCTCAG	60
AGCACAATCA	AGGTTGTTAT	CATTGGGGCG	ACAAAAGAGA	TTGGAAGAAC	GGCAATAGCG	120
GCAGTAAGTA	AAGCAAGGGG	AATGGAGCTT	GCAGGGGCCA	TAGATTCTCA	GTGTATAGGC	180
CTAGATGCAG	GAGAGATAAG	TGGCATGGGA	AGAACCCCTGG	AAATTCCGGT	GCTCAATGAT	240
CTCACAATGG	TTCTGGGCTC	AATTGCACAA	ACCAGAGCAA	CTGGAGTGTT	CGTTGATTT	300
AGTGAACCTT	CAACTGT'TTA	TGATAATGTC	AAACAGGCA			339

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 113 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: not relevant
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vii) IMMEDIATE SOURCE:
 (B) CLONE: rls2.pk0017.d3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Lys Ile Gly Arg Arg Asn Ala Ala Lys Val Leu Cys Ser Thr Gln Met
 1 5 10 15

Pro Pro Ser Gln Ser Thr Ile Lys Val Val Ile Ile Gly Ala Thr Lys
 20 25 30

Glu Ile Gly Arg Thr Ala Ile Ala Ala Val Ser Lys Ala Arg Gly Met
 35 40 45

Glu Leu Ala Gly Ala Ile Asp Ser Gln Cys Ile Gly Leu Asp Ala Gly
 50 55 60

Glu Ile Ser Gly Met Gly Arg Thr Leu Glu Ile Pro Val Leu Asn Asp
 65 70 75 80

Leu Thr Met Val Leu Gly Ser Ile Ala Gln Thr Arg Ala Thr Gly Val
 85 90 95

Val Val Asp Phe Ser Glu Pro Ser Thr Val Tyr Asp Asn Val Lys Gln
 100 105 110

Ala

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 275 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: not relevant
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vi) ORIGINAL SOURCE:
 (A) ORGANISM: Synechocystis sp

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met Ala Asn Gln Asp Leu Ile Pro Val Val Val Asn Gly Ala Ala Gly
 1 5 10 15

Lys Met Gly Arg Glu Val Ile Lys Ala Val Ala Gln Ala Pro Asp Leu
 20 25 30

Gln Leu Val Gly Ala Val Asp His Asn Pro Ser Leu Gln Gly Gln Asp
 35 40 45

Ile Gly Glu Val Val Gly Ile Ala Pro Leu Glu Val Pro Val Leu Ala
 50 55 60

Asp Leu Gln Ser Val Leu Val Leu Ala Thr Gln Glu Lys Ile Gln Gly
 65 70 75 80
 Val Met Val Asp Phe Thr His Pro Ser Gly Val Tyr Asp Asn Val Arg
 85 90 95
 Ser Ala Ile Ala Tyr Gly Val Arg Pro Val Val Gly Thr Thr Gly Leu
 100 105 110
 Ser Glu Gln Gln Ile Gln Asp Leu Gly Asp Phe Ala Glu Lys Ala Ser
 115 120 125
 Thr Gly Cys Leu Ile Ala Pro Asn Phe Ala Ile Gly Val Leu Leu Met
 130 135 140
 Gln Gln Ala Ala Val Gln Ala Cys Gln Tyr Phe Asp His Val Glu Ile
 145 150 155 160
 Ile Glu Leu His His Asn Gln Lys Ala Asp Ala Pro Ser Gly Thr Ala
 165 170 175
 Ile Lys Thr Ala Gln Met Leu Ala Glu Met Gly Lys Thr Phe Asn Pro
 180 185 190
 Pro Ala Val Glu Glu Lys Glu Thr Ile Ala Gly Ala Lys Gly Gly Leu
 195 200 205
 Gly Pro Gly Gln Ile Pro Ile His Ser Ile Arg Leu Pro Gly Leu Ile
 210 215 220
 Ala His Gln Glu Val Leu Phe Gly Ser Pro Gly Gln Leu Tyr Thr Ile
 225 230 235 240
 Arg His Asp Thr Thr Asp Arg Ala Cys Tyr Met Pro Gly Val Leu Leu
 245 250 255
 Gly Ile Arg Lys Val Val Glu Leu Lys Gly Leu Val Tyr Gly Leu Glu
 260 265 270
 Lys Leu Leu
 275

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1012 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: chp2.pk0008.h4
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

TATTGCCAGA GATGTGTGGT AATGGAGTCC GTTGCTTCGC TCGGTTTATA GCCGAGAGTTG	60
AAAATCTGCA GGGGACAAAT AGATTCACTA TTCATACTGG TGCTGGAAAG ATCGTTCCCTG	120
AAATACAAAG TGATGGCAG GTAAAGGTTG ATATGGCGA GCCTATCCTT TCTGGACTAG	180
ACATCCCCAC AAAACTGCTA GCTACCAAGA ACAAAAGCTGT TGTCAAGCT GAATTGGCAG	240
TTGAGGGCTT AACATGGCAT GTCACATGTG TTAGCATGGG AAACCCCTCAC TGTGTCACAT	300

TTGGTGCAAA	TGAGTTAAAG	GTATTGCAGG	TCGACGATTT	AAAACCTTGC	GAAATTGGGC	360
CTAAATTGAA	GCATCATGAA	ATGTTCTG	CTCGCACAAA	CACAGAATTG	GTACAGGTTT	420
TGTCTCGCTC	ACACCTCAAA	ATGCGGGTCT	GGGAACGTGG	TGCTGGAGCA	ACTCTTGCCT	480
GTGGTACTGG	TGCTTGTGCA	GTGGTTGTTG	CAGCTGTTCT	TGAGGGTCGA	GCTGAGCGGA	540
AATGTGTAGT	TGATTTGCCT	GGCGGGCCAT	TGGAAATTGA	GTGGAGGGAG	GATGACAATC	600
ATGTTTACAT	GACTGGTCCT	GCAGAGGTCG	TCTTTATGG	ATCTGTTGTT	CACTAGGTAC	660
TGGGGACCAA	GATAGAAGGG	TTGGCTGCCA	CTCAGAGCTT	GTGAGATTGG	TTATAGTATC	720
CATGAAACAG	AGTGTCTGG	TACCACTACA	CTTGTTAGA	TATTCTTAAT	TATGATTGCT	780
TGATTTGGGT	AGCMGTAGAG	GCTTCCTTT	GAAGCATTCT	AGTGTTCMCC	TTTTGTACTC	840
CTTTAGTTTG	TCAGGTTGA	ACACTACATG	GGTAACATGT	CYTCCCACC	ATTTCYGTT	900
TCTTTCTTT	GTAAGTGAAC	GCCAATGCAG	TTTTAGTATT	GTTTCTATA	GATTTGTCTT	960
GATGCACTGG	GCTTACTACT	TATTTCTGG	TATGAATGCT	GCCTATTCC	TG	1012

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 217 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: not relevant
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vii) IMMEDIATE SOURCE:
 (B) CLONE: chp2.pk0008.h4

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Leu	Pro	Glu	Met	Cys	Gly	Asn	Gly	Val	Arg	Cys	Phe	Ala	Arg	Phe	Ile
1				5				10					15		
Ala	Glu	Ile	Glu	Asn	Leu	Gln	Gly	Thr	Asn	Arg	Phe	Thr	Ile	His	Thr
					20			25					30		
Gly	Ala	Gly	Lys	Ile	Val	Pro	Glu	Ile	Gln	Ser	Asp	Gly	Gln	Val	Lys
			35			40			45						
Val	Asp	Met	Gly	Glu	Pro	Ile	Leu	Ser	Gly	Leu	Asp	Ile	Pro	Thr	Lys
	50				55				60						
Leu	Leu	Ala	Thr	Lys	Asn	Lys	Ala	Val	Val	Gln	Ala	Glu	Leu	Ala	Val
	65				70			75		80					
Glu	Gly	Leu	Thr	Trp	His	Val	Thr	Cys	Val	Ser	Met	Gly	Asn	Pro	His
			85			90				95					
Cys	Val	Thr	Phe	Gly	Ala	Asn	Glu	Leu	Lys	Val	Leu	Gln	Val	Asp	Asp
	100					105				110					
Leu	Lys	Leu	Ser	Glu	Ile	Gly	Pro	Lys	Phe	Glu	His	His	Glu	Met	Phe
	115					120				125					
Pro	Ala	Arg	Thr	Asn	Thr	Glu	Phe	Val	Gln	Val	Leu	Ser	Arg	Ser	His
	130					135				140					

Leu Lys Met Arg Val Trp Glu Arg Gly Ala Gly Ala Thr Leu Ala Cys
 145 150 155 160

Gly Thr Gly Ala Cys Ala Val Val Val Ala Ala Val Leu Glu Gly Arg
 165 170 175

Ala Glu Arg Lys Cys Val Val Asp Leu Pro Gly Gly Pro Leu Glu Ile
 180 185 190

Glu Trp Arg Glu Asp Asp Asn His Val Tyr Met Thr Gly Pro Ala Glu
 195 200 205

Val Val Phe Tyr Gly Ser Val Val His
 210 215

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 481 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: rls48.pk0036.h10
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

TGTATCCGGC	GCCGACGGTG	TGATCTTCGT	CATGCCGGGG	GTCAATGGCG	CGGACTACAC	60
CATGAGGATC	TTCAACTCGG	ACGGCAGTGA	GCGGGAGATG	TGTGGCAATG	GAGTCCGTTG	120
CTTGCCCCGG	TTTATAGCTG	AGCTTGAAAAA	CCTACAGGGA	ACACATAGCT	TCAAATTCA	180
CACTGGCGCT	GGGCTAATCA	TTCTGAAAT	ACAAAATGAT	GGCAAGGTA	AGGTTGATAT	240
GGGCCAGCCC	ATTCTCTCTG	GACCAGATAT	TCCAACAAAAA	CTGCCATCCA	CCAAGAATGA	300
AGCCGTTGTC	CAAGCTGATT	TGGCAGTTG	ATGGCTCAAC	ATGGCAAGTA	ACCTGTGTTA	360
GCATGGCAA	TCCACATTGT	GTCACATTTG	GCACAAAGGA	GCTCAAGGTT	TTGCATGTTG	420
ATGATTAAG	CTTAATGATA	TTGGGGCCTA	AATTCAAGCAT	CATGAAATGT	TCCTGCCCA	480
C						481

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 85 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: rls48.pk0036.h10
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Val Ser Gly Ala Asp Gly Val Ile Phe Val Met Pro Gly Val Asn Gly			
1	5	10	15

Ala Asp Tyr Thr Met Arg Ile Phe Asn Ser Asp Gly Ser Glu Pro Glu
 20 25 30

Met Cys Gly Asn Gly Val Arg Cys Phe Ala Arg Phe Ile Ala Glu Leu
 35 40 45

Glu Asn Leu Gln Gly Thr His Ser Phe Lys Ile His Thr Gly Ala Gly
 50 55 60

Leu Ile Ile Pro Glu Ile Gln Asn Asp Gly Lys Val Lys Val Asp Met
 65 70 75 80

Gly Gln Pro Ile Leu
 85

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1301 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

ATCCCTTATT AAGCAGGGGT TTTCGCGCGC GAGACGGTGA CACTGGCAGA GTGGAATTTC	60
CGCCGCCATT CGAACGCTACA GCGATGGCCA TAACCGCCAC CATTTCGTT CCCCTCACAT	120
CCCCCAGTCG CCGCACTCTC ACCTCCGTCA ATAGCCTCTC TCCCCTTTCT ACCCGATCCA	180
CTTTGCCAC ACCGCAACGC ACTTTCAAAT ACCCTAATTG GCGCCTCGTC GTGTCTCCA	240
TGAGCACCAG AACAGCCGTC AAAACTTCAT CCGCCTCCTT CCTCAACCGC AAGGAGTCCG	300
GCTTCCTCCA TTTCGCCAAG TACCACGGCC TCGGAAACGA CTTCGTTTG ATTGACAATA	360
GAGACTCCTC CGAGCCAAG ATCAGTGCTG AGAAAGCGGT GCAACTGTGT GATCGGAAC	420
TCGGCGTTGG AGCTGACGGA GTTATCTTTG TCTTGCTGG CATCAGTGGC ACCGATTATA	480
CCATGAGGAT TTTAACTCT GATGGTAGTG AGCCTGAGAT GTGTGGCAAT GGAGTTCGAT	540
GCTTGCCAA ATTTGTTCT CAGCTTGAGA ATTTACATGG GAGGCATAGT TTTACCATTC	600
ATACTGGTGC TGGTCTGATT ATTCCTGAAG TCTTGAGGA TGGAAATGTC AGAGTTGATA	660
TGGGGGAGCC AGTTCTAAA GCCTTGGATG TGCCTACTAA ATTACCTGCA AATAAGGATA	720
ATGCTGTTGT TAAATCACAG CTAGTTGTAG ATGGAGTTAT TTGGCATGTG ACCTGTGTTA	780
GCATGGGAA TCCACACTGT GTAACTTCA GTAGAGAAGG AAGCCAGAAT TTGCTTGTG	840
ATGAATTGAA GCTAGCAGAA ATTGGGCCAA AATTGAAACA TCATGAGGTG TTCCCTGCAC	900
GAACTAACAC AGAGTTGTG CAAGTATTAT CTAACCTCTCA CTTGAAAATG CGTGTGTTGG	960
AGCGGGGAGC AGGAGCAACC CTAGCCTGTG GAACTGGAGC TTGTGCTACT GTTGTGTCAG	1020
CAGTTCTTGA GGGTCGTGCT GGGAGGAATT GCACGGTTGA TCTACCTGGA GGGCCTCTTC	1080
AGATTGAGTG GAGGGAGGAA GATAATCATG TTTATATGAC AGGCTCAGCC GATGTAGTTT	1140

ATTATGGTTC TTTGCCCTT TGATATGTTG CCCCCATTGT TAAACCCAAT ATGGAATTAG	1200
GAATTGGTGA ATAATATTTG TATGAGAGGT GGACTTCTG CTTGTTCCCTA ATATTTGCC	1260
ACGTCTTAT AAAAAAAA AAAAAAAA AAAAAAAA A	1301

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 359 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Met Ala Ile Thr Ala Thr Ile Ser Val Pro Leu Thr Ser Pro Ser Arg	
1 5 10 15	
Arg Thr Leu Thr Ser Val Asn Ser Leu Ser Pro Leu Ser Thr Arg Ser	
20 25 30	
Thr Leu Pro Thr Pro Gln Arg Thr Phe Lys Tyr Pro Asn Ser Arg Leu	
35 40 45	
Val Val Ser Ser Met Ser Thr Glu Thr Ala Val Lys Thr Ser Ser Ala	
50 55 60	
Ser Phe Leu Asn Arg Lys Glu Ser Gly Phe Leu His Phe Ala Lys Tyr	
65 70 75 80	
His Gly Leu Gly Asn Asp Phe Val Leu Ile Asp Asn Arg Asp Ser Ser	
85 90 95	
Glu Pro Lys Ile Ser Ala Glu Lys Ala Val Gln Leu Cys Asp Arg Asn	
100 105 110	
Phe Gly Val Gly Ala Asp Gly Val Ile Phe Val Leu Pro Gly Ile Ser	
115 120 125	
Gly Thr Asp Tyr Thr Met Arg Ile Phe Asn Ser Asp Gly Ser Glu Pro	
130 135 140	
Glu Met Cys Gly Asn Gly Val Arg Cys Phe Ala Lys Phe Val Ser Gln	
145 150 155 160	
Leu Glu Asn Leu His Gly Arg His Ser Phe Thr Ile His Thr Gly Ala	
165 170 175	
Gly Leu Ile Ile Pro Glu Val Leu Glu Asp Gly Asn Val Arg Val Asp	
180 185 190	
Met Gly Glu Pro Val Leu Lys Ala Leu Asp Val Pro Thr Lys Leu Pro	
195 200 205	
Ala Asn Lys Asp Asn Ala Val Val Lys Ser Gln Leu Val Val Asp Gly	
210 215 220	
Val Ile Trp His Val Thr Cys Val Ser Met Gly Asn Pro His Cys Val	
225 230 235 240	
Thr Phe Ser Arg Glu Gly Ser Gln Asn Leu Leu Val Asp Glu Leu Lys	
245 250 255	

Leu Ala Glu Ile Gly Pro Lys Phe Glu His His Glu Val Phe Pro Ala
 260 265 270
 Arg Thr Asn Thr Glu Phe Val Gln Val Leu Ser Asn Ser His Leu Lys
 275 280 285
 Met Arg Val Trp Glu Arg Gly Ala Gly Ala Thr Leu Ala Cys Gly Thr
 290 295 300
 Gly Ala Cys Ala Thr Val Val Ala Ala Val Leu Glu Gly Arg Ala Gly
 305 310 315 320
 Arg Asn Cys Thr Val Asp Leu Pro Gly Gly Pro Leu Gln Ile Glu Trp
 325 330 335
 Arg Glu Glu Asp Asn His Val Tyr Met Thr Gly Ser Ala Asp Val Val
 340 345 350
 Tyr Tyr Gly Ser Leu Pro Leu
 355

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 602 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: wlm24.pk0030.g4
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

CTCCACCGCC	CCCTCCTCGG	GCGGTCGCCT	CCTCCGTCCG	TTCTGTGGGA	ATCCGCGCCC	60
CCGCCGCGCC	GTCGCCTCGA	TGGCCGTGTC	CGCTCCCAAG	TCGCCAGCCG	CCGCCTCGTT	120
CCTCGAGCGC	CGCGAGTCCG	AGCGCGCGCT	CCACTTCGTG	AAGTACCAGG	GCCTCGGCAA	180
CGACTTCATA	ATGGTCGACA	ACAGGGATTC	GGCCGTACCG	AAGGTGACAC	CGGAGGAGGC	240
GGCGAAGCTA	TGCGACCGAA	ACTTTGGGTA	TTGGGTGCTG	ATGGCGTCAT	CTTCGTCTG	300
CCGGGGGTCA	ACGGCGCGGA	CTACACTATG	AGGATATTCA	ACTCCGATGG	CAGCAACCAG	360
AATGTNTGGN	ATGGATTCTGT	TGCTTGCTCG	CTTTATACGG	AGTTGAAATC	TACANGGAAA	420
CATACTTCAA	AACAANAGGG	GGCTGGATTAA	ATATCCTGAA	ATANANACAT	GNAAGTTANG	480
TNATATGGGC	AACAATCTTA	TGGCANATT	CANAAAATGC	ATCACAAAGAT	AACTTNTAAA	540
ACGATTGAAT	TAGGCAANAG	AANTACCGTT	ATAGGAACCC	ATGAANCTTG	TNAAATTAAG	600
GT						602

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 80 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: wlm24.pk0030.g4
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Ala Leu His Phe Val Lys Tyr Gln Gly Leu Gly Asn Asp Phe Ile Met
 1 5 10 15

Val Asp Asn Arg Asp Ser Ala Val Pro Lys Val Thr Pro Glu Glu Ala
 20 25 30

Ala Lys Leu Cys Asp Arg Asn Phe Gly Xaa Gly Ala Asp Gly Val Ile
 35 40 45

Phe Val Leu Pro Gly Val Asn Gly Ala Asp Tyr Thr Met Arg Ile Phe
 50 55 60

Asn Ser Asp Gly Ser Asn Arg Asn Val Trp Xaa Gly Phe Val Ala Cys
 65 70 75 80

(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 279 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Synechocystis sp
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Ala Leu Ser Phe Ser Lys Tyr His Gly Leu Gly Asn Asp Phe Ile
 1 5 10 15

Leu Val Asp Asn Arg Gln Ser Thr Glu Pro Cys Leu Thr Pro Asp Gln
 20 25 30

Ala Gln Gln Leu Cys Asp Arg His Phe Gly Ile Gly Ala Asp Gly Val
 35 40 45

Ile Phe Ala Leu Pro Gly Gln Gly Gly Thr Asp Tyr Thr Met Arg Ile
 50 55 60

Phe Asn Ser Asp Gly Ser Glu Pro Glu Met Cys Gly Asn Gly Ile Arg
 65 70 75 80

Cys Leu Ala Lys Phe Leu Ala Asp Leu Glu Gly Val Glu Glu Lys Thr
 85 90 95

Tyr Arg Ile His Thr Leu Ala Gly Val Ile Thr Pro Gln Leu Leu Ala
 100 105 110

Asp Gly Gln Val Lys Val Asp Met Gly Glu Pro Gln Leu Leu Ala Glu
 115 120 125

Leu Ile Pro Thr Thr Leu Ala Pro Ala Gly Glu Lys Val Val Asp Leu
 130 135 140

Pro Leu Ala Val Ala Gly Gln Thr Trp Ala Val Thr Cys Val Ser Met
 145 150 155 160

Gly Asn Pro His Cys Leu Thr Phe Val Asp Asp Val Asp Ser Leu Asn
 165 170 175
 Leu Thr Glu Ile Gly Pro Leu Phe Glu His His Pro Gln Phe Ser Gln
 180 185 190
 Arg Thr Asn Thr Glu Phe Ile Gln Val Leu Gly Ser Asp Arg Leu Lys
 195 200 205
 Met Arg Val Trp Glu Arg Gly Ala Gly Ile Thr Leu Ala Cys Gly Thr
 210 215 220
 Gly Ala Cys Ala Thr Val Val Ala Ala Val Leu Thr Gly Arg Gly Asp
 225 230 235 240
 Arg Arg Cys Thr Val Glu Leu Pro Gly Gly Asn Leu Glu Ile Glu Trp
 245 250 255
 Ser Ala Gln Asp Asn Arg Leu Tyr Met Thr Gly Pro Ala Gln Arg Val
 260 265 270
 Phe Ser Gly Gln Ala Glu Ile
 275

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1160 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: cc2.pk0031.c9
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GTGGGCTGCG CGTCCACGGG AGACACCTCC	GGCGCGCTCT CGGCCTACTG CGCAGCCGCG	60
GGAATCCCCG CCATCGTGT	CTGCCAGCG GACCGCATCT CGCTGCAGCA	120
CCGATGCCA ACGGCCAC CGTGCTCTCT	CTAGACACTG ATTTGATGG CTGCATGCGG	180
CTCATTGCG AGGTCACTGC	AGAGCTGCCA ATCTACCTTG CCAATTGCT	240
CGCTTGAGG GGCAGAACAC	AGCGGCCATC GAGATATTGC AGCAGTTCAA	300
CCAGATTGGG TCATTGTTCC	AGGAGGCAAT CTTGGGAATA TCTATGCATT	360
TTTGAGATGT GCCGCGTTCT	TGGACTTGT GATCGCGTGC CACGGCTTGT	420
GCTGAAATG CAAATCCATT	GTACCGGTAC TACAAGTCAG GTGGACTGA	480
CAAATGCCG AGACTACATT	ATACAGATTG GTGATCCTGT ATCTGTTGAC	540
CGTGGGTGG TCGCGCTGAA	GGCCACTGAC GGTATTGTGG AGGAGGCTAC	600
CTAATGGATG CAACGGCGCT	TGCTGACCGC ACTGGGATGT TTGCTTGCCC	660
GTTGCACTTG CTGCTTGT	TAAGCTTCAG GGTCAGCGTA TAATTGGCCC	720
ACTGTGGTTG TTAGCACAGC	TCATGGGCTG AAGTTCACGC AGTCAGAGAT	780

GACAAAAACA TCAAAGACAT GGTTGCCAG TATGCTAATC CACCGATCAG TGTGAAGGCT	840
GACTTTGGTT CTGTGATGGA TGTTCTCCAG AAAAATCTCA ATGGTAAGAT ATAAAGTTAT	900
ATGATTAATT AACCCCTCCAA ACTGTTTTT TTTGTTTTT CGTTCCAGGA ATTTTATTCC	960
TGAGTCTTTC AACTTGTTT GGTGAACATG GTATGGTGCT AAAATCTAGA CCTAATACCT	1020
TGTAGTACTA GTTCTGGAGG CTCTTTGGT TGTAGGTCGA AGTGGATAGA GCTGTTCCCT	1080
GTACTTTATC TGTTTCAATGT AATATGAATA ATAAATTATG GTCTAAATAT TTGAATAAAA	1140
AATCGTTTGG AATGACCCAC	1160

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 297 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: not relevant
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vii) IMMEDIATE SOURCE:
 (B) CLONE: cc2.pk0031.c9

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Val Gly Cys Ala Ser Thr Gly Asp Thr Ser Ala Ala Leu Ser Ala Tyr	
1 5 10 15	
Cys Ala Ala Ala Gly Ile Pro Ala Ile Val Phe Leu Pro Ala Asp Arg	
20 25 30	
Ile Ser Leu Gln Gln Leu Ile Gln Pro Ile Ala Asn Gly Ala Thr Val	
35 40 45	
Leu Ser Leu Asp Thr Asp Phe Asp Gly Cys Met Arg Leu Ile Arg Glu	
50 55 60	
Val Thr Ala Glu Leu Pro Ile Tyr Leu Ala Asn Ser Leu Asn Pro Leu	
65 70 75 80	
Arg Leu Glu Gly Gln Lys Thr Ala Ala Ile Glu Ile Leu Gln Gln Phe	
85 90 95	
Asn Trp Gln Val Pro Asp Trp Val Ile Val Pro Gly Gly Asn Leu Gly	
100 105 110	
Asn Ile Tyr Ala Phe Tyr Lys Gly Phe Glu Met Cys Arg Val Leu Gly	
115 120 125	
Leu Val Asp Arg Val Pro Arg Leu Val Cys Ala Gln Ala Ala Asn Ala	
130 135 140	
Asn Pro Leu Tyr Arg Tyr Tyr Lys Ser Gly Trp Thr Glu Phe Glu Pro	
145 150 155 160	
Gln Thr Ala Glu Thr Thr Phe Ala Ser Ala Ile Gln Ile Gly Asp Pro	
165 170 175	
Val Ser Val Asp Arg Ala Val Val Ala Leu Lys Ala Thr Asp Gly Ile	
180 185 190	

Val Glu Glu Ala Thr Glu Glu Glu Leu Met Asp Ala Thr Ala Leu Ala
 195 200 205

Asp Arg Thr Gly Met Phe Ala Cys Pro His Thr Gly Val Ala Leu Ala
 210 215 220

Ala Leu Phe Lys Leu Gln Gly Gln Arg Ile Ile Gly Pro Asn Asp Arg
 225 230 235 240

Thr Val Val Val Ser Thr Ala His Gly Leu Lys Phe Thr Gln Ser Lys
 245 250 255

Ile Asp Tyr His Asp Lys Asn Ile Lys Asp Met Val Cys Gln Tyr Ala
 260 265 270

Asn Pro Pro Ile Ser Val Lys Ala Asp Phe Gly Ser Val Met Asp Val
 275 280 285

Leu Gln Lys Asn Leu Asn Gly Lys Ile
 290 295

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 325 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: csl.pk0058.g5
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

ATGGCTTGCA AGTACTCCAA CCCGCCTGTG AGCGTGAAGG CTGACTTGG CGCCGTGATG	60
GATGTGCTGA AGAAGAGGCT CAAGGGCAAG CTCTGAGCGC CTGTGCCTGG CTAATGCAAT	120
CAACTGATTG GAATGCAGTG GTTTCGTCGG TATCGGGGGG TCTTTAGGC TTCAGAAATT	180
CTGTCTGGGT TAGACTATTT GTTTGTGGAG TTTAGCAGGA GAATGGCTAT CTCTCCTGCA	240
AGACTGGCGC TCTTCCTGT GCTACGAATG TGTTACCATG GATAATAAGT GTAGTCGCTG	300
TCGGATTGAA TAATCAAAAA AAAAN	325

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 31 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: csl.pk0058.g5
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Met Ala Cys Lys Tyr Ser Asn Pro Pro Val Ser Val Lys Ala Asp Phe	
1 5 10 15	

Gly Ala Val Met Asp Val Leu Lys Lys Arg Leu Lys Gly Lys Leu
 20 25 30

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 528 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vii) IMMEDIATE SOURCE:
 (B) CLONE: rls72.pk0018.e7

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

ACACCCAACA CGCAGACTTG ACAGATTCTG CTACTACAAA TCCTGCATAT TTAACAGCGC	60
TGCAACTCGA CGATGGAGAA CGGTGCTGCA ACCAACGGGG CGTCGGAGAA GTCGCACTCT	120
CCTTCACAGA CCTACCTCTC CACAAGGGGA GACGATTATG GGCTCTCATT CGAGACCGTC	180
GTCCTCAAAG GTCTTGCAGC TGACGGGGGT CTTTCTCTGC CGAGAGGAAGT GCGCGCGCA	240
ACCGAGTGGC AAAGCTGGAA AGACCTGCC TACACCGAGC TTGCCGTCAA GGTTCTCAGC	300
TTGTACATCT CCCCCGCCGA GGTGCCGACG GAAGACCTCA GGGCGCTCGT CGAGCGCAGC	360
TACTCGACCT TCCGATCCAA GGAGGTTGTG CCGCTGGTGA AGCTGGAGGA CAACCTTCAC	420
CTGCTGGAGC TATTCCACGG CCCCAACTAC TCGTTCAAGG ACTGCGCGCT GCAATTCTT	480
GGTAACCTCN TCGAGTACTT TTGACTCNCA AGAACAAAGGG AAAGGAGG	528

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 143 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: not relevant
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vii) IMMEDIATE SOURCE:
 (B) CLONE: rls72.pk0018.e7

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Met Glu Asn Gly Ala Ala Thr Asn Gly Ala Ser Glu Lys Ser His Ser
 1 5 10 15

Pro Ser Gln Thr Tyr Leu Ser Thr Arg Gly Asp Asp Tyr Gly Leu Ser
 20 25 30

Phe Glu Thr Val Val Leu Lys Gly Leu Ala Ala Asp Gly Gly Leu Phe
 35 40 45

Leu Pro Glu Glu Val Pro Ala Ala Thr Glu Trp Gln Ser Trp Lys Asp
 50 55 60

Leu Pro Tyr Thr Glu Leu Ala Val Lys Val Leu Ser Leu Tyr Ile Ser
 65 70 75 80

Pro	Ala	Glu	Val	Pro	Thr	Glu	Asp	Leu	Arg	Ala	Leu	Val	Glu	Arg	Ser
85						90							95		
Tyr	Ser	Thr	Phe	Arg	Ser	Lys	Glu	Val	Val	Pro	Leu	Val	Lys	Leu	Glu
100						105							110		
Asp	Asn	Leu	His	Leu	Leu	Glu	Leu	Phe	His	Gly	Pro	Asn	Tyr	Ser	Phe
115						120							125		
Lys	Asp	Cys	Ala	Leu	Gln	Phe	Leu	Gly	Asn	Leu	Xaa	Glu	Tyr	Phe	
130						135						140			

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 571 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: sel.06a03
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

GGATGCAATG	GTGCAGGCTG	ATTCCACTGG	AATGTTCATATA	TGTCCACACAC	CTGGGGTGGC	60
TCTGGCGGCG	CTTATTAAGC	TGAGGAATCG	TGGGGTTATC	GGTGCCGGTG	AGAGGGTTGT	120
GGTGGTGAGC	ACTGCACATG	GATTGAAGTT	TGCACAGAGC	AAGATTGATT	ATCATTCTGG	180
GCTCATTCCCT	GGAAATGGGCC	GCTATGCTAA	CCCGCTGGTT	TCGGTTAAGG	CGGATTTGG	240
ATCGGTATG	GATGTTCTCA	AGGATTCTTG	CACAACAAGT	CCCCCGACTT	TAACAAGTCT	300
TGACGTTGCC	AAAGTAAGTTT	TAGTTCGGGG	TTTTTTCTGA	TTAAAGATGT	TTTTAAACAT	360
GTTTGTGTNC	ACTTTCGGTC	GTTATTATGG	ATTTGTAAGA	TTGGGCCCAA	GTATTGGAGG	420
GTTTGATTTC	AAACAACATG	CTTCTGGTGA	CGCAATGCAA	ATTCGGNGC	ATAACATCAT	480
TGTCGAAGAT	GGATCNCGAC	CGATGAAACT	GTGTGGCAAG	TAATGAGAAG	AAAATAGGGC	540
ACTTGTACAG	AGATTTNAAA	GNTTAATTTC	N			571

(2) INFORMATION FOR SEQ ID NO:22:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 104 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: sel.06a03
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Asp	Ala	Met	Val	Gln	Ala	Asp	Ser	Thr	Gly	Met	Phe	Ile	Cys	Pro	His
1						5			10				15		

Thr Gly Val Ala Leu Ala Ala Leu Ile Lys Leu Arg Asn Arg Gly Val
 20 25 30
 Ile Gly Ala Gly Glu Arg Val Val Val Val Ser Thr Ala His Gly Leu
 35 40 45
 Lys Phe Ala Gln Ser Lys Ile Asp Tyr His Ser Gly Leu Ile Pro Gly
 50 55 60
 Met Gly Arg Tyr Ala Asn Pro Leu Val Ser Val Lys Ala Asp Phe Gly
 65 70 75 80
 Ser Val Met Asp Val Leu Lys Asp Ser Cys Thr Thr Ser Pro Pro Thr
 85 90 95
 Leu Thr Ser Leu Asp Val Ala Lys
 100

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 2191 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vii) IMMEDIATE SOURCE:
 (B) CLONE: srl.pk0003.f6

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

GCTTCCTCTT	CTCTGTTCA	GTCTCTCCCT	TTCTCTCTCC	AAACCTCTAA	ACCCTACGGG	60
CCTCCCAAAC	CCGCCGCCCA	CTTCGTTGTC	CGCGCCCAAT	CCCCCCTCAC	TCAGAACAAAC	120
AACTCCTCCT	CCAAGCATCG	CCGCCCCGCC	GACGAGAACAA	TCCGCGACGA	GGCCCGCCGC	180
ATCAATGCGC	CCCACGACCA	CCACCTCTTC	TCGGCCAAGT	ACGTCCCCTT	CAACGCCGAC	240
TCCTCCTCCT	CCTCCTCCAC	GGAGTCCTAC	TCGCTCGACG	AGATCGTCTA	CCGCTCCCAA	300
TCCGGCGGCC	TCCTGGACGT	CCAGCACGAC	ATGGATGCC	TCAAGCGTTT	CGACGGCGAG	360
TACTGGCGCA	ACCTCTTCGA	CTCGCGCGTG	GGCAAAACCA	CCTGGCCTTA	CGGCTCCGGC	420
GTCTGGAGCA	AAAAAGAATG	GGTCCTCCCC	GAGATCCACG	ACGACGATAT	CGTCTCCGCC	480
TTCGAGGGTA	ACTCCAACCT	CTTCTGGGCC	GAGCGTTTCG	GAAACAGTT	CCTCGGCATG	540
AACGATTGT	GGGTCAAACA	CTGCGGAATC	AGCCACACGG	GCAGCTTCAA	GGATCTCGGC	600
ATGACCGTCC	TCGTCAGCCA	GGTCAATCGC	TTGAGAAAAA	TGAACCGCCC	CGTCGTCGGT	660
GTTGGTTGCG	CCTCCACCGG	TGACACATCG	GCCGCTTAT	CCGCCTATTG	CGCTTCCGCT	720
GCCATTCCCT	CCATTGTGTT	TTGCGCTGCT	AATAAAATCT	CTCTGCCCA	ACTTGTTCAG	780
CCTATTGCCA	ATGGAGCCTT	TGTGTTGAGT	ATCGACACTG	ATTTGATGG	TTGCATGCAG	840
TTGATCAGAG	AAGTCACTGC	TGAATTGCC	ATTTATTGG	CTAACTCTCT	CAACAGTTG	900
AAGTTGGAAG	GGCAGAAAAC	TGCTGCTATT	GAGATTCTGC	AGCAGTTGA	TTGGCAGGTT	960
CCTGATTGGG	TCATTGTGCC	TGGAAGCAAC	CTTGGCAACA	TTTATGCC	TTACAAAGGG	1020

TTTAAGATGT	TTCAAGAGCT	TGGGCTTG	GATAAGATTC	CAAGGCTTGT	TTGTGCTCAG	1080
GCTGCCAATG	CTGATCCTT	GTATTTGTAC	TTAAATCCG	GGTGGAAAGGA	GTTTAAGCCT	1140
GTGAAGTCGA	GCACTACATT	TGCTTCTGCC	ATTCAAATTG	GTGATCCTGT	TTCCATTGAC	1200
AGGGCGGTT	ACGCGCTAAA	GAGTTGCGAT	GGGATTGTGG	AGGAGGCCAC	GGAGGAGGAG	1260
TTGATGGATG	CTACAGCGCA	GGCGGATTCT	ACTGGGATGT	TTATTTGCC	CCACACCGGG	1320
GTTGCTTAA	CTGCATTGTT	TAAGCTCAGG	AACAGCGGGG	TTATTAAGGC	CACTGATAGG	1380
ACTGTGGTGG	TTAGCACTGC	TCATGGCTTG	AAGTTCACTC	AGTCCAAGAT	TGATTACCAT	1440
TCTAAGGACA	TCAAGGACAT	GGCTTGC	TATGCTAAC	CGCCCAGC	AGTGAAGGCA	1500
GACTTTGGCT	CGGTTATGGA	TGTTTGAAG	ACGTATTGC	AGAGTAAGGC	TCATTAGGTT	1560
AGCATTGCAA	GT	TCCTGAGTTT	GCTCATTATT	TACTTACTTT	TAGGC	1620
TGCTGTATTG	TCTTTCTAT	GAGCTAGGTT	TGAGTGTGT	AATAATTGC	TTGCTGCATT	1680
ATGTATGCCG	TCTAGTGTTC	CATATTGGC	ATCATCCTTA	GTATTTGTTG	TAGATTTCT	1740
TTGCTGAGCA	TTTGATATAA	TAGCTCAAGT	AGGAAAATGA	ATTGGGTACT	ATGAGGAATG	1800
CATATCATTG	GCTTGTATT	ACTGGATTCC	AGACCACCCC	AAAAGAAAAT	AATTCCAAAA	1860
AATATAATT	GAACAAATT	CGTCCTGTT	ATGCTGTGG	CATTAAGCTC	AGTGTGGTA	1920
TTACCAAGCA	ACTCGAAATC	AAGAGAAAAA	AAAATTGACA	GCAAAGGAGC	TGCATTGTTG	1980
GACTGAGTCA	CATCACTTCA	TTGCTATGTC	GTCATATTTC	GTTGAATTAC	GGGAAGGCAG	2040
CATGCACAGC	AATATGCAGC	GATTA	AGCCACACCG	CACACATTGA	AGTAGTAGTC	2100
AATTTAGACA	CTCCATCTT	TACTTCTAC	AAAAATGAAT	TTTCTTAGC	CATTAAGTAT	2160
AATATTTTAT	TCTAAAAAAA	AAAAAAA	A			2191

(2) INFORMATION FOR SEQ ID NO:24:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 518 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: srl.pk0003.f6
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Ala	Ser	Ser	Ser	L	e	u	P	h	e	G	l	n	S	e	r	1
																5
																10
																15

L	y	s	P	o	T	Y	A	l	a	P	o	l	s	P	o	l	20
																	25
																	30

G	l	n	S	e	r	P	o	l	a	u	h	l	o	u	h	l	35
																	40
																	45

Pro Ala Asp Glu Asn Ile Arg Asp Glu Ala Arg Arg Ile Asn Ala Pro
 50 55 60
 His Asp His His Leu Phe Ser Ala Lys Tyr Val Pro Phe Asn Ala Asp
 65 70 75 80
 Ser Ser Ser Ser Ser Thr Glu Ser Tyr Ser Leu Asp Glu Ile Val
 85 90 95
 Tyr Arg Ser Gln Ser Gly Gly Leu Leu Asp Val Gln His Asp Met Asp
 100 105 110
 Ala Leu Lys Arg Phe Asp Gly Glu Tyr Trp Arg Asn Leu Phe Asp Ser
 115 120 125
 Arg Val Gly Lys Thr Thr Trp Pro Tyr Gly Ser Gly Val Trp Ser Lys
 130 135 140
 Lys Glu Trp Val Leu Pro Glu Ile His Asp Asp Asp Ile Val Ser Ala
 145 150 155 160
 Phe Glu Gly Asn Ser Asn Leu Phe Trp Ala Glu Arg Phe Gly Lys Gln
 165 170 175
 Phe Leu Gly Met Asn Asp Leu Trp Val Lys His Cys Gly Ile Ser His
 180 185 190
 Thr Gly Ser Phe Lys Asp Leu Gly Met Thr Val Leu Val Ser Gln Val
 195 200 205
 Asn Arg Leu Arg Lys Met Asn Arg Pro Val Val Gly Val Gly Cys Ala
 210 215 220
 Ser Thr Gly Asp Thr Ser Ala Ala Leu Ser Ala Tyr Cys Ala Ser Ala
 225 230 235 240
 Ala Ile Pro Ser Ile Val Phe Leu Pro Ala Asn Lys Ile Ser Leu Ala
 245 250 255
 Gln Leu Val Gln Pro Ile Ala Asn Gly Ala Phe Val Leu Ser Ile Asp
 260 265 270
 Thr Asp Phe Asp Gly Cys Met Gln Leu Ile Arg Glu Val Thr Ala Glu
 275 280 285
 Leu Pro Ile Tyr Leu Ala Asn Ser Leu Asn Ser Leu Lys Leu Glu Gly
 290 295 300
 Gln Lys Thr Ala Ala Ile Glu Ile Leu Gln Gln Phe Asp Trp Gln Val
 305 310 315 320
 Pro Asp Trp Val Ile Val Pro Gly Ser Asn Leu Gly Asn Ile Tyr Ala
 325 330 335
 Phe Tyr Lys Gly Phe Lys Met Phe Gln Glu Leu Gly Leu Val Asp Lys
 340 345 350
 Ile Pro Arg Leu Val Cys Ala Gln Ala Ala Asn Ala Asp Pro Leu Tyr
 355 360 365
 Leu Tyr Phe Lys Ser Gly Trp Lys Glu Phe Lys Pro Val Lys Ser Ser
 370 375 380
 Thr Thr Phe Ala Ser Ala Ile Gln Ile Gly Asp Pro Val Ser Ile Asp
 385 390 395 400

Arg Ala Val His Ala Leu Lys Ser Cys Asp Gly Ile Val Glu Glu Ala
 405 410 415
 Thr Glu Glu Glu Leu Met Asp Ala Thr Ala Gln Ala Asp Ser Thr Gly
 420 425 430
 Met Phe Ile Cys Pro His Thr Gly Val Ala Leu Thr Ala Leu Phe Lys
 435 440 445
 Leu Arg Asn Ser Gly Val Ile Lys Ala Thr Asp Arg Thr Val Val Val
 450 455 460
 Ser Thr Ala His Gly Leu Lys Phe Thr Gln Ser Lys Ile Asp Tyr His
 465 470 475 480
 Ser Lys Asp Ile Lys Asp Met Ala Cys Arg Tyr Ala Asn Pro Pro Met
 485 490 495
 Gln Val Lys Ala Asp Phe Gly Ser Val Met Asp Val Leu Lys Thr Tyr
 500 505 510
 Leu Gln Ser Lys Ala His
 515

(2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 643 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: wr1.pk0085.h2
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

GCTCATCCAG CCCATGCCA ACGGCGCCAC GGTGCTCTCG CTTGACACGG ATTCGACGG	60
ATGCATGCGG CTTATCAGGG AGGTGACAGC TGAGCTGCC ATATACCTCG CAAACTCACT	120
CAACTCGCTT CCGGCTGGAG GGGCAGAAGA CTGCAGCCAT CCGAGATATT GCAACANTCA	180
ATTGGCAGGT GCCCGGACTG GGTACATCC CAAGGAGGCA ATCTGGGGGA ACATTTATG	240
CTTTCTACA AGGATTNAA TTTCCGTGTC CTTNGCTAGT TGATTNCCTT CCNACTCCTT	300
GTTANTNCAA NAGGCCGCCA ACGCAAACCC ACTGTACCCG TACTACAATC CTGGGGTGAC	360
TGATTTCCAT CCACTTGNTT GCCGGGACAA TTTNCATCCN GCAACAATTG GGGGATTCCA	420
TATCNATTAC CNTCGGTTTT TTCNCCCTNA AAGGACNNAT GATTNTCCNA GGAACCTCCNN	480
AGGNGGATCA AGGATCCAAA GGCTTTCTAC TCACTGGAAN TTGCTTCCCA ANACGGGGTT	540
CACTNCCGCC CGTTAACCC NTGACAAGTA TAATGGACAA CACNCCGGGG TNTATNACAA	600
CGGCAANTTN AAANCAAGTT NATCATTAGA ACNGGAANTT NCC	643

(2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 84 amino acids
 - (B) TYPE: amino acid

(C) STRANDEDNESS: not relevant
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vii) IMMEDIATE SOURCE:
 (B) CLONE: wr1.pk0085.h2

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Leu Ile Gln Pro Ile Ala Asn Gly Ala Thr Val Leu Ser Leu Asp Thr
 1 5 10 15

Asp Phe Asp Gly Cys Met Arg Leu Ile Arg Glu Val Thr Ala Glu Leu
 20 25 30

Pro Ile Tyr Leu Ala Asn Ser Leu Asn Ser Leu Xaa Leu Glu Gly Gln
 35 40 45

Lys Thr Ala Ala Ile Arg Asp Ile Ala Thr Xaa Asn Trp Gln Val Pro
 50 55 60

Gly Leu Gly His Ile Pro Arg Arg Gln Ser Xaa Thr Phe Tyr Ala Phe
 65 70 75 80

Leu Gln Gly Phe

(2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 525 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: not relevant
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vi) ORIGINAL SOURCE:
 (A) ORGANISM: Arabidopsis thaliana

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

Leu Ser Ser Cys Leu Phe Asn Ala Ser Val Ser Ser Leu Asn Pro Lys
 1 5 10 15

Gln Asp Pro Ile Arg Arg His Arg Ser Thr Ser Leu Leu Arg His Arg
 20 25 30

Pro Val Val Ile Ser Cys Thr Ala Asp Gly Asn Asn Ile Lys Ala Pro
 35 40 45

Ile Glu Thr Ala Val Lys Pro Pro His Arg Thr Glu Asp Asn Ile Arg
 50 55 60

Asp Glu Ala Arg Arg Asn Arg Ser Asn Ala Val Asn Pro Phe Ser Ala
 65 70 75 80

Lys Tyr Val Pro Phe Asn Ala Ala Pro Gly Ser Thr Glu Ser Tyr Ser
 85 90 95

Leu Asp Glu Ile Val Tyr Arg Ser Arg Ser Gly Gly Leu Leu Asp Val
 100 105 110

Glu His Asp Met Glu Ala Leu Lys Arg Phe Asp Gly Ala Tyr Trp Arg
 115 120 125

Asp Leu Phe Asp Ser Arg Val Gly Lys Ser Thr Trp Pro Tyr Gly Ser
130 135 140

Gly Val Trp Ser Lys Lys Glu Trp Val Leu Pro Glu Ile Asp Asp Asp
145 150 155 160

Asp Ile Val Ser Ala Phe Glu Gly Asn Ser Asn Leu Phe Trp Ala Glu
165 170 175

Arg Phe Gly Lys Gln Phe Leu Gly Met Asn Asp Leu Trp Val Lys His
180 185 190

Cys Gly Ile Ser His Thr Gly Ser Phe Lys Asp Leu Gly Met Thr Val
195 200 205

Leu Val Ser Gln Val Asn Arg Leu Arg Lys Met Lys Arg Pro Val Val
210 215 220

Gly Val Gly Cys Ala Ser Thr Gly Asp Thr Ser Ala Ala Leu Ser Ala
225 230 235 240

Tyr Cys Ala Ser Ala Gly Ile Pro Ser Ile Val Phe Leu Pro Ala Asn
245 250 255

Lys Ile Ser Met Ala Gln Leu Val Gln Pro Ile Ala Asn Gly Ala Phe
260 265 270

Val Leu Ser Ile Asp Thr Asp Phe Asp Gly Cys Met Lys Leu Ile Arg
275 280 285

Glu Ile Thr Ala Glu Leu Pro Ile Tyr Leu Ala Asn Ser Leu Asn Ser
290 295 300

Leu Arg Leu Glu Gly Gln Lys Thr Ala Ala Ile Glu Ile Leu Gln Gln
305 310 315 320

Phe Asp Trp Gln Val Pro Asp Trp Val Ile Val Pro Gly Gly Asn Leu
325 330 335

Gly Asn Ile Tyr Ala Phe Tyr Lys Gly Phe Lys Met Cys Gln Glu Leu
340 345 350

Gly Leu Val Asp Arg Ile Pro Arg Met Val Cys Ala Gln Ala Ala Asn
355 360 365

Ala Asn Pro Leu Tyr Leu His Tyr Lys Ser Gly Trp Lys Asp Phe Lys
370 375 380

Pro Met Thr Ala Ser Thr Thr Phe Ala Ser Ala Ile Gln Ile Gly Asp
385 390 395 400

Pro Val Ser Ile Asp Arg Ala Val Tyr Ala Leu Lys Lys Cys Asn Gly
405 410 415

Ile Val Glu Glu Ala Thr Glu Glu Leu Met Asp Ala Met Ala Gln
420 425 430

Ala Asp Ser Thr Gly Met Phe Ile Cys Pro His Thr Gly Val Ala Leu
435 440 445

Thr Ala Leu Phe Lys Leu Arg Asn Gln Gly Val Ile Ala Pro Thr Asp
450 455 460

Arg Thr Val Val Val Ser Thr Ala His Gly Leu Lys Phe Thr Gln Ser
465 470 475 480

Lys Ile Asp Tyr His Ser Asn Ala Ile Pro Asp Met Ala Cys Arg Phe
 485 490 495

Ser Asn Pro Pro Val Asp Val Lys Ala Asp Phe Gly Ala Val Met Asp
 500 505 510

Val Leu Lys Ser Tyr Leu Gly Ser Asn Thr Leu Thr Ser
 515 520 525

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1478 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vii) IMMEDIATE SOURCE:
 (B) CLONE: cen1.pk0064.f4

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

CAACAGTGGT CCTTGAGGGG GACTCATATG ATGAAGCTCA GTCATATGCA AAATTGCGTT	60
GCCAGCAGGA AGGCCGCACA TTTGTACCTC CTTTGACCA TCCTGATGTC ATCACTGGAC	120
AAGGAACATAT CGGCATGGAA ATTGTTAGGC AGCTGCA AGG TCCACTGCAT GCAATATTTG	180
TACCTGTTGG AGGTGGTGG A TTAATTGCTG GAATTGCTGC CTATGTAAAA CGGGTTCGCC	240
CAGAGGTGAA AATAATTGGA GTGGAACCC CAGATGCAAA TCGAATGGCA TTATCCTTGT	300
GTCATGGTAA GAGGGTCATG TTGGAGCATG TTGGTGGTT TGCTGATGGT GTAGCTGTCA	360
AAGCTGTTGG GGAAGAAACA TTTCGCCTGT GCAGAGAGCT AGTAGATGGC ATTGTTATGG	420
TCAGTCGAGA TGCTATTGT GCTTCATAA AGGATATGTT TGAGGGAGAAA AGAAAGTATCC	480
TTGAACCTGC TGGTGCCCTT GCATTGGCTG GGGCTGAAGC CTACTGCAAA TACTATAACT	540
TGAAAGGAGA AACTGTTGGT GCAATAACTA GTGGGGCAAA TATGAACCTT GATCGACTTA	600
GACTAGAAC CGAGCTAGCT GATGTTGGCC GAAAACGGGA AGCAGTGTGTA GCTACATTT	660
TGCCAGAGCG GCAGGGAAAGC TTCAAAAAAT TCACAGAATT GGTGGCAGG ATGAATATTA	720
CTGAATTCAA ATACAGATAC GATTCTAATG CAAAAGATGC CCTTGTCTT TACAGTGTG	780
GCATCTACAC TGACAATGAG CTTGGAGCAA TGATGGATCG CATGGAATCT GCGAAACTGA	840
GGACTGTTAA CCTTACTGAC AATGATTGGA CAAAGGACCA CCTTAGATAC TTTATTGGAG	900
GAAGATCAGA AATAAAAGAT GAACTGGTTT ACCGGTTCAT TTTCCCGGAA AGGCCTGGGG	960
CCCTTATGAA ATTTTGGAC ACGTTTAGTC CTCGTTGGAA CATCAGCCTT TTCCATTACC	1020
GTGCACAGGG TGAACCTGGA GCAAATGTAT TAGTTGGTAT ACAAGTGCCG CCAGCAGAAT	1080
TTGATGAATT CAAGAGTCAT GCCAACAATC TTGGGTACGA GTACATGTCA GAGCACAAACA	1140
ATGAGATATA CCGGTTGCTG TTGCGTGACC CAAAGGTCTA ATGTATATGC CTTTGCTCCC	1200
ATAATAAGTT GGTGACACTT TTCAAGGAAG ATTTGCTCC AAGGTAGAAG TTGCGAGTTT	1260

CTTCAAGTTG AAATGAAGCC ATCACCAAAT GTAGCTTCGG TGTGCCATCT GTTTACTCAG	1320
TTAGATCATG TAGTGTATCA GTTGTGTATC TTTGTTGTTG TGCTTCGTGA TCTCAATTAA	1380
TTGCTTTGTG CACCTAGAGG TTGTCAAATA ATGATAACCG ATATGTTATC TAAATATCTA	1440
ATAATGATTA TGTGATTGTG ATTAAAAAGG GGGGGCCC	1478

(2) INFORMATION FOR SEQ ID NO:29:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 392 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: not relevant
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vii) IMMEDIATE SOURCE:
 (B) CLONE: cen1.pk0064.f4

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

Thr Val Val Leu Glu Gly Asp Ser Tyr Asp Glu Ala Gln Ser Tyr Ala			
1	5	10	15
Lys Leu Arg Cys Gln Gln Glu Gly Arg Thr Phe Val Pro Pro Phe Asp			
20	25	30	
His Pro Asp Val Ile Thr Gly Gln Gly Thr Ile Gly Met Glu Ile Val			
35	40	45	
Arg Gln Leu Gln Gly Pro Leu His Ala Ile Phe Val Pro Val Gly Gly			
50	55	60	
Gly Gly Leu Ile Ala Gly Ile Ala Ala Tyr Val Lys Arg Val Arg Pro			
65	70	75	80
Glu Val Lys Ile Ile Gly Val Glu Pro Ser Asp Ala Asn Ala Met Ala			
85	90	95	
Leu Ser Leu Cys His Gly Lys Arg Val Met Leu Glu His Val Gly Gly			
100	105	110	
Phe Ala Asp Gly Val Ala Val Lys Ala Val Gly Glu Glu Thr Phe Arg			
115	120	125	
Leu Cys Arg Glu Leu Val Asp Gly Ile Val Met Val Ser Arg Asp Ala			
130	135	140	
Ile Cys Ala Ser Ile Lys Asp Met Phe Glu Glu Lys Arg Ser Ile Leu			
145	150	155	160
Glu Pro Ala Gly Ala Leu Ala Leu Ala Gly Ala Glu Ala Tyr Cys Lys			
165	170	175	
Tyr Tyr Asn Leu Lys Gly Glu Thr Val Val Ala Ile Thr Ser Gly Ala			
180	185	190	
Asn Met Asn Phe Asp Arg Leu Arg Leu Val Thr Glu Leu Ala Asp Val			
195	200	205	
Gly Arg Lys Arg Glu Ala Val Leu Ala Thr Phe Leu Pro Glu Arg Gln			
210	215	220	

Gly Ser Phe Lys Lys Phe Thr Glu Leu Val Gly Arg Met Asn Ile Thr
 225 230 235 240
 Glu Phe Lys Tyr Arg Tyr Asp Ser Asn Ala Lys Asp Ala Leu Val Leu
 245 250 255
 Tyr Ser Val Gly Ile Tyr Thr Asp Asn Glu Leu Gly Ala Met Met Asp
 260 265 270
 Arg Met Glu Ser Ala Lys Leu Arg Thr Val Asn Leu Thr Asp Asn Asp
 275 280 285
 Leu Ala Lys Asp His Leu Arg Tyr Phe Ile Gly Gly Arg Ser Glu Ile
 290 295 300
 Lys Asp Glu Leu Val Tyr Arg Phe Ile Phe Pro Glu Arg Pro Gly Ala
 305 310 315 320
 Leu Met Lys Phe Leu Asp Thr Phe Ser Pro Arg Trp Asn Ile Ser Leu
 325 330 335
 Phe His Tyr Arg Ala Gln Gly Glu Ala Gly Ala Asn Val Leu Val Gly
 340 345 350
 Ile Gln Val Pro Pro Ala Glu Phe Asp Glu Phe Lys Ser His Ala Asn
 355 360 365
 Asn Leu Gly Tyr Glu Tyr Met Ser Glu His Asn Asn Glu Ile Tyr Arg
 370 375 380
 Leu Leu Leu Arg Asp Pro Lys Val
 385 390

(2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 728 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: sf11.pk0055.h7

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

AAAAATATTGT AGCAATAACC AGTGGAGCAA ACATGAAATT TGATAAACTT CGGGTTGTAA	60
CTGAACCTTGC TAATGTTGGT CGTAAACAAG AGGCTGTGCT GGCAACTGTT ATGGCAGAGG	120
AGCCTGGCAG TTTCAAACAA TTTTGTGAAT TGGTGGGGCA GATGAACATA ACAGAATTCA	180
AATACAGATA TAACTCAAAT GAGAAGGCAG TTGTCCTTTA CAGTGTGGG GTTCACACAA	240
TCTCCGAACT AAGAGCAATG CAGGAGAGGA TGGAACTTTC TCAGCTCAAAC TTACAATC	300
TCACAGAAAG TGACTTGGTG AAAGACCACT TGCCTTACTT GATGGGAGGC CGATCAAACG	360
TTCAGAAATGA GGTCTTGTC GTCTCACCTT TCCAAGAAAG ACTGGTGCTT TGATGAAATT	420
TTGGGACCCCT TCAGTCCACG TTGGGATATT AGTTTATCCA TTACCGAGGG GAGGTGAAAC	480
TGGAGCAAAC TGCTAGTTGG NTACAGGTAC CAAAATGAGA TAGATGAGTC CATGATCGTG	540

CTAACAAACT GGATATGATT ATAAGTGGNA ATATGTGATG NCTCAGCTCA ATCNCGATGG	600
GGNTTAAGCA CTGCATATGG GNATTAGGGG NAGNTACANT TAAATTACAG GCCTCAAGNT	660
AAGCATANTN TAGGAACTAG CTTTACAGGG GGCTACNANT TAACCGNGTA TTTTTTTGTA	720
GATGANNG	728

(2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 152 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: sf11.pk0055.h7
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

Asn Ile Val Ala Ile Thr Ser Gly Ala Asn Met Asn Phe Asp Lys Leu			
1	5	10	15
Arg Val Val Thr Glu Leu Ala Asn Val Gly Arg Lys Gln Glu Ala Val			
20	25	30	
Leu Ala Thr Val Met Ala Glu Glu Pro Gly Ser Phe Lys Gln Phe Cys			
35	40	45	
Glu Leu Val Gly Gln Met Asn Ile Thr Glu Phe Lys Tyr Arg Tyr Asn			
50	55	60	
Ser Asn Glu Lys Ala Val Val Leu Tyr Ser Val Gly Val His Thr Ile			
65	70	75	80
Ser Glu Leu Arg Ala Met Gln Glu Arg Met Glu Ser Ser Gln Leu Lys			
85	90	95	
Thr Tyr Asn Leu Thr Glu Ser Asp Leu Val Lys Asp His Leu Arg Tyr			
100	105	110	
Leu Met Gly Gly Arg Ser Asn Val Gln Asn Glu Val Phe Val Val Ser			
115	120	125	
Pro Xaa Pro Arg Lys Thr Gly Ala Leu Met Lys Phe Leu Asp Xaa Phe			
130	135	140	
Ser Pro Arg Trp Asp Ile Ser Leu			
145	150		

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 572 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: sre.pk0044.f3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

AAAGACCTGG	TGCTTGATG	AAATTTGG	ACCCCTTCAG	TCCACGTTGG	AATATCAGTT	60
TATTCCATTA	CCGAGGGGAG	GGTGAAACTG	GACCAAATGT	GCTAGTTGGA	ATACAGGTAC	120
CCAAAAGTGA	GATGGATGAG	TTCCACGATC	GTGCCAACAA	ACTTGGATAT	GATTATAAAAG	180
TGGTGAATAA	TGATGATGAC	TTCCAGCTTC	TAATGCACTG	ATGATGGTTT	TAGGCACTTG	240
CCATTATTGT	GTATTTAGT	CAACAAAGTTT	GCCATATTAA	ATATTCAC	GGTCGTTCT	300
AAAAGTTGGA	TGGGGAAAAA	AGGTGGAAAG	GAAGTGGCCT	TCAGACATGT	CATTAGTTGA	360
TTAGAGGAAC	AACTAGTTCT	TTTACCTAA	TGCGGCGTCT	TATTACATT	TTTATAATCT	420
GTAATTTATG	TTTTTTGTT	GTTGTTAACAA	TTGGAATCTT	ATAATGTTGT	TGCCTGGTCT	480
TTTGTGTCTG	TAATATAAGT	GTCTCAAAA	GGTTGTTGC	TAAATTCAG	CAGCCTAAAAA	540
AAAAAAAAAA	AAAAAAAAAA	AAAAAAAAAA	AA			572

(2) INFORMATION FOR SEQ ID NO:33:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 72 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: not relevant
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vii) IMMEDIATE SOURCE:

- (B) CLONE: sre.pk0044.f3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

Arg	Pro	Gly	Ala	Leu	Met	Lys	Phe	Leu	Asp	Pro	Phe	Ser	Pro	Arg	Trp
1				5				10						15	
Asn	Ile	Ser	Leu	Phe	His	Tyr	Arg	Gly	Glu	Gly	Glu	Thr	Gly	Ala	Asn
				20				25				30			
Val	Leu	Val	Gly	Ile	Gln	Val	Pro	Lys	Ser	Glu	Met	Asp	Glu	Phe	His
				35			40				45				
Asp	Arg	Ala	Asn	Lys	Leu	Gly	Tyr	Asp	Tyr	Lys	Val	Val	Asn	Asn	Asp
	50				55				60						
Asp	Asp	Phe	Gln	Leu	Leu	Met	His								
	65				70										

(2) INFORMATION FOR SEQ ID NO:34:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 507 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: not relevant
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Burkholderia capacia

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Met Ala Ser His Asp Tyr Leu Lys Lys Ile Leu Thr Ala Arg Val Tyr
 1 5 10 15

Asp Val Ala Phe Glu Thr Glu Leu Glu Pro Ala Arg Asn Leu Ser Ala
 20 25 30

Arg Leu Arg Asn Pro Val Tyr Leu Lys Arg Glu Asp Asn Gln Pro Val
 35 40 45

Phe Ser Phe Lys Leu Arg Gly Ala Tyr Asn Lys Met Ala His Ile Pro
 50 55 60

Ala Asp Ala Leu Ala Arg Gly Val Ile Thr Ala Ser Ala Gly Asn His
 65 70 75 80

Ala Gln Gly Val Ala Phe Ser Ala Ala Arg Met Gly Val Lys Ala Val
 85 90 95

Ile Val Val Pro Val Thr Thr Pro Gln Val Lys Val Asp Ala Val Arg
 100 105 110

Ala His Gly Gly Pro Gly Val Glu Val Ile Gln Ala Gly Glu Ser Tyr
 115 120 125

Ser Asp Ala Tyr Ala His Ala Leu Lys Val Gln Glu Glu Arg Gly Leu
 130 135 140

Thr Phe Val His Pro Phe Asp Asp Pro Tyr Val Ile Ala Gly Gln Gly
 145 150 155 160

Thr Ile Ala Met Glu Ile Leu Arg Gln His Gln Gly Pro Ile His Ala
 165 170 175

Ile Phe Val Pro Ile Gly Gly Gly Leu Ala Ala Gly Val Ala Ala
 180 185 190

Tyr Val Lys Ala Val Arg Pro Glu Ile Lys Val Ile Gly Val Gln Ala
 195 200 205

Glu Asp Ser Cys Ala Met Ala Gln Ser Leu Gln Ala Gly Lys Arg Val
 210 215 220

Glu Leu Ala Glu Val Gly Leu Phe Ala Asp Gly Thr Ala Val Lys Leu
 225 230 235 240

Val Gly Glu Glu Thr Phe Arg Leu Cys Lys Glu Tyr Leu Asp Gly Val
 245 250 255

Val Thr Val Asp Thr Asp Ala Leu Cys Ala Ala Ile Lys Asp Val Phe
 260 265 270

Gln Asp Thr Arg Ser Val Leu Glu Pro Ser Gly Ala Leu Ala Val Ala
 275 280 285

Gly Ala Lys Leu Tyr Ala Glu Arg Glu Gly Ile Glu Asn Gln Thr Leu
 290 295 300

Val Ala Val Thr Ser Gly Ala Asn Met Asn Phe Asp Arg Met Arg Phe
 305 310 315 320

Val Ala Glu Arg Ala Glu Val Gly Glu Ala Arg Glu Ala Val Phe Ala
 325 330 335

Val Thr Ile Pro Glu Glu Arg Gly Ser Phe Lys Arg Phe Cys Ser Leu
 340 345 350
 Val Gly Asp Arg Asn Val Thr Glu Phe Asn Tyr Arg Ile Ala Asp Ala
 355 360 365
 Gln Ser Ala His Ile Phe Val Gly Val Gln Ile Arg Arg Arg Gly Glu
 370 375 380
 Ser Ala Asp Ile Ala Ala Asn Phe Glu Ser His Gly Phe Lys Thr Ala
 385 390 395 400
 Asp Leu Thr His Asp Glu Leu Ser Lys Glu His Ile Arg Tyr Met Val
 405 410 415
 Gly Gly Arg Ser Pro Leu Ala Leu Asp Glu Arg Leu Phe Arg Phe Glu
 420 425 430
 Phe Pro Glu Arg Pro Gly Ala Leu Met Lys Phe Leu Ser Ser Met Ala
 435 440 445
 Pro Asp Trp Asn Ile Ser Leu Phe His Tyr Arg Asn Gln Gly Ala Asp
 450 455 460
 Tyr Ser Ser Ile Leu Val Gly Leu Gln Val Pro Gln Ala Asp His Ala
 465 470 475 480
 Glu Phe Glu Arg Phe Leu Ala Ala Leu Gly Tyr Pro Tyr Val Glu Glu
 485 490 495
 Ser Ala Asn Pro Ala Tyr Arg Leu Phe Leu Ser
 500 505

(2) INFORMATION FOR SEQ ID NO:35:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1582 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: cc3.mn0002d2
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

ACGAGACGAG	TCCCCTCCCC	CCACCTCGCC	TCACCCAAACC	GGAACGAACA	AGTTACCATC	60
TCATCCCAAC	CCCGCCTCGA	CCGGATCTCG	TCGGACTCGG	ATCCGCCCGA	CCACCCCGCG	120
CCGCCGCAGA	TCAAAGAAGA	TGGCAGCTCT	CGACACCTTC	CTCTTCACCT	CGGAGTCTGT	180
GAACGAGGGA	CACCTGACA	AGCTCTGCGA	CCAGGTCTCA	GATGCCGTT	TTGACGCTTG	240
CCTTGCTGAG	GACCCTGACA	GCAAGGTTGC	TTGTGAGACC	TGCACCAAGA	CCAACATCGT	300
CATGGTCTTT	GGTGAGATCA	CCACCAAGGC	CAATGTCGAC	TACGAGAAGA	TTGTCAGGGA	360
GACCTGCCGC	AACATTGGTT	TTGTGTCAAA	CGATGTCGGG	CTTGACGCTG	ACCACTGCAA	420
GGTGCTCGTG	AACATTGAGC	AGCAGTCCCC	TGATATTGCT	CAGGGTGTGC	ATGGCCACTT	480
CACCAAGCGC	CCCGAGGGAGA	TTGGAGCTGG	TGACCAGGGA	CACATGTTCG	GGTATGCGAC	540

CGATGAGACC CCTGAGTTGA TGCCCCTCAG CCATGTCCTT GCCACCAAGC TAGGTGCTCG	600
TCTCACCGAG GTCCGCAAGA ACGGAACCTG CCCCTGGCTC AGGCCTGATG GGAAGACCCA	660
GGTGACAGTC GAGTACCGCA ATGAGGGTGG TGCCATGGTC CCCATCCGTG TCCACACCGT	720
CCTCATCTCC ACCCAGCACG ACGAGACAGT GACCAATGAT GAGATCGCTG CTGACCTGAA	780
GGAGCATGTC ATCAAGCCTA TCATCCCTGA GCAGTACCTT GACCGAGAAGA CCATCTTCCA	840
CCTTAACCCA TCCGGCCGCT TTGTCATTGG TGGACCTCAC GCGCATGCTG GCCTCACTGG	900
CCGCAAGATC ATCATTGACA CCTACGGTGG CTGGGGAGCC CATGGCGGTG GCGCTTCTC	960
CGGCAAGGAC CCAACCAAGG TTGACCGCAG CGGAGCCTAT GTCCGAGGC AGGCTGCCAA	1020
GAGCATCGTC GCCAGCGGCC TTGCTCGCCG CGCCATCGTC CAGGTGTCCCT ACGCCATCGG	1080
CGTCCCCGAG CCTCTCTCCG TGTTTGTGCA CACGTACGGC ACCGGCGCGA TCCCCGACAA	1140
GGAGATCCTC AAGATTGTCA AGGAGAACTT CGATTCAGG CCTGGCATGA TTATCATCAA	1200
CCTTGACCTC AAGAAAGGCG GCAACGGGCG CTACCTCAAG ACGGCAGCCT ACGGCCACTT	1260
CGGAAGGGAC GACCCCTGACT TCACCTGGGA GGTGGTGAAG CCACTCAAGT CGGAGAAACC	1320
TTCTGCCTAA GGC GGCCCTTT TTTTCAGTAA GAAGCTTTG GTGGTCTGCT GTGCTTAATC	1380
ATGCTTTAT ATGGCTTCTA CATGTTGTGG TTCTTTCTTG ATCTGCACCG CGCTTATCGT	1440
TTGTGTTGTA CTGCCCTAAT AAGTGGTGCT TATGAGGACT GTTTCTGGTT TTGCTGCTTA	1500
TGTTGTAATG CTTTGAAACA ATGAAAGAAG CTACAGGCCA CAGCTATTTT GAGAAGTAAT	1560
GGAACCTCGT GCCGTTTGA TT	1582

(2) INFORMATION FOR SEQ ID NO:36:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 396 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: cc3.mn0002.d2
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Met Ala Ala Leu Asp Thr Phe Leu Phe Thr Ser Glu Ser Val Asn Glu			
1	5	10	15
Gly His Pro Asp Lys Leu Cys Asp Gln Val Ser Asp Ala Val Leu Asp			
20	25	30	
Ala Cys Leu Ala Glu Asp Pro Asp Ser Lys Val Ala Cys Glu Thr Cys			
35	40	45	
Thr Lys Thr Asn Met Val Met Val Phe Gly Glu Ile Thr Thr Lys Ala			
50	55	60	
Asn Val Asp Tyr Glu Lys Ile Val Arg Glu Thr Cys Arg Asn Ile Gly			
65	70	75	80

Phe Val Ser Asn Asp Val Gly Ile Asp Ala Asp His Cys Lys Val Leu
 85 90 95
 Val Asn Ile Glu Gln Gln Ser Pro Asp Ile Ala Gln Gly Val His Gly
 100 105 110
 His Phe Thr Lys Arg Pro Glu Glu Ile Gly Ala Gly Asp Gln Gly His
 115 120 125
 Met Phe Gly Tyr Ala Thr Asp Glu Thr Pro Glu Leu Met Pro Leu Ser
 130 135 140
 His Val Leu Ala Thr Lys Leu Gly Ala Arg Leu Thr Glu Val Arg Lys
 145 150 155 160
 Asn Gly Thr Cys Pro Trp Leu Arg Pro Asp Gly Lys Thr Gln Val Thr
 165 170 175
 Val Glu Tyr Arg Asn Glu Gly Gly Ala Met Val Pro Ile Arg Val His
 180 185 190
 Thr Val Leu Ile Ser Thr Gln His Asp Glu Thr Val Thr Asn Asp Glu
 195 200 205
 Ile Ala Ala Asp Leu Lys Glu His Val Ile Lys Pro Ile Ile Pro Glu
 210 215 220
 Gln Tyr Leu Asp Glu Lys Thr Ile Phe His Leu Asn Pro Ser Gly Arg
 225 230 235 240
 Phe Val Ile Gly Gly Pro His Gly Asp Ala Gly Leu Thr Gly Arg Lys
 245 250 255
 Ile Ile Ile Asp Thr Tyr Gly Gly Trp Gly Ala His Gly Gly Ala
 260 265 270
 Phe Ser Gly Lys Asp Pro Thr Lys Val Asp Arg Ser Gly Ala Tyr Val
 275 280 285
 Ala Arg Gln Ala Ala Lys Ser Ile Val Ala Ser Gly Leu Ala Arg Arg
 290 295 300
 Ala Ile Val Gln Val Ser Tyr Ala Ile Gly Val Pro Glu Pro Leu Ser
 305 310 315 320
 Val Phe Val Asp Thr Tyr Gly Thr Gly Ala Ile Pro Asp Lys Glu Ile
 325 330 335
 Leu Lys Ile Val Lys Glu Asn Phe Asp Phe Arg Pro Gly Met Ile Ile
 340 345 350
 Ile Asn Leu Asp Leu Lys Lys Gly Gly Asn Gly Arg Tyr Leu Lys Thr
 355 360 365
 Ala Ala Tyr Gly His Phe Gly Arg Asp Asp Pro Asp Phe Thr Trp Glu
 370 375 380
 Val Val Lys Pro Leu Lys Ser Glu Lys Pro Ser Ala
 385 390 395

(2) INFORMATION FOR SEQ ID NO:37:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 2183 base pairs
(B) TYPE: nucleic acid

(C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Oryza sativa

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

GAATTCTTAT AAATGAACGG AAAATGGAAA AAAAAATTGA TTGGTGCCAC TTCAAAGTTA	60
AATATGCCAA GACGAATTGA TATGTTCTG CTGTTGTTT ATGCTCTTGA TTAGTTGATG	120
CGCATGTTCA ATGATTATG ATGTTGTCT TTGTGAAAG ATTACATGTA AAGAGTATAG	180
TAGAACCCCT AAAAGCTAGC CAGCGATTTC GCTCTTTT TCCAGGTCTC CATGATATGT	240
TTACCCCTAA AAGTGGTATA TTTATGTGAT AGTTACAATA CATACTGGAC CACGATTGAT	300
TATGCGTTA TGCTGATTCC GGCAGAAAAT TGTTAGATT CTTGTGCTCT ATACCTGCTT	360
GTTGCGCTTC TAGAGAATAT TACAAATACC TAACACTTGC CCAAGGAAC TAGGAACCTA	420
GTCAACTCTT TGTAGGGACA ACTATTTAG CCCAAAATTG TGGTCTTGTC AGGTGCCAAC	480
AAAACAGCAT CTTGGCGTAC ATAAGCTATA TAGAGGATTA AAAGGAATGT TTTGTTCCCTT	540
GCTACTGTTT TTTAACCTG TTTACTCAGG ACAAAATTTG TTGCATAAAC CATTGTTCT	600
AGGGATCAGT ATTGTCCTCT CAGTGTGTTA TGTAAGCATT TCCAGAAATC AATTGTCGCT	660
ATCAGCTTCC CTCACATTAG CTATCACTTA TACCCCTTT TTTCTCATAG GCTCACCATG	720
TCCATTTAT TCATGATATT TCTTGTCTA AAGTATGTGA AATACCATTT TATGCAGATA	780
GGAGAAGATG GCCGCACTTG ATACCTTCCT CTTTACCTCG GAGTCTGTGA ACGAGGGCCA	840
CCCTGACAAG CTCTGCGACC AAGTCTCAGA TGCTGTCCT GATGCCCTGCC TCGCCGAGGA	900
CCCTGACAGC AAGGTCGCTT GTGAGACCTG CACCAAGACA AACATGGTCA TGGTCTTGG	960
TGAGATCACC ACCAAGGCTA ACGTTGACTA TGAGAAGATT GTCAGGGAGA CATGCCGTAA	1020
CATCGGTTTT GTGTCAGCTG ATGTCGGTCT CGATGCTGAC CACTGCAAGG TGCTTGTGAA	1080
CATCGAGCAG CAGTCCCTG ACATTGCACA GGGTGTGCAC GGGCACTTCA CCAAGCGCCC	1140
TGAGGAGATT GGTGCTGGTG ACCAGGGACA CATGTTGGA TATGCAACTG ATGAGACCCC	1200
TGAGTTGATG CCCCTCAGCC ATGTCCTTGC TACCAAGCTT GGCGCTCGTC TTACGGAGGT	1260
TCGCAAGAAT GGGACCTGCG CATGGCTCAG GCCTGACGGG AAGACCCAAG TGACTGTTGA	1320
GTACCGCAAT GAGAGCGGTG CCAGGGTCCC TGTCCGTGTC CACACCGTCC TCATCTCTAC	1380
CCAGCATGAT GAGACAGTCA CCAACGATGA GATTGCTGCT GACCTGAAGG AGCATGTCAT	1440
CAAGCCTGTC ATTCCCGAGC AGTACCTTGA TGAGAAGACA ATCTTCCATC TTAACCCATC	1500
TGGTCGCTTC GTCATTGGCG GACCTCATGG TGATGCTGGT CTCACTGGCC GGAAGATCAT	1560
CATTGACACT TATGGTGGCT GGGGAGCTA CGGTGGTGGT GCCTTCTCTG GCAAGGACCC	1620
AACCAAGGTT GACCGCAGTG GAGCATACTGT CGCAAGGCAA GCTGCCAAGA GCATTGTTGC	1680

TAGTGGCCTT GCTCGCCGCT GCATTGTCCA AGTATCATAAC	GCCATCGGTG TCCCAGAGCC	1740
ACTGTCCGTA TTCGTCGACA CATA CGGCAC TGGCAGGATC	CCTGACAAGG AGATCCTCAA	1800
GATTGTGAAG GAGAACTTCG ACTTCAGGCC TGGCATGATC	ATCATCAACC TTGACCTCAA	1860
GAAAGGCGGC AACGGACGCT ACCTCAAGAC GGCGGCTTAC	GGTCACCTCG GAAGGGACGA	1920
CCCAGACTTC ACCTGGGAGG TGCTGAAGCC CCTCAAGTGG	GAGAAGCCTT CTGCCTAAAA	1980
GCTCCCTTC GGAGGCTTT GCTCTGTCCC ATTATGGTGT	TTTGTTCCT CGCTGCTCAG	2040
CATTGTGATT CTTAACCTGC CCCCCGCTGC CATTATGCC	CATGCACGCT ACTTTCTAA	2100
TAATAAGTAC TTATAAGGGT ATTGTGTTG AATATTTCAC	CTAGAGGAGG AGGAGGATTT	2160
GTTATCTGTT ATTGCTTAAG CTT		2183

(2) INFORMATION FOR SEQ ID NO:38:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1485 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vii) IMMEDIATE SOURCE:
 (B) CLONE: s2.12b06

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

AGCCAAGCCC CACTCAACCA CCACACCACT CTCTCTGTC TTCTTCTACC	TTTCAAGTTT	60
TTAAAGTATT AAGATGGCAG AGACATTCTT ATTACCTCA GAGTCAGTGA	ACGAGGGACA	120
CCCTGACAAG CTCTGCGACC AAATCTCCGA TGCTGTCTC GACGCTTGCC	TTGAACAGGA	180
CCCAGACAGC AAGGTTGCCT GCGAAACATG CACCAAGACC AACTTGGTCA	TGGTCTTCGG	240
AGAGATCACC ACCAAGGCCA ACGTTGACTA CGAGAAGATC GTGCGTGACA	CCTGCAGGAA	300
CATCGCTTC GTCTCAAACG ATGTGGGACT TGATGCTGAC AACTGCAAGG	TCCTTGTAAA	360
CATTGAGCAG CAGAGCCCTG ATATTGCCCA GGGTGTGCAC GGCCACCTTA	CCAAAAGACC	420
CGAGGAAATC GGTGCTGGAG ACCAGGGTCA CATGTTGGC TATGCCACGG	ACGAAACCCC	480
AGAATTGATG CCATTGAGTC ATGTTCTTGC AACTAAACTC GGTGCTCGTC	TCACCGAGGT	540
TCGCAAGAAC GGAACCTGCC CATGGTTGAG GCCTGATGGG AAAACCCAAG	TGACTGTTGA	600
GTATTACAAT GACAACGGTG CCATGGTTCC AGTTCTGTC CACACTGTGC	TTATCTCCAC	660
CCAACATGAT GAGACTGTGA CCAACGACGA AATTGCAGCT GACCTCAAGG	AGCATGTGAT	720
CAAGCCGGTG ATCCGGAGA AGTACCTTGA TGAGAAAGACC ATTTTCCACT	TGAACCCCTC	780
TGGCCGTTT GTCATTGGAG GTCCTCACGG TGATGCTGGT CTCACCGGCC	GCAAGATCAT	840
CATCGATACT TACGGAGGAT GGGGTGCTCA TGGTGGTGGT GCTTTCTCCG	GGAAGGATCC	900
CACCAAGGTT GATAGGAGTG GTGCTTACAT TGTGAGACAG GCTGCTAAGA	GCATTGTGGC	960
AAGTGGACTA GCCAGAAGGT GCATTGTGCA AGTGTCTTAT	GCCATTGGTG TGCCCGAGCC	1020

TTTGTCTGTC	TTTGTGACA	CCTATGGCAC	CGGGAAGATC	CATGATAAGG	AGATTCTCAA	1080
CATTGTGAAG	GAGAACTTG	ATTCAGGCC	CGGTATGATC	TCCATCAACC	TTGATCTCAA	1140
GAGGGGTGGG	AATAACAGGT	TCTTGAAGAC	TGCTGCATAT	GGACACTTCG	GCAGAGAGGA	1200
CCCTGACTTC	ACATGGGAAG	TGGTCAAGCC	CCTCAAGTGG	GAGAAGGCCT	AAGGCCATTC	1260
ATTCCACTGC	AATGTGCTGG	GAGTTTTTA	GCGTTGCCCT	TATAATGTCT	ATTATCCATA	1320
ACTTTCCACG	TCCCTTGCTC	TGTGTTTTC	TCTCGTCGTC	CTCCTCCTAT	TTTGTTCCTC	1380
CTGCCTTTCA	TTTGTAAATT	TTTACATGAT	CAACTAAAAA	ATGTAATCTC	TGTTTCCGA	1440
CCATTGTGTC	TCTTAATATC	AGTATCAAAA	AGAATGTTCC	AAGTT		1485

(2) INFORMATION FOR SEQ ID NO:39:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 392 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: not relevant
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vii) IMMEDIATE SOURCE:

- (B) CLONE: s2.12b06

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

Met	Ala	Glu	Thr	Phe	Leu	Phe	Thr	Ser	Glu	Ser	Val	Asn	Glu	Gly	His
1				5				10					15		
Pro	Asp	Lys	Leu	Cys	Asp	Gln	Ile	Ser	Asp	Ala	Val	Leu	Asp	Ala	Cys
				20				25				30			
Leu	Glu	Gln	Asp	Pro	Asp	Ser	Lys	Val	Ala	Cys	Glu	Thr	Cys	Thr	Lys
				35			40				45				
Thr	Asn	Leu	Val	Met	Val	Phe	Gly	Glu	Ile	Thr	Thr	Lys	Ala	Asn	Val
				50		55			60						
Asp	Tyr	Glu	Lys	Ile	Val	Arg	Asp	Thr	Cys	Arg	Asn	Ile	Gly	Phe	Val
				65		70			75			80			
Ser	Asn	Asp	Val	Gly	Leu	Asp	Ala	Asp	Asn	Cys	Lys	Val	Leu	Val	Asn
				85			90				95				
Ile	Glu	Gln	Gln	Ser	Pro	Asp	Ile	Ala	Gln	Gly	Val	His	Gly	His	Leu
				100			105				110				
Thr	Lys	Arg	Pro	Glu	Glu	Ile	Gly	Ala	Gly	Asp	Gln	Gly	His	Met	Phe
				115		120			125						
Gly	Tyr	Ala	Thr	Asp	Glu	Thr	Pro	Glu	Leu	Met	Pro	Leu	Scr	His	Val
				130		135			140						
Leu	Ala	Thr	Lys	Leu	Gly	Ala	Arg	Leu	Thr	Glu	Val	Arg	Lys	Asn	Gly
				145		150			155			160			
Thr	Cys	Pro	Trp	Leu	Arg	Pro	Asp	Gly	Lys	Thr	Gln	Val	Thr	Val	Glu
				165			170				175				

Tyr Tyr Asn Asp Asn Gly Ala Met Val Pro Val Arg Val His Thr Val
 180 185 190
 Leu Ile Ser Thr Gln His Asp Glu Thr Val Thr Asn Asp Glu Ile Ala
 195 200 205
 Ala Asp Leu Lys Glu His Val Ile Lys Pro Val Ile Pro Glu Lys Tyr
 210 215 220
 Leu Asp Glu Lys Thr Ile Phe His Leu Asn Pro Ser Gly Arg Phe Val
 225 230 235 240
 Ile Gly Gly Pro His Gly Asp Ala Gly Leu Thr Gly Arg Lys Ile Ile
 245 250 255
 Ile Asp Thr Tyr Gly Gly Trp Gly Ala His Gly Gly Ala Phe Ser
 260 265 270
 Gly Lys Asp Pro Thr Lys Val Asp Arg Ser Gly Ala Tyr Ile Val Arg
 275 280 285
 Gln Ala Ala Lys Ser Ile Val Ala Ser Gly Leu Ala Arg Arg Cys Ile
 290 295 300
 Val Gln Val Ser Tyr Ala Ile Gly Val Pro Glu Pro Leu Ser Val Phe
 305 310 315 320
 Val Asp Thr Tyr Gly Thr Gly Lys Ile His Asp Lys Glu Ile Leu Asn
 325 330 335
 Ile Val Lys Glu Asn Phe Asp Phe Arg Pro Gly Met Ile Ser Ile Asn
 340 345 350
 Leu Asp Leu Lys Arg Gly Gly Asn Asn Arg Phe Leu Lys Thr Ala Ala
 355 360 365
 Tyr Gly His Phe Gly Arg Glu Asp Pro Asp Phe Thr Trp Glu Val Val
 370 375 380
 Lys Pro Leu Lys Trp Glu Lys Ala
 385 390

(2) INFORMATION FOR SEQ ID NO:40:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1479 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Lycopersicon esculentum
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

GAATTCTTAC AAAGAGGTTA TTTCTCTCAA GGGGTAAAAAA GATTGCCCTT TTTGACATT	60
TATAATCCTC TTTTTCTCTT TGTTGCCGT TGGGTTCTTC ACTTTCTGT TTCTTGAGAA	120
TGGAAACTTT CTTATTCAAC TCCGAGTCTG TGAACGAGGG TCACCCAGAC AAGCTCTGTG	180
ATCAGATCTC TGATGCAGTT CTTGATGCCT GCCTTGAGCA AGATCCCGAG AGCAAAGTTG	240
CATGTGAAAC TTGCACCAAG ACCAACTTGG TCATGGTCTT TGGTGAGATC ACAACCAAGG	300

CTATTGTAGA CTATGAGAAG ATTGTGCGTG ACACATGCCG TAATATTGGA TTTGTTCTG	360
ATGATGTTGG TCTTGATGCT GACAACGTGCA AGGTCCCTGT TTACATTGAG CAGCAAAGTC	420
CTGATATTGC TCAAGGTGTC CACGGCCATC TGACCAAACG CCCCAGGGAG ATTGGTGCTG	480
GTGACCAGGG CCACATGTT GGCTATGCAA CAGATGAGAC CCCTGAATTA ATGCCTCTCA	540
GTCACGTGCT TGCAACTAAA CTTGGTGCCC GTCTTACAGA AGTCCGCAAG AATGGCACCT	600
GCGCCTGGTT GAGGCCTGAT GGCAAGACCC AAGTTACTGT TGAGTATAGC AATGACAATG	660
GTGCCATGGT TCCAATTAGG GTACACACTG TTCTTATCTC CACCCAACAC GATGAGACCG	720
TTACCAATGA TGAGATTGCC CGCGACCTTA AGGAGCATGT CATCAAACCA GTCATCCAG	780
AGAAGTACCT TGATGAGAAT ACTATTTCC ACCTTAACCC ATCTGGCCGA TTCGTTATTG	840
GTGGACCTCA TGGTGTGCT GGTCTCACTG GTCGAAAAT CATCATCGAC ACTTATGGTG	900
CTTGGGGTGC TCATGGTGGT GGTGTTTCT CGGGCAAAGA CCCAACCAAG GTCGACAGGA	960
GTGGTGCATA CATTGTAAGG CAGGCTGCAA AGAGTATCGT AGCTAGTGGA CTTGCTCGTA	1020
GATGCATCGT GCAGGTATCT TATGCCATCG GTGTGCCCTGA GCCATTGTCT GTATTGTTG	1080
ACACCTATGG CACTGGAAAG ATCCCTGACA GGGAAATTTT GAAGATCGTT AAGGAGAACT	1140
TTGACTTCAG ACCTGGAATG ATGTCCATTA ACTTGGATTG GAAGAGGGGT GGCAATAGAA	1200
GATTCTTGAA AACTGCTGCC TATGGTCACT TTGGACGTGA TGACCCCGAT TTCACATGGG	1260
AAGTTGTCAA GCCCCTCAAG TGGGAAAAGC CCCAACACTA ATAAGTGCCTT GCCTATGTT	1320
TTGTTCTTG TTGTTGCTT GTGGCTTAG AATCTCCCC GTGTTGCTT GTTTGTCTT	1380
GTATTTCTC TTTTGACCCCT TTATTTGTT ATTGTCTGT TTCCATTGTG TTGGATGGAT	1440
ATCTTAGGCC TTGGAATATT AAGGAAAGAA AAGGAATTC	1479

(2) INFORMATION FOR SEQ ID NO:41:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1380 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

CCCTCCCTTC GGTCATCGG CCTCCCGATC GAGCAGTAGA AGCAGCGCAA GGGCATCGCT	60
AGCACTAAAG AAATGGCGAC CGAGACGTTG CTCTTCACGT CCGAGTCTGT GAACGAGGGC	120
CATCCCGACA AGCTCTGTGA CCAAGTCTCC GACGCCGTCT TGGATGCCCTG CTTGGCCAG	180
GATGCCGACA GCAAGGTCGC CTGCGAGACC GTCACCAAGA CCAACATGGT CATGGTCTTG	240
GGCGAGATCA CCACCAAGGC CACCGTCGAC TATGAGAAGA TCGTGCCTGAA CACCTGCCGC	300
AACATCGGTT TCATCTCTGA TGACGTTGGT CTCGACGCCG ACCGTTGCAA RGTGCTCGTC	360
AACATCGAGC AGCAGTCCCC TGACATTGCC CAGGGTGTTC ATGGACACTT CACCAAGCGT	420

CCCGAAGAAG	TCGGCCCGG	TGACCAGGGC	ATCATGTTG	GCTATGCCAC	CGATGAGACC	480
CCTGAGCTGA	TGCCCCTCAA	GCACGTGCTT	GCCACCAAGC	TYGGAGCTCG	CCTCACSGAG	540
GTCCGCAAGA	ATGGCACCTG	CGCCTGGTC	AGGCCTGACG	GAAAGACCCA	GGTCACAGTC	600
GAGTACCTAA	ACGAGGATGG	TGCCATGGTA	CCTGTTGCTG	TGCACACCGT	CCTCATCTCC	660
ACCCAGCACG	ACGAGACCGT	CACCAACGAC	GAGATTGCTG	CGGACCTCAA	GGAGCATGTC	720
ATCAAGCCGG	TGATCCCCGC	AAAGTACCTC	GATGAGAACAA	CCATCTTCCA	CCTGAACCCG	780
TCTGGCCGCT	TCGTCATCGG	CGGCCCCAC	GGTGACGCCG	GTCTCACCGG	CCGCAAGATC	840
ATCATCGACA	CCTATGGTGG	CTGGGGAGCC	CACGGCCGCG	GTGCCCTCTC	TGGCAAGGAC	900
CCAACCAAGG	TCGACCGYAG	TGGCGCCTAC	ATTGCCAGGC	ARGCCGCCAA	GAGCATCATC	960
GCCAGCGGCC	TCGCACGCCG	CTGCATTGTG	CAGATCTCAT	ACGCCATCGG	TGTGCCTGAG	1020
CCTTTGTCTG	TGTTCGTCGA	CTCCTACGGC	ACCGGCAAGA	TCCCCGACAG	GGAGATCCTC	1080
AAGCTCGTGA	AGGAGAACTT	TGACTTCAGG	CCCGGGATGA	TCAGCATCAA	CCTGGACTTG	1140
AAGAAAGGTG	GAAACAGGTT	CATCAAGACC	GCTGCTTACG	GTCACTTTGG	CCGTGATGAT	1200
GCCGACTTCA	CCTGGGAGGT	GGTGAAGCCC	CTCAAGTTCG	ACAAGGCATC	TGCCTAAGAG	1260
CATGGCATTTC	TCTTGGTCTG	CCGCCTCTCA	AGTTCGTCAA	GACGGGATCA	TGTTGCTCCT	1320
GGGAAGTGGG	AAGAACATT	AGACATTGAA	GCGACGCTCT	ACACTGGTCT	TGTTGTATGG	1380

(2) INFORMATION FOR SEQ ID NO:42:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 394 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

Met	Ala	Ala	Glu	Thr	Phe	Leu	Phe	Thr	Ser	Glu	Ser	Val	Asn	Glu	Gly
1						5			10					15	
His	Pro	Asp	Lys	Leu	Cys	Asp	Gln	Val	Ser	Asp	Ala	Val	Leu	Asp	Ala
								20		25			30		
Cys	Leu	Ala	Gln	Asp	Ala	Asp	Ser	Lys	Val	Ala	Cys	Glu	Thr	Val	Thr
								35		40			45		
Lys	Thr	Asn	Met	Val	Met	Val	Leu	Gly	Glu	Ile	Thr	Thr	Lys	Ala	Thr
							50		55		60				
Val	Asp	Tyr	Glu	Lys	Ile	Val	Arg	Asp	Thr	Cys	Arg	Asn	Ile	Gly	Phe
							65		70		75		80		
Ile	Ser	Asp	Asp	Val	Gly	Leu	Asp	Ala	Asp	Arg	Cys	Lys	Val	Leu	Val
								85		90			95		
Asn	Ile	Glu	Gln	Gln	Ser	Pro	Asp	Ile	Ala	Gln	Gly	Val	His	Gly	His
								100		105			110		

Phe Thr Lys Arg Pro Glu Glu Val Gly Ala Gly Asp Gln Gly Ile Met
 115 120 125
 Phe Gly Tyr Ala Thr Asp Glu Thr Pro Glu Leu Met Pro Leu Lys His
 130 135 140
 Val Leu Ala Thr Lys Leu Gly Ala Arg Leu Thr Glu Val Arg Lys Asn
 145 150 155 160
 Gly Thr Cys Ala Trp Val Arg Pro Asp Gly Lys Thr Gln Val Thr Val
 165 170 175
 Glu Tyr Leu Asn Glu Asp Gly Ala Met Val Pro Val Arg Val His Thr
 180 185 190
 Val Leu Ile Ser Thr Gln His Asp Glu Thr Val Thr Asn Asp Glu Ile
 195 200 205
 Ala Ala Asp Leu Lys Glu His Val Ile Lys Pro Val Ile Pro Ala Lys
 210 215 220
 Tyr Leu Asp Glu Asn Thr Ile Phe His Leu Asn Pro Ser Gly Arg Phe
 225 230 235 240
 Val Ile Gly Gly Pro His Gly Asp Ala Gly Leu Thr Gly Arg Lys Ile
 245 250 255
 Ile Ile Asp Thr Tyr Gly Gly Trp Gly Ala His Gly Gly Ala Phe
 260 265 270
 Ser Gly Lys Asp Pro Thr Lys Val Asp Arg Ser Gly Ala Tyr Ile Ala
 275 280 285
 Arg Gln Ala Ala Lys Ser Ile Ile Ala Ser Gly Leu Ala Arg Arg Cys
 290 295 300
 Ile Val Gln Ile Ser Tyr Ala Ile Gly Val Pro Glu Pro Leu Ser Val
 305 310 315 320
 Phe Val Asp Ser Tyr Gly Thr Gly Lys Ile Pro Asp Arg Glu Ile Leu
 325 330 335
 Lys Leu Val Lys Glu Asn Phe Asp Phe Arg Pro Gly Met Ile Scr Ile
 340 345 350
 Asn Leu Asp Leu Lys Lys Gly Asn Arg Phe Ile Lys Thr Ala Ala
 355 360 365
 Tyr Gly His Phe Gly Arg Asp Asp Ala Asp Phe Thr Trp Glu Val Val
 370 375 380
 Lys Pro Leu Lys Phe Asp Lys Ala Ser Ala
 385 390

(2) INFORMATION FOR SEQ ID NO:43:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1353 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Hordeum vulgare

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

GAATTCCGGA TAGCATCAGC ACAACTGCAC GAGAGCATCT CTACCACCAA AGAAATGGCG	60
GCCGAGACGT TCCTCTTCAC GTCCGAGTCC GTGAACGGAGG GCCATCCCGA CAAGCTGTGC	120
GACCAGGTCT CTGACGCCGT CTTGGACGCC TGCTTGGCCC AGGATCCTGA CAGCAAGGTT	180
GCTTGCAGAGA CCTGCACCAA GACCAACATG GTCACTGGTCT TCGGCAGAGAT CACCAACCAAG	240
GCCACCGTTG ACTATGAGAA GATTGTGCGC GACACCTGCC GTGACATCGG CTTCATCTCT	300
GACGACGTCG GTCTCGATGC CGACCAATTGC AAGGTGCTCG TCAACATCGA GCAGCAATCC	360
CCTGACATTG CCCAGGGTGT TCACGGACAC TTCACCAAGC GTCCAGAAGA GGTCGGGCC	420
GGTGACCAGG GCATCATGTT TGGCTACGCC ACTGATGAGA CCCCTGAGCT GATGCCCTC	480
ACCCACATGC TTGCCACCAA GCTCGGAGCT CGCCTCACCG AGGTCCGCAA GAATGGCACC	540
TGCGCCTGGC TCAGGCCTGA TGGAAAGACC CAGGTACCA TTGAGTACCT AAACGAGGGT	600
GGTGCCATGG TGCCCGTTCG TGTGCACACC GTCTCATCT CCACCCAGCA TGATGAGACC	660
GTCACCAACG ATGAGATCGC TGCAGACCTC AAGGAGCATG TCATCAAGCC GGTGATTCCC	720
GGGAAGTACC TCGATGAGAA CACCATCTTC CACCTGAACC CATCGGGCCG CTTTGTACATC	780
GGTGGCCCTC ACGGCGATGC CGGTCTCACC GCCCCAAGA TCATCATCGA CACCTATGGT	840
GGCTGGGGAG CCCACGGCGG CGGTGCCTTC TCTGGCAAGG ACCCTACCAA GGTCGACCGC	900
AGTGGCGCCT ACATTGCCAG GCAGGCTGCC AAGAGCATCA TCGCCAGCGG CCTCGCACGC	960
CGGTGCATTG TGCAGATCTC ATATGCCATC GGTGTACCTG AGCCTTGTC TGTGTTCGTC	1020
GACTCCTACG GCACTGGCAA GATCCCTGAC AGGGAGATCC TCAAGCTCGT GAAGGAGAAC	1080
TTTGACTTCA GACCCGGGAT GATCACGATC AACCTCGACT TGAAGAAAGG TGGAAACAGG	1140
TTCATCAAGA CAGCTGCTTA CGGTCACTTT GGCGCGATG ATGCTGACTT CACCTGGGAG	1200
GTGGTGAAGC CCCTCAAGTT CGACAAGGCA TCTGCTTAAG AAGAAGACAT CACATTGAGG	1260
GTTCTTCTTG GTCTGATGCC TCTCAAGTTC GGCAAGGCAG GATCCTTTG CTCCCTGGAA	1320
GTAAGAAGAA GCATTCAACA TCGCCCGGAA TTC	1353

CLAIMS

What is claimed is:

1. An isolated nucleic acid fragment encoding all or a substantial portion of a plant dihydropicolinate reductase comprising a member selected from the group consisting of:
 - 5 (a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:2 and 4;
 - (b) an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:2 and 4; and
 - (c) an isolated nucleic acid fragment that is complementary to (a) or (b).
- 10 2. The isolated nucleic acid fragment of Claim 1 wherein the nucleotide sequence of the fragment comprises all or a portion of the sequence set forth in a member selected from the group consisting of SEQ ID NO:1 and 3.
- 15 3. A chimeric gene comprising the nucleic acid fragment of Claim 1 operably linked to suitable regulatory sequences.
4. A transformed host cell comprising the chimeric gene of Claim 3.
- 20 5. A dihydropicolinate reductase polypeptide comprising all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:2 and 4.
6. An isolated nucleic acid fragment encoding all or a substantial portion of a plant diaminopimelate epimerase comprising a member selected from the group consisting of:
 - 25 (a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:7, 9, 11, and 13;
 - (b) an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:7, 9, 11, and 13; and
 - (c) an isolated nucleic acid fragment that is complementary to (a) or (b).
- 30 7. The isolated nucleic acid fragment of Claim 6 wherein the nucleotide sequence of the fragment comprises all or a portion of the sequence set forth in a member selected from the group consisting of SEQ ID NO:6, 8, 10, and 12.
8. A chimeric gene comprising the nucleic acid fragment of Claim 6 operably linked to suitable regulatory sequences.
- 35 9. A transformed host cell comprising the chimeric gene of Claim 8.

10. A diaminopimelate epimerase polypeptide comprising all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO: 7, 9, 11, and 13.

11. An isolated nucleic acid fragment encoding all or a substantial portion of a

5 plant threonine synthase comprising a member selected from the group consisting of:

(a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:16 and 18;

(b) an isolated nucleic acid fragment that is substantially similar to an

10 isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:16 and 18; and

(c) an isolated nucleic acid fragment that is complementary to (a) or (b).

12. The isolated nucleic acid fragment of Claim 11 wherein the nucleotide

15 sequence of the fragment comprises all or a portion of the sequence set forth in a member selected from the group consisting of SEQ ID NO:15 and 17.

13. A chimeric gene comprising the nucleic acid fragment of Claim 11 operably linked to suitable regulatory sequences.

14. A transformed host cell comprising the chimeric gene of Claim 13.

20 15. A threonine synthase polypeptide comprising all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:16 and 18.

16. An isolated nucleic acid fragment encoding all or a substantial portion of a plant threonine synthase comprising a member selected from the group consisting of:

25 (a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:20;

(b) an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:20; and

30 (c) an isolated nucleic acid fragment that is complementary to (a) or (b).

17. The isolated nucleic acid fragment of Claim 16 wherein the nucleotide sequence of the fragment comprises all or a portion of the sequence set forth in SEQ ID NO:19.

35 18. A chimeric gene comprising the nucleic acid fragment of Claim 16 operably linked to suitable regulatory sequences.

19. A transformed host cell comprising the chimeric gene of Claim 18.

20. A threonine synthase polypeptide comprising all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:20.

21. An isolated nucleic acid fragment encoding all or a substantial portion of a plant threonine synthase comprising a member selected from the group consisting of:

- (a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:22 and 24;
- (b) an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:22 and 24; and
- (c) an isolated nucleic acid fragment that is complementary to (a) or (b).

5 22. The isolated nucleic acid fragment of Claim 21 wherein the nucleotide sequence of the fragment comprises all or a portion of the sequence set forth in a member selected from the group consisting of SEQ ID NO:21 and 23.

10 23. A chimeric gene comprising the nucleic acid fragment of Claim 21 operably linked to suitable regulatory sequences.

15 24. A transformed host cell comprising the chimeric gene of Claim 23.

20 25. A threonine synthase polypeptide comprising all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:22 and 24.

25 26. An isolated nucleic acid fragment encoding all or a substantial portion of a plant threonine synthase comprising a member selected from the group consisting of:

- (a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:26;
- (b) an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:26; and
- (c) an isolated nucleic acid fragment that is complementary to (a) or (b).

30 27. The isolated nucleic acid fragment of Claim 26 wherein the nucleotide sequence of the fragment comprises all or a portion of the sequence set forth in SEQ ID NO:25.

35 28. A chimeric gene comprising the nucleic acid fragment of Claim 26 operably linked to suitable regulatory sequences.

29. A transformed host cell comprising the chimeric gene of Claim 28.

30 30. A threonine synthase polypeptide comprising all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:26.

35 31. An isolated nucleic acid fragment encoding all or a substantial portion of a plant threonine deaminase comprising a member selected from the group consisting of:

- (a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:29;

- (b) an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:29; and
- (c) an isolated nucleic acid fragment that is complementary to (a) or (b).

5 32. The isolated nucleic acid fragment of Claim 31 wherein the nucleotide sequence of the fragment comprises all or a portion of the sequence set forth in SEQ ID NO:28.

33. A chimeric gene comprising the nucleic acid fragment of Claim 31 operably linked to suitable regulatory sequences.

10 34. A transformed host cell comprising the chimeric gene of Claim 33.

35. A threonine deaminase polypeptide comprising all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:29.

36. An isolated nucleic acid fragment encoding all or a substantial portion of a plant threonine deaminase comprising a member selected from the group consisting of:

- (a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:31 and 33;
- (b) an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:31 and 33; and
- (c) an isolated nucleic acid fragment that is complementary to (a) or (b).

20 37. The isolated nucleic acid fragment of Claim 36 wherein the nucleotide sequence of the fragment comprises all or a portion of the sequence set forth in a member selected from the group consisting of SEQ ID NO:30 and 32.

25 38. A chimeric gene comprising the nucleic acid fragment of Claim 36 operably linked to suitable regulatory sequences.

39. A transformed host cell comprising the chimeric gene of Claim 38.

40. A threonine deaminase polypeptide comprising all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:31 and 33.

41. An isolated nucleic acid fragment encoding all or a substantial portion of a plant S-adenosylmethionine synthetase wherein the nucleotide sequence of the fragment comprises all or a portion of the sequence set forth in SEQ ID NO:35.

35 42. A chimeric gene comprising the nucleic acid fragment of Claim 41 operably linked to suitable regulatory sequences.

43. A transformed host cell comprising the chimeric gene of Claim 42.

44. An isolated nucleic acid fragment encoding all or a substantial portion of a plant S-adenosylmethionine synthetase wherein the nucleotide sequence of the fragment comprises all or a portion of the sequence set forth in SEQ ID NO:38.

5 45. A chimeric gene comprising the nucleic acid fragment of Claim 44 operably linked to suitable regulatory sequences.

46. A transformed host cell comprising the chimeric gene of Claim 45.

47. An isolated nucleic acid fragment encoding all or a substantial portion of a plant S-adenosylmethionine synthetase wherein the nucleotide sequence of the fragment comprises all or a portion of the sequence set forth in SEQ ID NO:41.

10 48. A chimeric gene comprising the nucleic acid fragment of Claim 47 operably linked to suitable regulatory sequences.

49. A transformed host cell comprising the chimeric gene of Claim 48.

50. A method of altering the level of expression of a plant amino acid biosynthetic enzyme in a host cell comprising:

15 (a) transforming a host cell with the chimeric gene of any of Claims 3, 8, 13, 18, 23, 28, 33, 38, 42, 45, and 48; and
(b) growing the transformed host cell produced in step (a) under conditions that are suitable for expression of the chimeric gene

wherein expression of the chimeric gene results in production of altered levels of a plant 20 amino acid biosynthetic enzyme in the transformed host cell.

51. A method of obtaining a nucleic acid fragment encoding all or substantially all of the amino acid sequence encoding a plant amino acid biosynthetic enzyme comprising:

25 (a) probing a cDNA or genomic library with the nucleic acid fragment of any of Claims 1, 6, 11, 16, 21, 26, 31, 36, 41, 44, and 47;
(b) identifying a DNA clone that hybridizes with the nucleic acid fragment of any of Claims 1, 6, 11, 16, 21, 26, 31, 36, 41, 44, and 47;
(c) isolating the DNA clone identified in step (b); and
(d) sequencing the cDNA or genomic fragment that comprises the clone isolated in step (c)

30 wherein the sequenced nucleic acid fragment encodes all or substantially all of the amino acid sequence encoding a plant amino acid biosynthetic enzyme.

52. A method of obtaining a nucleic acid fragment encoding a portion of an amino acid sequence encoding a plant amino acid biosynthetic enzyme comprising:

35 (a) synthesizing an oligonucleotide primer corresponding to a portion of the sequence set forth in any of SEQ ID NOs:1, 3, 6, 8, 10, 12, 15, 17, 19, 21, 23, 25, 28, 30, 32, 35, 38, and 41; and
(b) amplifying a cDNA insert present in a cloning vector using the oligonucleotide primer of step (a) and a primer representing sequences of the cloning vector

wherein the amplified nucleic acid fragment encodes a portion of an amino acid sequence encoding a plant amino acid biosynthetic enzyme.

53. The product of the method of Claim 51.

54. The product of the method of Claim 52.

5 55. A method for evaluating at least one compound for its ability to inhibit the activity of a plant biosynthetic enzyme selected from the group consisting of dihydrodipicolinate reductase, diaminopimelate epimerase, threonine synthase, threonine deaminase and S-adenosylmethionine synthetase, the method comprising the steps of:

10 (a) transforming a host cell with a chimeric gene comprising a nucleic acid fragment encoding a plant biosynthetic enzyme selected from the group consisting of dihydrodipicolinate reductase, diaminopimelate epimerase, threonine synthase, threonine deaminase and S-adenosyl-methionine synthetase, operably linked to suitable regulatory sequences;

15 (b) growing the transformed host cell under conditions that are suitable for expression of the chimeric gene wherein expression of the chimeric gene results in production of the biosynthetic enzyme encoded by the operably linked nucleic acid fragment in the transformed host cell;

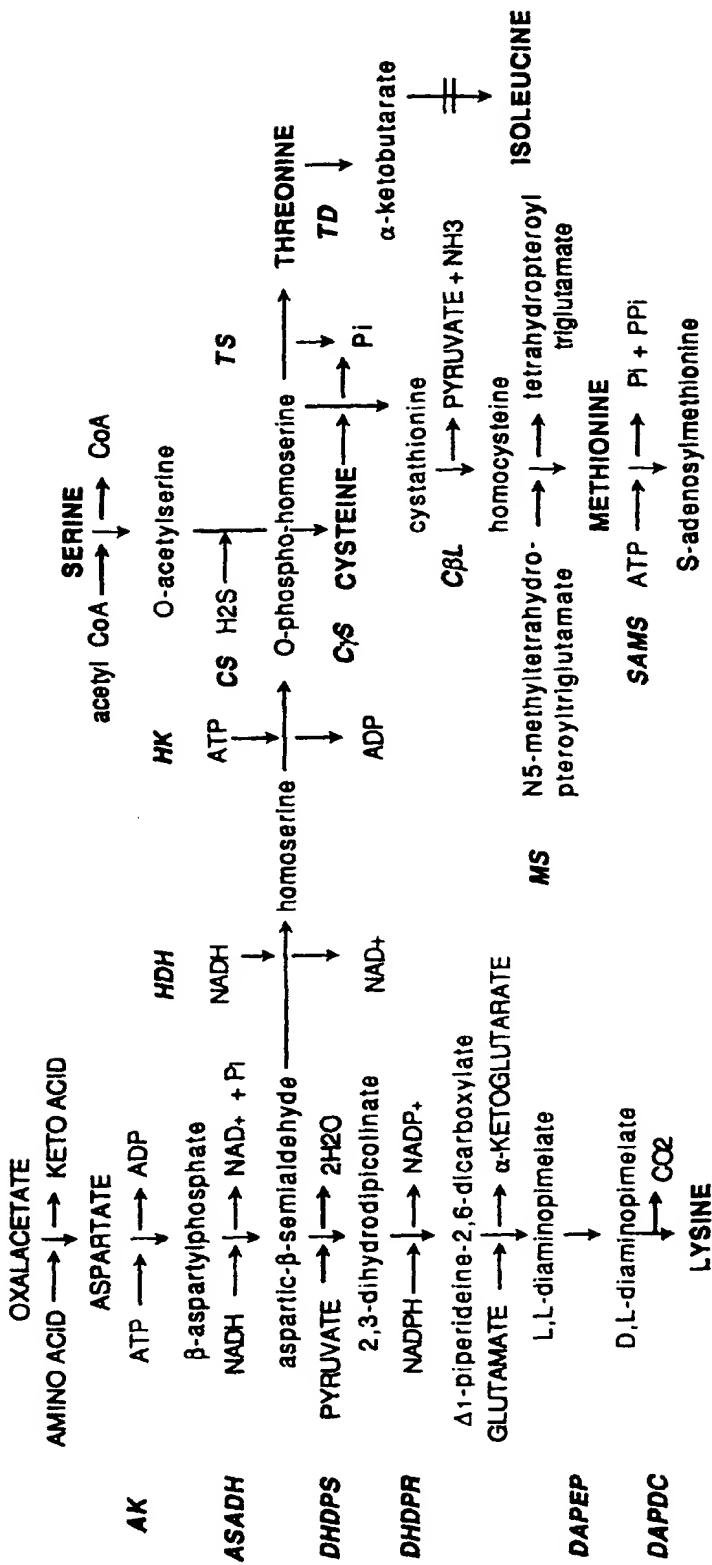
20 (c) optionally purifying the biosynthetic enzyme expressed by the transformed host cell;

(d) treating the biosynthetic enzyme with a compound to be tested; and

(e) comparing the activity of the biosynthetic enzyme that has been treated with a test compound to the activity of an untreated biosynthetic enzyme,

25 thereby selecting compounds with potential for inhibitory activity.

FIG. 1



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FIG. 2

SEQ ID NO: 4	NO: 4	1	60
SEQ ID NO: 2	NO: 2		KIGRRNAA.
SEQ ID NO: 5	NO: 5		AGQISGMD.EPLEI
			MANQDLIPVVNGAAGKMGREVIKAVAQARPDLQLVGAUDHNPSLQQDIGEVVGIAPLEV
		61	
SEQ ID NO: 4	NO: 4	1.20	
SEQ ID NO: 2	NO: 2		KVVIIGATKEIGRTAIIAVSKARGMELAGAID.
SEQ ID NO: 5	NO: 5		PVLNDLTMVLSIAQSRAATGVVVDFSEPSAVYDNVKQAAAFGLSSVVYPKIELETVTTEL
			PVLADLQSVLVLAQEIKIQGVMDFTHPSCGVYDNVRSAIAYGVRPVVGTGSEQQIQDL
		1.21	
SEQ ID NO: 4	NO: 4	180	
SEQ ID NO: 2	NO: 2		S.QCI. QCLAGEI.
SEQ ID NO: 5	NO: 5		SGMGRTLEIP.
			SAFCEKAS.GCLVAPTLSIGSVLQQAAIQASFHYSNVEIVESRPNP.SDLPSQDRIQIA
			GDFAEKASTGCLIAPIFNAGVILMQAAVQACQYFDHVEELHNQKADAPSGTAIKTA
		181	
SEQ ID NO: 4	NO: 4	240	
SEQ ID NO: 2	NO: 2		..LNDLTMV. LGSIAQTRA.
SEQ ID NO: 5	NO: 5		TGVV. VDFSEPSTVYD
			NNISDLGQIYNR. EDMDSSSPARGOLLEDGVVRVHSMVLPGLVSSTSINFSGPGEMYT
			QMLAEMGKTENPPAVEEERETIAGAKGGL. GPGQIPIHSIRLPGLIAHQEVLFGSPGQLYT
		241	276
SEQ ID NO: 4	NO: 4		NVKQA.
SEQ ID NO: 2	NO: 2		LRHDVANVQCLMPGLLAIRKVRFRNLIYGLEKFL
SEQ ID NO: 5	NO: 5		IRHDTTDRACYMPGVLLGIRKVELKGLVYGLEKLL

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FIG. 3

1	SEQ ID NO: 7	L...	60
SEQ ID NO: 9	VS...
SEQ ID NO: 11	MAITATISVPLTSRRTLTSVNLSPLSTRSTLPTPORTFKYPNSRSLVSSMSTETAVK
SEQ ID NO: 13
SEQ ID NO: 14
61	SEQ ID NO: 7	120
SEQ ID NO: 9
SEQ ID NO: 11	TSSASFNLNRKESGFLHFAKYHGLGNDFVLIDNRDSSSEP	KISAEKAVQLCDORNFGVGADGV
SEQ ID NO: 13
SEQ ID NO: 14
121	SEQ ID NO: 7	PEMCNGNGVRCFARFIAELENLOGTNRFTIHTGAGKIV	180
SEQ ID NO: 9	IFVMPGVNGADDTYMRIFNSDGSPEPMCNGNGVRCFARFIAELENLOGTNSFKIHTGAGLII
SEQ ID NO: 11	IFVLPGISGTDYMRIFNSDGSPEPMCNGNGVRCFAKEVSQLENLHGRHSFTIHTGAGLII
SEQ ID NO: 13	IFVLPGVNGADDTYMRIFNSDGSNRYWYK.GFV
SEQ ID NO: 14	IFALPGQQGTDYMRIFNSDGSPEPMCNGNGIRCLAKFLADLEGEEK.TYRIHTLAGVIT
181	SEQ ID NO: 7	PEIQSDGQVKVDMGEPLISGLDIPTRKLLATKNAVVAELAVEGLTWHVTCVSMGNPHCV	240
SEQ ID NO: 9	PEIQONDGKVVKVDMGQPLAC
SEQ ID NO: 11	PEVLEDGNVRVDMGEPLVPLAKDVTPLPANKDNAYVKSQVVDGVVHVTCVSMGNPHCV
SEQ ID NO: 13
SEQ ID NO: 14	PQLLADGQVKVDMGEPLQLLAELIPTLAPAGEK.VVDLPLAVAGQTWAVTCVSMGNPHCL

FIG. 3 (Continued)

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SEQ	ID NO: 7	TEGANELKVQVDDKLSEIGPKFEHHEMFPARTNTTEFQVLSSRSHLKMVRVWERGAGATL
SEQ	ID NO: 9
SEQ	ID NO: 11	TESREGSQNLVDELKLAEIGPKFEHHEVFPARTNTTEFQVLSSNHLKMVRVWERGAGATL
SEQ	ID NO: 13
SEQ	ID NO: 14	TFVDD.....VDSLNLTEIGPLFEHHPQFSQRTNTTEFIQVLGSDRLKMVRVWERGAGITL

300

SEQ	ID NO: 7	ACGTGACATVVAAVLEGRAERKCVVDPGGPLEIEWREDDNHVYMTGPAEVVFYGSVVFH
SEQ	ID NO: 9
SEQ	ID NO: 11	ACGTGACATVVAAVLEGRAGRNCTVDPGGPLQIEWREDDNHVYMTGSAADVYYGSLPL
SEQ	ID NO: 13
SEQ	ID NO: 14	ACGTGACATVVAAVLITGRGDRRCTVELPGGNLEIWSAQDNRLYMTGPAQRVFGQAEI

301

SEQ	ID NO: 7	ACGTGACATVVAAVLEGRAERKCVVDPGGPLEIEWREDDNHVYMTGPAEVVFYGSVVFH
SEQ	ID NO: 9
SEQ	ID NO: 11	ACGTGACATVVAAVLEGRAGRNCTVDPGGPLQIEWREDDNHVYMTGSAADVYYGSLPL
SEQ	ID NO: 13
SEQ	ID NO: 14	ACGTGACATVVAAVLITGRGDRRCTVELPGGNLEIWSAQDNRLYMTGPAQRVFGQAEI

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SEQ	ID NO: 7	ACGTGACATVVAAVLEGRAERKCVVDPGGPLEIEWREDDNHVYMTGPAEVVFYGSVVFH
SEQ	ID NO: 9
SEQ	ID NO: 11	ACGTGACATVVAAVLEGRAGRNCTVDPGGPLQIEWREDDNHVYMTGSAADVYYGSLPL
SEQ	ID NO: 13
SEQ	ID NO: 14	ACGTGACATVVAAVLITGRGDRRCTVELPGGNLEIWSAQDNRLYMTGPAQRVFGQAEI

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FIG. 4

1	SEQ ID NO: 24	ASSSLFQSLPPFLQTSK.PYAPPKPAAHF'VRA.....QSPLTQNNNNSSSKHRRPAD	120
2	SEQ ID NO: 25	
3	SEQ ID NO: 27	
4	SEQ ID NO: 20	61
5	SEQ ID NO: 22	
6	SEQ ID NO: 24	ENIRDEARRINAPHDHHILESAKYVPFNA.....MENGAATNGASEKSHSPS	
7	SEQ ID NO: 26	
8	SEQ ID NO: 27	DNIRDEARR.NRSNAVNPFSAKYVPFNA.....APGSTESYSLDEIVYRSRGGLDVEHDM	
9	SEQ ID NO: 13	121
10	SEQ ID NO: 20	
11	SEQ ID NO: 13	
12	SEQ ID NO: 20	QTYLSTRGDCIGLSFET'V	
13	SEQ ID NO: 22	
14	SEQ ID NO: 24	DALKRFDGEYWRNLFDLSRVGKTTWPGSGVWSKKKEWVLPEIHDDEIVSAFEGNNSNLFWAE	
15	SEQ ID NO: 26	
16	SEQ ID NO: 27	EALKRFDGAYWRDLFDSRVGKSTWPGSGVWSKKKEWVLPEIDDDIVSAFEGNNSNLFWAE	
17	SEQ ID NO: 16	181
18	SEQ ID NO: 18	
19	SEQ ID NO: 20	LKGGLAADGGLFLPEEVPRATEQSWKDLPYTELAKVY	
20	SEQ ID NO: 22	
21	SEQ ID NO: 24	RGFKQFLGMNDLWVKHCGISHTGSEFKDLMGTVLVSQVNRLRERMNRPVVGCASTGDTSA	
22	SEQ ID NO: 26	
23	SEQ ID NO: 27	RGFKQFLGMNDLWVKHGGISHTGSEFKDLMGTVLVSQVNRLRERMNRPVVGCASTGDTSA	

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FIG. 4 (Continued)

300	241	SEQ ID NO: 16 SEQ ID NO: 18 SEQ ID NO: 20 SEQ ID NO: 22 SEQ ID NO: 24 SEQ ID NO: 26 SEQ ID NO: 27	ALSAAYCAAAGGPAIVFLEADRISLQQLIQPIANGATVLSLDTDFGCMRLIREVTAELPI
360	301	SEQ ID NO: 16 SEQ ID NO: 18 SEQ ID NO: 20 SEQ ID NO: 22 SEQ ID NO: 24 SEQ ID NO: 26 SEQ ID NO: 27	YLANSLNPL.RLEGQKTAIEIILQQFNWQVPDWVIVPGGNLGNIIYAFYKGFMCRVGLV
420	361	SEQ ID NO: 16 SEQ ID NO: 18 SEQ ID NO: 20 SEQ ID NO: 22 SEQ ID NO: 24 SEQ ID NO: 26 SEQ ID NO: 27	DRVPRLVCAQQAANANPPLYKSGWTEFEPQTAETTFSAAIQIGDPPSVDRAVVVALKATD
480	421	SEQ ID NO: 16 SEQ ID NO: 18 SEQ ID NO: 20 SEQ ID NO: 22 SEQ ID NO: 24	KDCALQFLGNLXKEYF..... DK1PRLVCAQQAANADPPLYLYFKSGWKEFKPVKSSTTFSAAIQIGDPPSIDRAVHALKS CD DRIPRMVCAQQAANANPPLYHYSKGMKDFKPMTASTTFASAAIQIGDPPSIDRAVYALKKC N
540	482	SEQ ID NO: 16 SEQ ID NO: 18 SEQ ID NO: 20 SEQ ID NO: 22 SEQ ID NO: 24	GIVEEATEEEELMDATAADRGMFACPHTGVALAALFKLOGQRIIGPNDRTVVVSTA HGL

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FIG. 4 (Continued)

SEQ ID NO: 27	GIVEEATEEELMDAMAQADSTGMFICPFHTGVALTAFLKLRNQGVIAPTDRTVYVSTAHGL	
	481	537
SEQ ID NO: 16	KFTQSKIDYHDKNIKDMVCCQYANPPISVKADFGSVMMDVLQKN.....LNGKI.....	
SEQ ID NO: 18MACKYSNPPVSVKADEFGAVMMDVLKKR.....LKGKL.....	
SEQ ID NO: 20	
SEQ ID NO: 22	KFAQSKIDYHSGLIPGMG.RYANPLVSVKADFGSVMMDVLKDSCTTSPPPTLTSLDVAK.....	
SEQ ID NO: 24	KFTQSKIDYHSKDIKDMACRYANPPMQVKADFGSVMMDVLKTY.....LQSKA..H.....	
SEQ ID NO: 26	
SEQ ID NO: 27	KFTQSKIDYHSNAIPDMACRFSSNPFDVKADEFGAVMMDVLKSY.....LGSNTLTS.....	

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FIG. 5

SEQ	ID	NO: 29	1	60
SEQ	ID	NO: 31
SEQ	ID	NO: 33
SEQ	ID	NO: 34	MASHDYLKKILTA RYDVAFETELEPAR NLSARLRNPVYLK REDNQPVFSFKL RGAYNKM
SEQ	ID	NO: 29	61	120
SEQ	ID	NO: 31
SEQ	ID	NO: 33
SEQ	ID	NO: 34	AHIPADALRGVIT ASAGNHAQGVAF SAARMGVKA VIVV PVTT PQVKV DAVRAH HGGPGVE
SEQ	ID	NO: 29	121	130
SEQ	ID	NO: 31	SYDEAQSYAK.....	SYDEAQSYAK.....
SEQ	ID	NO: 33
SEQ	ID	NO: 34	VIQAGESYS DAYAHALKV QEERGLTFV HFPFDDPY VIAGQGT TIAIMEI ILRQHQGP IHAIFV P
SEQ	ID	NO: 29	181	240
SEQ	ID	NO: 31	VGGGGGLIAGIA AYV KVR PEVK IIG VE PS D A N A M A L S L C H G K R V M L E H V G G F A D G V A K A
SEQ	ID	NO: 33
SEQ	ID	NO: 34	IGGGGLAAGV A A Y V K A R P E I K V I G Q A E D S C A M A Q S L Q A G K R V E L A E V G L F A D G T A V K L
SEQ	ID	NO: 29	241	300
SEQ	ID	NO: 31	VGEETFR LCREL V D G I V M V S R D A I C A S I K D M F E E K R S I L E P A G A L A G A E A Y C K Y N L K
SEQ	ID	NO: 33
SEQ	ID	NO: 34	VGEETFR LCREL V D G I V M V S R D A I C A S I K D M F E E K R S I L E P A G A L A G A E A Y C K Y N L K

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FIG. 5 (Continued)

301	SEQ ID NO: 29	GETVVAITSGANMNFDRLRLVTELADVGKREAVLATFLPERQSFKKFTELVGRMNITE	360
	SEQ ID NO: 31	..NIVAITSGANMNFDKLRLVVTTELAVGRKQEAVLATVMAEEPGSFKFQFCELVGQMNITE	
	SEQ ID NO: 33	
	SEQ ID NO: 34	NQTLVAVITSGANMNFDRLRFVAAERAEGEAREAVFAVTPEERGSFKRFCSLVGDRNVE	
361	SEQ ID NO: 29	FKYRYDSNAKDALVLYSVGIFTYTDNELGAMMDRMEASAKLRTVNLTNDLAKDHLYFIGGR	420
	SEQ ID NO: 31	FKYRYNSNEK.AVVLYSVGVHTISELRAMQERMESSQLKTYNLTESDLVKDHLYLMGGR	
	SEQ ID NO: 33	
	SEQ ID NO: 34	ENYRI.ADAQOSAHIFVGVQIRRGEASDIAANFEFSGEFTADLTHDELSKEHRYMVGGR	
421	SEQ ID NO: 29	SEIK.DELVYRFIFPERPGALMKFLDTFSPRWNISLFLHYRAQGEAGANVLGIQVPPAEF	480
	SEQ ID NO: 31	SNVQ.NEVFVVSPXPXPRKTGALMKFLDXFSPRWDISL.....	
	SEQ ID NO: 33RPGALMKFLDPFSPRWNISLFLHYRGEGETGANVLGIQVPKSEM	
	SEQ ID NO: 34	SPLALDERLRFEEPERPGALMKFLSSMAPDWNISLFLHYRNQGADYSSILVGLQVPQADH	
481	SEQ ID NO: 29	DEFKSHANNLGYEYMSEHNNIEYRLLLROPKV	512
	SEQ ID NO: 31	
	SEQ ID NO: 33	DEFHDRAANKLGDYDKVNNNDDDFQLLMH.....	
	SEQ ID NO: 34	AEFERFLAALGYPYVEESANPAYRLFLS.....	

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FIG. 6

SEQ ID NO: 36	126	GCAGATCAAAGAAGATGGCAGGCTCTGACACCTTCACCTCGAGTCGTGAACG	185
SEQ ID NO: 37	774	GCAGATAGGAGAAAGTGGCCACTTGATACCTTCCTCTTACCTCGAGTCGTGAACG	833
SEQ ID NO: 36	186	AGGGACACCCCTGACAAGCTCTGGACCCAGGTCTAGATGCCGTTCTTGACGCTGCCCTTG	245
SEQ ID NO: 37	834	AGGGCCACCCCTGACAAGCTCTGGACCAAGTCTAGATGCTGCTGATGCCCTCG	893
SEQ ID NO: 36	246	CTGAGGACCCCTGACAGCAAGGTTGCTGTGAGACCTGACCAAGACCAACATGGTCATGG	305
SEQ ID NO: 37	894	CCGAGGACCCCTGACAGCAAGGTCGCTGTGAGACCTGACCAAGACAAACATGGTCATGG	953
SEQ ID NO: 36	306	TCTTTGGTGGAGATCACCAAGGCCAATGTGCACTACGAGAAAGATTGTCAGGGAGACCT	365
SEQ ID NO: 37	954	TCTTTGGTGGAGATCACCAAGGCCAAGCTAACGCTAACGTGACTATGAGAGATTGTCAGGGAGACAT	1013
SEQ ID NO: 36	366	GCGCAACACATTGGTTTGTGTCACACGATGTCAGGGCTGACCAAGGTGC	425
SEQ ID NO: 37	1014	GCGTAACATCGGTTTGTGTCAGCTGACATGCTGACACTGCAAGGTGC	1073
SEQ ID NO: 36	426	TGTTGAAACATTTGAGCAGCAGTCGGCTGATATTGCTCAGGGTGTGCAATGGCCACTTCACCA	485
SEQ ID NO: 37	1074	TGTTGAAACATCGAGCAGCAGTCGGCTGACATGCAAGGGTGTGCACTTCACCA	1133
SEQ ID NO: 36	486	AGCGCCCCGAGGAGATTGGAGGCTGGTACACATGTTGGGTATGGCAGCGATG	545
SEQ ID NO: 37	1134	AGGGCCCTGAGGTGATGCCCATGTCCTGCCACATGGTGGTACAGGGACATGTTGGATATGCAACTGATG	1193
SEQ ID NO: 36	546	AGACCCCTGAGGTGATGCCCATGTCCTGCCACATGGTGGTACAGGGACATGTTGGATATGCAACTGATG	605
SEQ ID NO: 37	1194	AGACCCCTGAGGTGATGCCCATGTCCTGCCACATGGTGGTACAGGGACATGTTGGATATGCAACTGATG	1253

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FIG. 6 (Continued)

SEQ ID NO: 36	606	CCGAGGTCCGGCAAAGAACCGGAACTGCCCTGGCTCAGGCCCTGATGGGRAGACCAGGTGA	665
SEQ ID NO: 37	1254	CGGAGGTGGCAAAGAACCGGAACTGCCCTGGCTCAGGCCCTGACGGRAAGCCAAGTGA	1313
SEQ ID NO: 36	666	CAGTCGAGTACCGCAATGAGGTGGTGCATGGTCCCACATCGGTGTCACACCGTCTCA	725
SEQ ID NO: 37	1314	CTGTTAGTACCGAATGAGAGCGGGTCCAGGGTCCCTGTCCGTGTCACACCGTCTCA	1373
SEQ ID NO: 36	726	TCTCCACCCAGCACGGACAGTGGCAATGATGAGATCGCTGACTGAAGGGGC	785
SEQ ID NO: 37	1374	TCTCTACCCAGCATGATGAGACAGTCACCRACGRTGAGATTGCTGACCTGAAGGRC	1433
SEQ ID NO: 36	786	ATGTCATCAAGCCATATCCCTGAGCAGTACCTTGACGAGAGACCATCTTCACCTTA	845
SEQ ID NO: 37	1434	ATGTCATCAAGCCCTGTCATTCGCCGACGATACCTTGATGAGAGACATCTTCATCTTA	1493
SEQ ID NO: 36	846	ACCCATCCGGGGCTTGTCAATTGGGACCTCACGGGAGTGCCTCACTGGCGCA	905
SEQ ID NO: 37	1494	ACCCATCTGGCTCGCTTCGTCATTTGGGGACCTCATGGTGTGCTCACTGGCGGA	1553
SEQ ID NO: 36	906	AGATCATCATGACCTAACCTAACGGTGGCTGGGAGCCATGGGAGCTCACGGTGGCTTCGGCA	965
SEQ ID NO: 37	1554	AGATCATCATGACCTAACCTAACGGTGGCTGGGAGCTCACGGTGGCTTCGGCA	1613
SEQ ID NO: 36	966	AGGACCCAAACCAAGGTGACCGAACGGGCTATGTCGGAGGGCTAGGCCATGGGAGCA	1025
SEQ ID NO: 37	1614	AGGACCCAAACCAAGGTGACCGAACGGTGGCTAACGTCGGCAAGGGAGCTGGCA	1673
SEQ ID NO: 36	1026	TCGTCGCCAGGGCCTTGGCTGGGGGGCAATGTCAGGTGTCCTACGCCATGGGAGCA	1085
SEQ ID NO: 37	1674	TGTTGCTAGTGGCCATTGCTGGGGGGTGCATTGTCAGGTGTCATAGGTATCATAGGCA	1733

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FIG. 6 (Continued)

FIG. 7

SEQ ID NO: 38	80	GAGACATTCCATTACCTCAGACTAGTGAACGAGGACACCTGACAAGCTCTGCGAC	139
SEQ ID NO: 40	123	GAAACTTCTTATTCACCTCAGACTAGTGAACGAGGACACCTGACAAGCTCTGCGAT	182
SEQ ID NO: 38	140	CAAATCTCGATGCTGCTCGACGCTTGCTGACAAGGAGTCACCCAGACAAGCTCTGCGAT	199
SEQ ID NO: 40	183	CAGATCTCTGATGCAAGTCTTGTGACAAGATCCCGAGGAAAGTGGCA	242
SEQ ID NO: 38	200	TGCGAACACATGCAACAAAGACCAACTTGGTATGGTCTCGAGAGATCACCACAAAGGCC	259
SEQ ID NO: 40	243	TGTGAACCTGCAACAAAGACCAACTTGGTATGGTCTTGGTGAAGATCACACAAAGGCT	302
SEQ ID NO: 38	260	AACGTTGACTACGAGAAAGATGTCGCTGACACCTGCAACATGGCTTCGTCACAC	319
SEQ ID NO: 40	303	ATTGTAACATGAGAAAGATTGTCGCTGACACATGGCTAAATATTGGATTGTTCTGAT	362
SEQ ID NO: 38	320	GATGTTGGACTTGATGCTGACAACACTGCAAGGTCCCTGTAACATTGAGCAGGCCCT	379
SEQ ID NO: 40	363	GATGTTGGCTTGATGCTGACAACACTGCAAGGTCCCTGTTACATTGAGCAGCAAAGTCCT	422
SEQ ID NO: 38	380	GATATGCCAGGGTGTGCAAGGCCACCTTACCAAAAGACCCGAGGAATCGGTGCTGGA	439
SEQ ID NO: 40	423	GATATGCTCAAGGTGTCCAGGGCCATCTGACCAAAGCCCCTGAGGAGATGGTGTGGT	482
SEQ ID NO: 38	440	GACCAGGGTCACTGTTGGCTATGCCACGGACGAAACCCCAAGATTGATGCCATTGAGT	499
SEQ ID NO: 40	483	GACCAGGGCCACATGTTGGCTATGCCAACAGATGAGAACCTGAAATTGCTCTCAGT	542
SEQ ID NO: 38	500	CATGTCCTGCAAACTAACCTGGCTCAGCTGGTCTCGTCAACGGAGGTTGCC	559
SEQ ID NO: 40	543	CACGTGCTGCAAACTAACCTGGCTCAGCTGGTCTCGTCAACGGAGGTTGCC	602

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FIG. 7 (Continued)

SEQ ID NO:38	560	CCATGGTTGAGGCCCTGATGGAAAACCCAAAGTGACTGTTGAGTATTACATGACAACGGT	619
SEQ ID NO:40	603	GCCTGGTTGAGGCCCTGATGGCAAGACCCAAAGTTACTGTTGAGTATTACATGACAATGGT	662
SEQ ID NO:38	620	GCCATGGTTCCAGTTCTGTGTCACACTGTGCTTATCTCCACCCAAACATGATGAGCTGTG	679
SEQ ID NO:40	663	GCCATGGTTCCATTAGGGTACACACTGTCTTATCTCCACCCAAACAGTGTGAGCCGTT	722
SEQ ID NO:38	680	ACCAACGACGAAATTGAGCTGACCTCAGGGCATGTGATCAGGCCGTATCCGGAG	739
SEQ ID NO:40	723	ACCAATGAGATTGCCGGACCTTAAGGGCATGTCTCAAAACCTGTATCCCAGAG	782
SEQ ID NO:38	740	AAGTACCTTGATGAGAAAGACCATTTCCACTTGTGACCCCTCTGGCCCTTGTGATTTGGA	799
SEQ ID NO:40	783	AAGTACCTTGATGAGAAATCTATTTCACCTTAACCCATCTGGCCGATTGTATTGGT	842
SEQ ID NO:38	800	GGTCCTCACGGTATGGCTCTACCCGGCAAGGATCATCATGATACTTACGGAGGA	859
SEQ ID NO:40	843	GGACTCTATGGTATGGCTCACTGGCTGTTAAATCATCATGACACTTATGGTGGT	902
SEQ ID NO:38	860	TGGGGTGTCTATGGGGGGCTTCACCGGCCGCAAGGATCCCACCAAGGGTAGGGAGT	919
SEQ ID NO:40	903	TGGGGTGTCTATGGGGGGCTTCACCGGCCGCAAGGACCCAAACCAAGGGTAGGGAGT	962
SEQ ID NO:38	920	GGTCTTACATTGTGAGACAGGTGCTAAGAGCATTGGCAAGGGACTAGCCAGAAGG	979
SEQ ID NO:40	963	GGTCATACTTGTGAGGTGCTGAAAGAGTATCGTAGCTGGACTTGTCTGTAGA	1022
SEQ ID NO:38	980	TGCATTGTGCAAGTGTCTTATGCCATTGGTGTGCCATTGGCTTGTCTGTGAC	1039
SEQ ID NO:40	1023	TGCATCGTGCAGGTATGCCATTGGTGTGCCATTGGCTGAGCCATTGTCTGTGAC	1082

FIG. 7 (Continued)

SEQ ID NO: 38 1040 ACCTATGGCACCGGAAAGTCCATGATAGGAGATTCTCAACATGTGAAGGAGACTTT 1099
 SEQ ID NO: 40 1083 ACCTATGGCACTGGAAAGTCCCTGACAGGGAAATTGTAAGATGTTAAGGAGAACTTT 1142
 SEQ ID NO: 38 1100 GATTTCAGGGCCCGTATGATCTCCATCAACCTTGATCTGATCTCAAGAGGGGGGGAAATACAGG 1159
 SEQ ID NO: 40 1143 GACTTCAGACCTGGAAATGATGTCCATTAACTTGGATTGAAGAGGGGGCAATAGAAGA 1202
 SEQ ID NO: 38 1160 TTCTTGAAAGCTGCTGGCATATGGACACTTCGGCAGAGGACCCCTGACCTTCACATGGAA 1219
 SEQ ID NO: 40 1203 TTCTTGAAAACTGGCTATGGTCACTTGGACGTGATGACCCGATTTCACATGGAA 1262
 SEQ ID NO: 38 1220 GTGGTCAAGGCCCTCAAGTGGGAAAGGCCCTAAGGCATTCACTGCATGTGCTG 1279
 SEQ ID NO: 40 1263 GTTGTCAGGCCCTCAAGTGGAAAGGCCCAAGACTAATAAGTGGCTATGTTTTT 1322
 SEQ ID NO: 38 1280 GGAGTTTTTT 1289
 SEQ ID NO: 40 1323 GTTCTTGTGTT 1332

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8.
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H.

FIG. 8 (Continued)

SEQ ID NO: 42	461	GCTATGCCACCAGATGAGACCCCTGAGCTGATGCCCTCAAGCACGTGCTTGCACCAAGC	520
SEQ ID NO: 43	443	GCTACGCCACTGATGAGACCCCTGAGCTGATGCCCTCACGCCACTGCTTGCACCAAGC	502
SEQ ID NO: 42	521	TYGGAGCTGCCCTCACSGAGTCGCCAGAATGGCACCTGGCTGGGTCAAGCCTGAGC	580
SEQ ID NO: 42	503	TCGGAGCTGCCCTCACCGAGTCGCCAGAATGGCACCTGGCTGGGTCAAGCCTGATG	562
SEQ ID NO: 42	581	GAAGAGCCAGGTACAGTCAGGTACCTAAACAGGGATGGTACCTGTCGTG	640
SEQ ID NO: 43	563	GAAAGACCCAGGTACCATTTAGTACGTACCTAAACAGGGTGGCATGGTGCCTCGTG	622
SEQ ID NO: 42	641	TGCACACCGTCCCATCTCCACCCAGCACGAGACCGTACCAACGACGAGATTGCTG	700
SEQ ID NO: 43	623	TGCACACCGTCCCATCTCCACCCAGCATGAGACCCGTACCAACGATGAGATCGCTG	682
SEQ ID NO: 42	701	CGGACCTCAAGGAGCATGTCATCAAGCCGGTGTATCCCGCAAAGTACCTCTGATGAGAACAA	760
SEQ ID NO: 43	683	CAGACCTCAAGGAGCATGTCATCAAGCCGGTGTATCCGGGAAGTACCTCTGATGAGAACAA	742
SEQ ID NO: 42	761	CCATCTCCACCTGAACCCGGTCTGGCCGGTTCTGTCATCGGGGCCACGGTGACGCCG	820
SEQ ID NO: 43	743	CCATCTCCACCTGAACCCATCGGGCCGGTTCTGTCATCGGGGCCCTCACGGGATGCCG	802
SEQ ID NO: 42	821	GTCTCACGGGCCAGATCATCATCGAACCTATGGTGGCTGGGAGGCCACGGGGCG	880
SEQ ID NO: 43	803	GTCTCACCGGCCAGATCATCGAACCTATGGTGGCTGGGAGGCCACGGGGCG	862
SEQ ID NO: 42	881	GTGCCTTCTCTGCAAGGACCAACCAAGGTGACCGYAGTGGGCCATGCCAGGC	940
SEQ ID NO: 43	863	GTGCCTTCTCTGGCAAGGACCCATACCAAGGTGACCCGAGTGGGCCATGCCAGGC	922

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FIG. 8 (Continued)

SEQ ID NO:42	941	ARGCGCCAAAGGCCATCATGCCAGGGCTCGCACGCCGCTGCAATTGGAGATCTCAT	1000
SEQ ID NO:43	923	AGGCTGCCAAAGGCCATCATGCCAGGGCTCGCACGCCGCTGCAATTGGAGATCTCAT	982
SEQ ID NO:42	1001	ACGCCATCGGTGCTGAGCTTGTCTGTTGTCGACTCCATGGCACGGCAAGA	1060
SEQ ID NO:43	983	ATGCCATCGGTGACCTGAGCTTGTCTGACTCCATGGCACGGCAAGA	1042
SEQ ID NO:42	1061	TCCCCGACAGGGAGATCCTAAGCTCGTGAAGGAAGACTTGACTTCAGGCCGGATGA	1120
SEQ ID NO:43	1043	TCCCCGACAGGGAGATCCTCAGCTCGTGAAGGAAGACTTGACTTCAGGCCGGATGA	1102
SEQ ID NO:42	1121	TCAGCATCAACCTGGACTTGAGAAAGGTGGAACAGGTTCATCAAGACGGCTGCTTACG	1180
SEQ ID NO:43	1103	TCACGATCAACCTCGACTTGAGAAAGGTGGAACAGGTTCATCAAGACGGCTGCTTACG	1162
SEQ ID NO:42	1181	GTCACTTGGCCGTGATGATGCCGACTTACCTGGAGGTGGAACAGGTTCATCAAGACGGCTGCTTACG	1240
SEQ ID NO:43	1163	GTCACTTGGCCGTGATGATGCCGACTTACCTGGAGGTGGAAGGCCCTCAAGTTCG	1222
SEQ ID NO:42	1241	ACAAGGCATCTGCCATAAGGCATGGCAT	1268
SEQ ID NO:43	1223	ACAAGGCATCTGCCATAAGGCATGGCAT	1250
SEQ ID NO:42	1271	TCTTGGCTGCGCCCTCAAGTTCGTCAGACGGGATCATGTTGCTCCTGGAAAGTGGG	1330
SEQ ID NO:43	1266	TCTTGGCTGATGCCCTCAAGTTCGCAAGGGGGATCCTTTCGCTGGAAAGTAAG	1325
SEQ ID NO:42	1331	AAGAAGCATTAGACATTG	1348
SEQ ID NO:43	1326	AAGAAGCATTCAACATCG	1343